



Skin Color and Self-reported Sun Exposure Scores are Associated with Serum 25-Hydroxyvitamin D Concentrations in a Multi-ethnic Population Living in South Florida

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Authors' Contributions

This study was a result of collaborative work between all authors. Author SA wrote the manuscript, managed the manuscript, performed literature searches, performed statistical analysis and collected data. Author JCE collected the data, contributed in designing of the study and helped developing the methodology and discussion sections. Author GGZ managed the analyses of the study. Authors AN, LS and MML proofread and critically reviewed the manuscript. Author FGH initiated the idea, designed and secured funding, secured data collection and contributed in correction of draft and analysis. All authors read and approved the final manuscript.

Original Research Article

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ABSTRACT

Aims: The aim was to investigate the association between serum 25-hydroxyvitamin D [25(OH)D], skin color and sun exposure score.

Study Design: Cross-sectional.

Place and Duration of Study: Florida International University, Robert Stempel College of Public Health and Social Work, Department of Dietetics and Nutrition, Miami, Florida from July 2012 to October 2012.

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Methodology: Seventy six adults, ages 18-36 years living in South Florida participated in the study. Skin color was quantified by a IMS Smart Probe 400 scanner and 25(OH)D was measured by ELISA. A sun exposure questionnaire was used to record the weekly sun exposure scores. A food frequency questionnaire was used to record daily vitamin D intake.

Results: Multiple-linear regression analysis indicated that sun exposure, forearm skin color and vitamin D intake were significant predictors of 25(OH)D ($P=.004$, $P=.003$ and $P=.021$ respectively). This association held after controlling for covariates ($B=.371$, $P=.027$ for forearm, $B=.031$, $P=.005$ for total sun exposure and $B=.689$, $P=.003$ for vitamin D intake).

Conclusion: Skin color, sun exposure along with vitamin D intake may be used as an indirect non-invasive tools to estimate 25(OH)D levels in healthy individuals in South Florida.

Keywords: Serum vitamin D; vitamin D deficiency; UV exposure; vitamin D intake.

1. INTRODUCTION

Hypovitaminosis D is associated with a wide range of health concerns, such as osteomalacia, diabetes, cardiovascular complications, autoimmune conditions and certain cancers [1-7]. While vitamin D can be obtained through dietary sources [8-9], the vitally important level of bodily vitamin D is provided by its endogenous ultraviolet-dependent (UV) biosynthesis, through photochemical conversion of 7-dehydrocholesterol in the skin [7]. Hepatocytes convert vitamin D to 25-hydroxyvitamin D [25(OH)D] [10]. Circulating 25(OH)D concentration is widely used as the most acceptable indicator of vitamin D status due to its half-life which is indicative of exposure to sun within two months [9,11,12]. Environmental conditions such as geographical location, latitude, seasons and atmosphere along with personal characteristics such as age, gender, race, skin color, sun protective behaviors, dietary intakes and body mass index (BMI) can potentially affect the serum 25(OH)D level [13-26].

Previous studies using sun exposure questionnaires to predict serum vitamin D levels have shown significant results [27-31]. However, the correlations have been in the range of .29-.39, leaving a large percentage of the variation in serum vitamin D unexplained by sun exposure alone. This indicates that other factors such as age and skin color should be included in the model in order to increase the predictive power. Hanwell et al. [32] showed that the correlation between serum vitamin D and sun exposure is only significant for the summer months, but not for winter in hospital workers in Italy, meaning that even when workers spent several hours under the sun in winter it has no effect on serum vitamin D levels and other factors like vitamin D intake maybe more relevant during these months [32]. Therefore, using self-reported sun exposure alone leads to a high probability of misclassification of vitamin D status. Another variation in estimating vitamin D status could be ethnicity. Our previous study showed significant variation in serum vitamin D levels across ethnicity among individuals residing in South Florida [33].

Measuring serum 25(OH)D level requires an invasive and expensive process. Developing validated noninvasive methodology to predict vitamin D levels are needed. Methods that can easily be used in a clinical setting without training could facilitate screening for hypovitaminosis D. Providing a new methodology to clinicians to predict vitamin D deficiencies quickly will lead to prevention of vitamin D deficiency and related complications.

