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# Applying stable isotopes to examine food-web structure: an overview of analytical tools

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## **Applying stable isotopes to examine food-web structure: an overview of analytical tools**

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## 19 ABSTRACT

20 Stable isotope analysis has emerged as one of the primary means for examining the structure and  
 21 dynamics of food webs, and numerous analytical approaches are now commonly used in the  
 22 field. Techniques range from simple, qualitative inferences based on the isotopic niche, to  
 23 Bayesian mixing models that can be used to characterize food-web structure at multiple  
 24 hierarchical levels. We provide a comprehensive review of these techniques, and thus a single  
 25 reference source to help identify the most useful approaches to apply to a given data set. We  
 26 structure the review around four general questions: (1) what is the trophic position of an  
 27 organism in a food web; (2) which resource pools support consumers; (3) what additional  
 28 information does relative position of consumers in isotopic space reveal about food-web  
 29 structure; and (4) what is the degree of trophic variability at the intrapopulation level? For each  
 30 general question, we detail different approaches that have been applied, discussing the strengths  
 31 and weaknesses of each. We conclude with a set of suggestions that transcend individual  
 32 analytical approaches, and provide guidance for future applications in the field.

33

34 *Key words:* Bayesian statistics, dietary variation, individual specialization, mixing model,  
 35 predator-prey interactions, trophic structure.

36

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## 62 I. INTRODUCTION

63           Stable isotope analysis has emerged as one of the primary means to analyze the structure  
64 of food webs. Stable isotopes are especially useful because they provide time- and space-  
65 integrated insights into trophic relationships among organisms, and thus can be used to develop  
66 models of trophic structure. Many of the first applications of stable isotope data in a food-web  
67 context were critical advances, although largely qualitative, providing for broad inferences based  
68 on relative isotope values of consumers and resources (Haines & Montague, 1979; Peterson,  
69 Howarth & Garritt, 1985; Zieman, Macko & Mills, 1984). Over the past 10 years, a series of  
70 more quantitative approaches for analyzing stable isotope data has emerged. These approaches  
71 have dramatically improved our understanding of food webs, for example, providing new insight  
72 into food-chain length (Post, Pace & Hairston, 2000), niche variation (Martinez del Rio *et al.*,  
73 2009a; Moore & Semmens, 2008; Semmens *et al.*, 2009b; Votier *et al.*, 2010), and human-driven  
74 shifts in community structure (Layman *et al.*, 2007b; Schmidt *et al.*, 2007).

75           The emergence of new analytical approaches has led to some debate about which  
76 method(s) is most appropriate to apply to stable isotope data (Hoeinghaus & Zeug, 2008;  
77 Jackson *et al.*, 2009; Layman & Post, 2008; Newsome *et al.*, 2007; Semmens, Moore & Ward,  
78 2009a). At times, this discussion has focused on which analytical approach is “right” or  
79 “wrong”. But a more useful perspective is recognizing the specific types of questions for which  
80 different approaches are best suited. Analogously, use of stomach contents to evaluate dietary  
81 breadth has some very well-understood limitations (Votier *et al.*, 2003), but still provides critical  
82 insight into feeding relationships. Likewise, each stable isotope analytical approach has distinct  
83 strengths and weaknesses (Table 1), and each is more or less appropriate under specific  
84 circumstances. Information regarding these strengths and weaknesses is scattered among dozens  
85 of papers in the field, often rendering direct comparison among techniques difficult. Herein we  
86 provide a comprehensive review of these diverse approaches, structured around four core  
87 ecological questions: (1) what is the trophic position of an organism in a food web; (2) which  
88 resource pools support consumers; (3) what additional information does relative position of  
89 consumers in isotopic space reveal about food web structure; (4) what is the degree of trophic  
90 variability at an intrapopulation level?

91 This review is not intended to be a comprehensive catalogue of every food-web study that has  
92 employed stable isotopes, an endeavour which would be a monumental task given the rapid  
93 proliferation of such studies (Fig. 1). Instead, we emphasize those papers that are paradigmatic  
94 with respect to a particular analytical approach, as well as some of the most recent contributions  
95 to the field. Not expanded upon in this review are the many additional types of information that  
96 are necessarily relevant in interpreting isotope data sets (e.g. trophic discrimination factors,  
97 isotopic routing, tissue turnover rates, lipid extraction, etc.), as other reviews have discussed

98 these topics thoroughly (e.g. Bearhop *et al.*, 2004; Boecklen *et al.*, in press; Martinez del Rio *et*  
99 *al.*, 2009b; McCutchan *et al.*, 2003; Oppel & Powell, 2011; Phillips & Eldridge, 2006; Post *et*  
100 *al.*, 2007; Vanderklift & Ponsard, 2003). Our goal is to provide a single source that outlines  
101 analytical approaches currently being applied to answer questions about food-web structure, and  
102 provide guidelines as to which approaches are most appropriate with respect to a particular data  
103 set or question of interest.