Therefore, the aim of this study was to develop non-invasive and inexpensive methodology to predict serum 25(OH)D among a multi-ethnic population living in a subtropical climate, Miami, Florida.

2. MATERIALS AND METHODS

This study was approved by Institutional Review Board (IRB) and all participants signed an informed consent form.

2.1 Participants

In this cross-sectional study participants were recruited from the Florida International University communities by flyers and word of mouth. The inclusion criteria was to be older than 18 year, not taking vitamin D supplements, living in South Florida for more than one year and not majoring in the nutrition field. A sample of 85 participants aged between 18 and 36 years were enrolled. The study was conducted during the months of July 2012 to October 2012. The average daily temperature for the month of June was 82.6°F with an average rainfall of 12.56 inches in Miami [34]. Similarly, the average daily temperature was 83.4°F with an average rainfall of 8.92 inches and 83.8°F with 15.92 inches for the months of July and August, respectively.

2.2 Skin Color

In order to have an objective measure of sun exposure, skin color was determined by reflectance colorimetry using the IMS Smart Probe 400, Millford, CT, USA. This instrument uses the International Commission on Illumination Scale which ranges from 0 (black) to 100 (white) for skin color. Two readings at each measurement site (6 readings total) for each participant were taken: Two on the wrist of the right hand (area most exposed to sun), two on the inside of the right upper arm and the waist (areas less exposed to the sun). The mean values were used. The change in skin color due to sun exposure was calculated by finding the difference between the less exposed area (natural color) and the most exposed area.

2.3 Blood Collection

Fasted (8-10 hours) venous blood (10ml) was collected from each subject by a certified phlebotomist using standard laboratory methods. Serum vitamin D concentrations were measured with an enzyme-immunoassay kit by absorbance (Immunodiagnostic Systems Scottsdale, AZ, USA).

2.3.1 Demographics questionnaire

A self-administered socio-demographic questionnaire was used to collect data on age, education, race/ethnicity, employment status, health insurance, alcohol use, marital status and smoking status.

2.4 Anthropometrics

The participants were asked to step behind a screened area, remove their shoes and heavy clothing and step on a scale with height rod to measure their body weight and height. The participants' waist and hip circumferences were measured with a non-flexible tape measure.

2.5 Sun Exposure Questionnaire (SEQ)

The instrument developed by Hanwell et al. [32] was applied following the original rubric. Time spent outdoors during the previous week (0<5 minutes, 1=5-30 minutes and 2=>30 min) was self-reported. Four options for skin exposed while outdoors were offered (1=face and hands, 2=face, hands and arms, 3=face, hands and legs and 4=bathing suit). The daily sun exposure score for each day was calculated by multiplying the time spent outdoors score times the skin exposed while outdoors score. The scale for each day ranged from 0 to 8. The weekly sun exposure was calculated by adding the daily scores (min=0, max=56).

2.6 Short Vitamin D and Calcium Questionnaire (FFQ-D)

The short instrument for assessing dietary intakes of calcium and vitamin D by Blalock and colleagues was used [35]. The original instrument included 22 foods and beverages. The new calculator has added pizza for a total of 23 foods. The questionnaire asked for the frequency of consumption of each food and the serving size.

2.7 Statistical Analysis

The descriptive statistics for continues variables was presented as mean $\pm$ SD and proportions for categorical variables. Spearman's correlation among weekly sun exposure scores, delta of skin color, sun exposure scores with serum vitamin D and short food frequency questionnaire with serum vitamin D were run. The relationship between serum 25(OH)D, sun exposure and skin color was evaluated by a multi-linear regression models. Serum 25(OH)D was the dependent variable. Total sun exposure score and forearm skin color were independent variables. The three variables were all continuous. A simple model was run using total sun exposure and forearm skin color as predictors of serum 25(OH)D. Confounding factors including daily vitamin D intake, age, gender, BMI, years living in US, race, tobacco use and alcohol consumption were added to the adjusted model. Two-way and three-way interactions were tested between total sun exposure score, forearm skin color, daily vitamin D intake, and gender. Analyses were conducted using SPSS version 19 (Chicago, IL, US).

3. RESULTS AND DISCUSSION

This cross-sectional study included 85 participants. Nine participants were excluded; 4 subjects due to refusing blood draw and 5 subjects due to missing values for sun exposure questionnaire. Final sample considered for the analysis included 76 participants, males (n=39), females (n=37). Descriptive characteristics are provided in Table 1. The mean age was 25.32 ± 4.82 years. Forty-eight percent were females and the mean BMI was 24.49 ± 4.10 kg/m². The mean serum 25(OH)D, forearm skin color and total sun exposure were 24.53 ± 9.65 , 56.17 ± 7.35 and 28.05 ± 12.19 , respectively. The minimum score for weekly sun exposure was 14 and the maximum 40. About sixty percent of the participants were White and about forty percent were Blacks and Asians. Vitamin D insufficiency (25(OH)D <30 ng/ml) was present in 77.6% of the participants.

Table 1. Characteristics of participants (n = 76)

Variable	Mean $\pm$ SD
Age (years)	25.3 $\pm$ 4.8
Gender	
_Female	48.7%
_Male	51.3%
BMI (kg/m ²)*	24.5 $\pm$ 4.1
Living in US (years)	10.4 $\pm$ 9.3
Vitamin D intake (mcg/day)	7.4 $\pm$ 4.9
Serum 25(OH)D (ng/mL)	24.5 $\pm$ 9.7
Forearm skin color	56.2 $\pm$ 7.6
Sun exposure score	28.1 $\pm$ 12.2
Race	
_White	60.5%
_Black and Asian	39.5%
Tobacco use	
_No	92.1%
_Yes	7.9%
Alcohol consumption	
_No	57.9%
_Yes	42.1%

Data are % or mean $\pm$ standard deviation (SD). *BMI = Body mass index

Correlations between weekly sun exposure and change in skin color, weekly sun exposure and serum vitamin D, and vitamin D intake and serum vitamin D were significant ($P=.037$, $r=.24$ and $P=.05$, $r=.23$ and $P=.03$ and $r=.25$ respectively) (Table 2 and Figs.1-3).

Table 2. Correlation analysis (n = 76)

Variables compared	<i>r</i>	<i>P</i> -value
Sun exposure vs. change in skin color	.240	.04
Sun exposure vs. serum vitamin D	.226	.05
Vitamin D intake vs. serum vitamin D	.246	.03

† $P<.05$ is considered significant

The unadjusted model showed that total sun exposure ($P=.004$), forearm skin color ($P=.003$) and daily vitamin D intake ($P=.021$) were good predictors of serum 25(OH)D. In this model, 22 percent of the variation in serum 25(OH)D was explained by total sun exposure, skin color and daily vitamin D intake. This relationship remained significant ($P=.005$ for sun exposure, $P=.027$ forearm skin color and daily vitamin D intake $P=.003$) after controlling for covariates, including age, gender, BMI, years living in US, race, tobacco use and alcohol consumption. The fully adjusted model explained 34.5 percent of the variation in serum 25(OH)D. For every one unit increase in forearm skin color score, total sun exposure and daily vitamin D intake, there was a .37, .03 and .69 unit increase in 25(OH)D, respectively, keeping the other variables constant (Table 3). Two and three-way interactions between daily vitamin D intake, total sun exposure score, forearm skin color and gender were not significant.

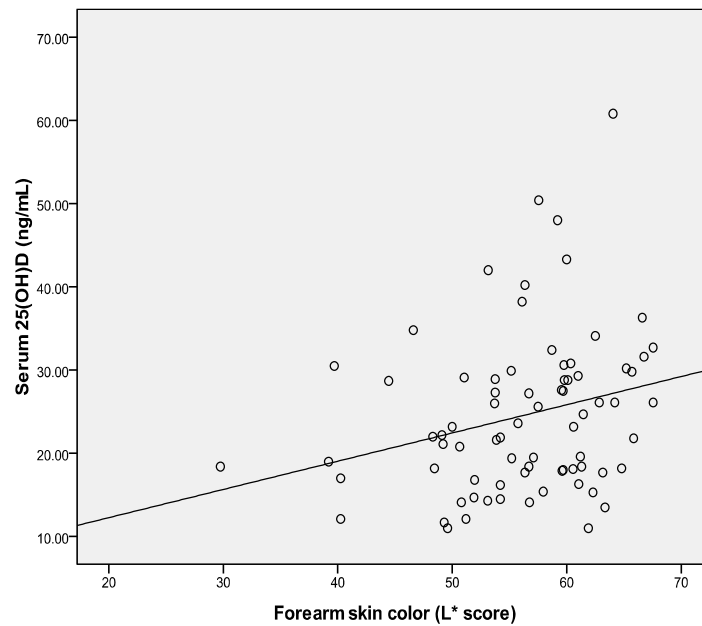


Fig. 1. Correlation between serum 25(OH)D and forearm skin color

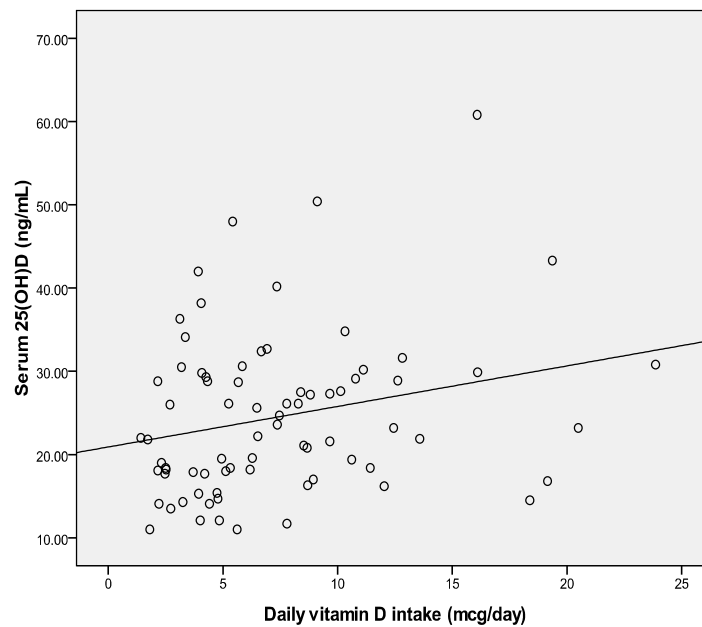


Fig. 2. Correlation between serum 25(OH)D and vitamin D intake

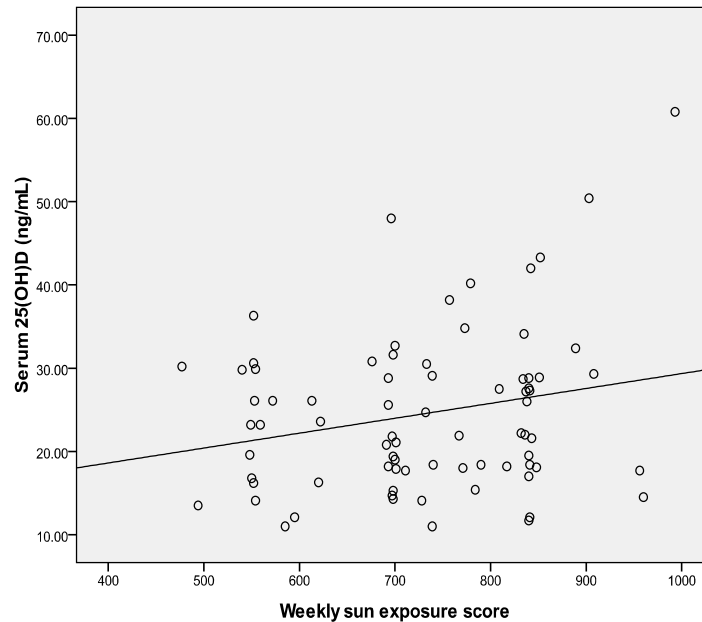


Fig. 3. Correlation between serum 25(OH)D and sun exposure score

Table 3. Relationship of serum 25(OH)D and covariates

Parameter*	β	SE	T	P-value [†]
Forearm skin color	.371	.164	2.258	.027
Weekly sun exposure score	.031	.011	2.898	.005
Daily vitamin D intake	.689	.225	3.059	.003

*Other covariates in the multiple linear regression were age, gender, BMI, years living in US, race, tobacco use and alcohol consumption; [†] $P < .05$ is considered significant

This study examined the association between forearm skin color, sun exposure score and daily vitamin D intake with serum 25(OH)D concentration. The results showed forearm skin color, sun exposure score and daily vitamin D intake predicted serum 25(OH)D among our participants. These findings suggest that people with lighter skin color have higher concentrations of serum 25(OH)D. Our results are consistent with similar studies that reported darker skin color is associated with serum 25(OH)D insufficiencies or deficiencies [36-38]. We also measured the unexposed skin color in two areas and calculated the delta between exposed and un-exposed areas; however, only exposed (forearm) skin color was significantly associated with serum 25(OH)D concentrations. The correlations between delta of exposed and un-exposed areas, sun exposure and serum vitamin D were not significant. This is in contrast to the findings of Nessvi et al. [15] who suggested that serum 25(OH)D is correlated with both constitutive (unexposed) and facultative (exposed) skin color. The reason for this difference may be because their study was done during non-summer seasons, in which people may not have significant exposure to sun. This can result in a non-significant difference between the exposed and unexposed areas skin color. Our study was conducted during summer. Also, the exposed areas in that study were forehead and outer arm, while we considered forearm skin color as our sun exposed area. However, in non-

summer seasons, the outer arm may be covered and it may not reflect the most sun exposed skin area. There are several studies confirming that forearm and inner upper arm skin colors are valid measures of exposed and unexposed areas [17,22,39,40]. Tanning behaviors can also affect the color of unexposed skin area and changes the delta between constitutive and facultative skin colors. Although the ratio between upper arm skin color and forearm skin color had a stronger correlation with serum 25(OH)D in comparison to the ratio between stomach and forearm skin color with serum vitamin D; none of these correlations were statistically significant.

The sun exposure questionnaire used in this study has not been validated for South Florida. Most studies validate sun exposure questionnaires against observed records of sun exposure or dosimetry [41-45]. However, these methods are not considered as a gold standard. Others use serum vitamin D as the gold standard due to its ability to reflect recent sun exposure [32]. In our study, weekly sun exposure scores were significantly correlated with change in skin color due to sun exposure and serum vitamin D. However, the correlations were weak (0.226 and 0.240), leaving a large proportion of variation in serum vitamin D unexplained. The fully adjusted model including sun exposure score, vitamin D intake and forearm skin color as predictors, explained 34.5% of the variation in serum vitamin D meaning that still 65% is not explained.

Other factors like the use of sunscreen, umbrellas, or days outside South Florida were not investigated and may have played a role in serum vitamin D prediction. Previous studies have shown the use of sunscreen with a SPF of 8 completely suppressed the cutaneous synthesis of vitamin D [24,46]. In addition, the vitamin D food frequency questionnaire did not include ethnic foods that may have contributed to the serum vitamin D levels like sardines and mixed ethnic dishes with cheese. The use of multivitamins containing vitamin D and the intake of vitamin D fortified foods, such as orange juice were not measured.

It is important to note that despite the young age of our sample and the majority being Caucasian, over three-quarters of the participants (77.6%) were vitamin D insufficient. It is almost paradoxical that in a tropical area like South Florida and during the summer months, the incident of vitamin D insufficiency is still high. Mean sun exposure scores were in the middle of the scale, reflecting not much outdoor activity. Similarly, the mean vitamin D intake was 7.43 mcg/day. The recommended dietary allowance for vitamin D between the ages of 19-50 years old is 15 mcg/day [47]. Alcohol intake can also interfere with vitamin D absorption and activation in the liver. In our sample, 42.1% of the subjects reported drinking one or more alcohol servings per week (12 oz of beer, 5 oz of wine or 1 oz hard liquor). The low vitamin D intake, limited sun exposure and alcohol consumption may explain in part the high incidence of insufficiency in our sample.

We suggested that higher sun exposure scores resulted in greater concentrations of 25(OH)D. These findings are consistent with Malvy et al. [48] who reported serum 25(OH)D concentration was associated with the results of self-reported sun exposure among 1191 French adults. Kimlin et al. [18] reported that regular winter sunscreen user who are involved in outdoor activities had some of the highest levels of 25(OH)D concentrations among adults living in New Zealand; however, we could not find any significant correlations between sun screen application and variance in 25(OH)D level. This may be due to our smaller group of participants and different climates. Limitations of our study included small sample size, limited age range (18-36 years old), self-reported questionnaires, cross-sectional study design, and lack of access to other regions in South Florida. Since approximately 60% of our participants were Whites and 40% were Blacks and Asians, we did not have enough power

to categorize our participants into ethnic groups and assess the sun exposure behaviors and vitamin D intake behaviors among them. Another limitation of our study was not considering hepatic function. After transportation of vitamin D to the liver by vitamin D binding protein, vitamin D-25-dehydroxylase converts it to 25(OH)D and the function of this enzyme can affect the amount of circulating 25(OH)D [49,50].

4. CONCLUSION

Forearm skin color, sun exposure scores along with daily vitamin D intake were indirect non-invasive tools to estimate serum 25(OH)D level in young healthy individuals living in South Florida. Sun exposure scores were significantly correlated with exposed (forearm) skin color and serum vitamin D which indicates that sun exposure questionnaire is appropriate for this particular sample. Further studies should be conducted among populations with more variation of skin colors and a greater age range to confirm these results.

CONSENT

All participants signed an informed consent form before data collection.

ETHICAL APPROVAL

All authors hereby declare that all experiments have been examined and approved by the Florida International University, Institutional Review Board for the inclusion of human subjects and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Cavalier E, Delanaye P, Chapelle JP, Souberbielle JC. Vitamin D: Current status and perspectives. *Clin Chem Lab Med.* 2009;47:120–127.
2. Munger KL, Levin LI, Hollis BW, Howard NS, Ascherio A. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. *JAMA.* 2006;296:2832–2838.
3. Grandi NC, Breitling LP, Brenner H. Vitamin D and cardiovascular disease: Systematic review and meta-analysis of prospective studies. *Prev Med.* 2010;51:228–233.
4. Lehmann B, Querings K, Reichrath J. Vitamin D and skin: new aspects for dermatology. *Exp Dermatol.* 2004;13(4):11–15.
5. Grau MV, Baron JA, Sandler RS, Haile RW, Beach ML, Church TR, et al. Vitamin D, calcium supplementation, and colorectal adenomas: results of a randomized trial. *J Natl Cancer Inst.* 2003;95(23):1765–1771.
6. Garland CF, Garland FC, Gorham ED, Lipkin M, Newmark H, Mohr SB, et al. The role of vitamin D in cancer prevention. *Am J Public Health.* 2006;96(2):252–261.
7. Holick MF. Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis. *Am J Clin Nutr.* 2004;79(3):362–371.
8. Hollis BW. Circulating 25-hydroxyvitamin D levels indicative of vitamin D sufficiency: Implications for establishing a new effective dietary intake recommendation for vitamin D. *J Nutr.* 2005;135(2):317–322.

9. Vieth R. The pharmacology of vitamin D, including fortification strategies. In: Feldman D, Pike JW, Glorieux FH, editors. Vitamin D. 2nd ed. New York: Elsevier. 2005;2:995–1015.
10. Holick MF. Vitamin D: Extraskeletal health. *Endocrinol Metab Clin North Am.* 2010;39:381–400.
11. Standing Committee on the Scientific Evaluation of Dietary Reference Intakes. Dietary reference intakes: Calcium, phosphorus, magnesium, vitamin D, fluoride. National Academy Press; 1997.
12. Hollis BW, Wagner CL, Drezner MK, Binkley NC. Circulating vitamin D3 and 25-hydroxyvitamin D in humans: An important tool to define adequate nutritional vitamin D status. *J Steroid Biochem Mol Biol.* 2007;103(3-5):631-634.
13. Salih FM. Effect of clothing varieties on solar photosynthesis of previtamin D3: An in vitro study. *Photodermatol Photoimmunol Photomed.* 2004;20(1):53-58.
14. Nessvi S, Johansson L, Jopson J, Stewart A, Reeder A, McKenzie R, et al. Association of 25-hydroxyvitamin D3) levels in adult New Zealanders with ethnicity, skin color and self-reported skin sensitivity to sun exposure. *Photochem Photobiol.* 2011;87(5):1173-1178.
15. Park S, Johnson MA. Living in low-latitude regions in the United States does not prevent poor vitamin D status. *Nutr Rev.* 2005;63:203-209.
16. Janda M, Kimlin M, Whiteman D, Aitken J, Neale R. Sun protection and low levels of vitamin D: Are people concerned? *Cancer Causes Control.* 2007;18(9):1015-1019.
17. Rockell JE, Skeaff CM, Williams SM, Green TJ. Association between quantitative measures of skin color and plasma 25-hydroxyvitamin D. *Osteoporos Int.* 2008;19(11):1639-1642.
18. Kimlin M, Harrison S, Nowak M, Moore M, Brodie A, Lang C. Does a high UV environment ensure adequate vitamin D status? *J Photochem Photobiol B.* 2007;14;89(2-3):139-147.
19. Webb AR, Kline L, Holick MF. Influence of season and latitude on cutaneous synthesis of vitamin D3: Exposure to winter sunlight in Boston and Edminton will not promote vitamin D3 synthesis in human skin. *J Clin Endocrinol.* 1988;68:373–378.
20. Ladizesky M, Lu Z, Oliveri B, San Roman N, Diaz S, Holick MF, et al. Solar ultraviolet B radiation and photoproduction of vitamin D3 in central and southern areas of Argentina. *J Bone Mineral Res.* 1995;10:545–549.
21. Matsuoka LY, Wortsman J, Chen TC, Holick MF. Compensation for the intraracial variance in cutaneous synthesis of vitamin D. *J Lab Clin Med.* 1995;126:452–457.
22. Matsuoka LY, Wortsman J, Haddad JG, Kolm P, Hollis BW. Racial pigmentation and the cutaneous synthesis of vitamin D. *Arch Dermatol.* 1991;127:536–538.
23. Matsuoka LY, Wortsman J, Hollis BW. Use of topical sunscreen for the evaluation of regional photosynthesis of Vitamin D3. *J Am Acad Dermatol.* 1990;22:772–775.
24. Matsuoka LY, Wortsman J, Hollis BW. Suntanning and cutaneous synthesis of vitamin D3. *J Lab Clin Med.* 1990;116:87–90.
25. MacLaughlin J, Holick MF. Aging decreases the capacity of human skin to produce Vitamin D3. *J Clin Invest.* 1985;76:1536–1538.
26. Holick MF, Matsuoka LY, Wortsman J. Age, vitamin D, and solar ultraviolet. *Lancet.* 1989;4:1104–1105.
27. Van der Maei IAF, Blizzard L, Ponsonby A-L, Dwyer T. Validity and reliability of adult recall of past sun exposure in a case-control study of multiple sclerosis. *Cancer Epidemiol Biomarkers Prev.* 2006;15:1538–1544.

28. Brot C, Vestergaard P, Kolthoff N, Gram J, Hermann AP, Sørensen OH. Vitamin D status and its adequacy in healthy Danish perimenopausal women: Relationships to dietary intake, sun exposure and serum parathyroid hormone. *Br J Nutr*. 2001;86:S97–103.
29. Pasco JA, Henry MJ, Nicholson GC, Sanders KM, Kotowicz MA. Vitamin D status of women in the Geelong Osteoporosis Study: Association with diet and casual exposure to sunlight. *Med J*. 2001;175:401–405.
30. Sullivan SS, Cobb JL, Rosen CJ, Holick MF, Chen TC, Kimlin Michael G, Parisi Alfio. Assessment of sun exposure in adolescent girls using activity diaries. *Nutr Res*. 2003;23:631–644.
31. McCarty CA. Sunlight exposure assessment: can we accurately assess vitamin D exposure from sunlight questionnaires? *Am J Clin Nutr*. 2008;87(4):1097S-101S.
32. Hanwell HE, Vieth R, Cole DE, Scillitani A, Modoni S, Frusciante V, et al. Sun exposure questionnaire predicts circulating 25-hydroxyvitamin D concentrations in Caucasian hospital workers in southern Italy. *J Steroid Biochem Mol Biol*. 2010;121(1-2):334-337.
33. Shaban LH, Zarini GG, Exebio JC, Sukhram SD, Huffman FG. Serum vitamin D insufficiency and diabetes status in three ethnic minority groups. *J Immigr Minor Health*. 2012;14(6):926-932.
34. Griffin M, Zierden DF. Climate Summary for Florida. 2012. Available at: <http://climatecenter.fsu.edu/products-services/summaries/climate-summary-for-florida-june-2012>.
35. Blalock SJ, Norton LL, Patel RA, Cabral K, Thomas CL. Development and assessment of a short instrument for assessing dietary intakes of calcium and vitamin D. *J Am Pharm Assoc*. 2003;43(6):685-693.
36. Armas LA, Dowell S, Akhter M, Duthuluru S, Huerter C, Hollis BW, et al. Ultraviolet-B radiation increases serum 25-hydroxyvitamin D levels: The effect of UVB dose and skin color. *J Am Acad Dermatol*. 2007;57(4):588-593.
37. Harris SS, Dawson-Hughes B. Seasonal changes in plasma 25-hydroxyvitamin D concentrations of young American black and white women. *Am J Clin Nutr*. 1998;67:1232–1236.
38. Nesby-O'Dell S, Scanlon KS, Cogswell ME, Gillespie C, Hollis BW, Looker AC, Allen C, Dougherty C, Gunter EW, Bowman BA. Hypovitaminosis D prevalence and determinants among African American and white women of reproductive age: Third national health and nutrition examination survey, 1988-1994. *Am J Clin Nutr*. 2002;76(1):187–192.
39. Pierard GE. EEMCO guidance for the assessment of skin colour. *J Eur Acad Dermatol Venereol*. 1998;10:1–11.
40. Shriver MD, Parra EJ. Comparison of narrow-band reflectance spectroscopy and tristimulus colorimetry for measurements of skin and hair color in persons of different biological ancestry. *Am J Phys Anthropol*. 2000;112:17–27.
41. Diffey BL, Saunders PJ. Behavior outdoors and its effect on personal ultraviolet exposure rate measured using an ambulatory data logging dosimeter. *Photochem Photobiol*. 1995;6:615–618.
42. Dwyer T, Blizzard L, Gies PH, Ashbolt R, Roy C. Assessment of habitual sun exposure in adolescents via questionnaire—a comparison with objective measurement using polysulphone badges. *Melanoma Res*. 1996;6:231–239.
43. O'Riordan DL, Stanton WR, Eyeson-Annan M, Gies P, Roy C. Correlations between reported and measured ultraviolet radiation exposure of mothers and young children. *Photochem Photobiol*. 2000;71:60–64.

44. Thieden E, Ågren MS, Wulf HC. Solar UVR exposures of indoor workers in a working and holiday period assessed by personal dosimeters and sun exposure diaries. *Photodermatol Photoimmunol Photomed*. 2001;17:249–255.
45. Theiden E, Philipsen PA, Heydenreich J, Wulf HC. UV radiation exposure related to age, sex, occupation, and sun behavior based on time-stamped personal dosimeter readings. *Arch Dermatol*. 2004;140:197–203.
46. Matsuoka LY, Ide L, Wortsman J, MacLaughlin JA, Holick MF. Sunscreens suppress cutaneous vitamin D3 synthesis. *J Clin Endocrinol Metab*. 1987;64:1165–1168.
47. National Institutes of Health. Vitamin D fact sheet for health professionals 2011. Available at: <http://ods.od.nih.gov/factsheets/VitaminD-HealthProfessional/>.
48. Malvy DJ, Guinot C, Preziosi P, Galan P, Chapuy MC, Maamer M, et al. Relationship between vitamin D status and skin phototype in general adult population. *Photochem Photobiol*. 2000;71(4):466–469.
49. Holick MF. Vitamin D deficiency. *N Engl J Med*. 2007;357:266–281.
50. Holick MF. Vitamin D and health: Evolution, biologic functions, and recommended dietary intakes for vitamin D. *Clin Rev Bone Miner Metab*. 2009;7:2–19.

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