## 104 **II. STABLE ISOTOPE RATIOS AND FOOD WEBS**

105         The two elements most commonly employed in a food-web context are nitrogen (N) and  
106 carbon (C), although sulphur (S), oxygen (O) and deuterium (D) are also useful in particular  
107 cases. The ratio of  $^{15}\text{N}$  to  $^{14}\text{N}$  (expressed relative to a standard,  $\delta^{15}\text{N}$ ) exhibits stepwise  
108 enrichment with trophic transfers, and is a powerful tool for estimating trophic position of  
109 organisms (Minagawa & Wada, 1984; Peterson & Fry, 1987; Post, 2002b). Ratios of carbon  
110 isotopes ( $\delta^{13}\text{C}$ ) vary substantially among primary producers with different photosynthetic  
111 pathways (e.g. C3 *versus* C4 photosynthetic pathways in plants), but change little with trophic  
112 transfers (DeNiro & Epstein, 1981; Inger & Bearhop, 2008; Peterson & Fry, 1987; Post, 2002b).  
113 Therefore,  $\delta^{13}\text{C}$  can be used to determine original sources of dietary carbon. Similarly, the ratio  
114 of sulphur isotopes ( $\delta^{34}\text{S}$ ) varies substantially among primary producers, but changes relatively  
115 little with progression through a food web, and also can be used to identify important resource  
116 pools. This has proven especially insightful in marine systems where the sulphur cycle often  
117 gives rise to distinct benthic and pelagic  $\delta^{34}\text{S}$  values (Currin, Newell & Paerl, 1995; Peterson &  
118 Howarth, 1987) and along marine ecotones to differentiate marine and fresh-water (or terrestrial)  
119 sources (Jones *et al.*, 2010). The  $\delta^{18}\text{O}$  and  $\delta^2\text{H}$  values of precipitation vary at multiple spatial  
120 scales, yielding insight into large-scale dietary patterns across geographic regions (Bowen &

121 Revenaugh, 2003), or across smaller-scale environmental gradients (Deines, Wooller & Grey,  
122 2009; Finlay, Doucett & McNeely, 2010; Solomon *et al.*, 2011, 2009). Newsome *et al.* (2007)  
123 and Oulhote *et al.* (2011) provide additional information regarding the insights that can be  
124 gleaned from various isotope tracers.

125         Most frequently,  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  (or one of these in combination with other elemental  
126 tracers) are plotted in bivariate fashion, a depiction that has been variously referred to as niche  
127 space, trophic space, isotope space, or the isotopic niche. Herein, we adopt the term “isotopic  
128 niche”. In this sense, we view the ecological information contained in stable isotope plots as a  
129 proxy for a subset of the Hutchinsonian  $n$ -dimensional hypervolume (Hutchinson, 1957). We  
130 emphasize that the isotopic niche is distinct from, but in many circumstances should align  
131 closely with, aspects of the actual trophic niche (e.g. particular resource pools utilized or relative  
132 trophic position within a web).

133         Examining food-web structure involves analyzing and comparing the relative position of  
134 species, populations, or individuals within this niche space, i.e. concomitantly examining the  
135 relative positions along one (or more) isotopic axes. These data provide for inference regarding  
136 feeding relationships and food-web structure, but they are not direct characterizations of diet  
137 such as those provided by stomach-content analysis, feeding observations, or fecal analysis.  
138 Because of the indirect nature of the data, there are various sources of potential ambiguity in  
139 interpretation of isotope values that relate to all of the analytical approaches discussed herein.

140         Stable isotope values are a product not only of the actual trophic interactions, but are also  
141 driven by myriad underlying biological and chemical processes. For example, when isotopic  
142 routing occurs, i.e. when elemental isotopes from resources are broken down and assimilated  
143 differently among consumer tissue types, direct interpretation of the underlying trophic



144 relationships may be more problematic (Martinez del Rio *et al.*, 2009b). In such a case, a  
145 consumer tissue does not reflect isotopic composition of the bulk diet, but rather the isotopic  
146 composition of the nutrient component of the diet from which the tissue was synthesized. This is  
147 especially important to consider when an individual consumes diverse resources (for example,  
148 feeding on both plants and animals, Kelly & Martinez del Rio, 2010; Martinez del Rio *et al.*,  
149 2009b; Voigt *et al.*, 2008). Ignoring such biochemical processes driving variation in stable  
150 isotope values can result in biased interpretations of trophic interactions.

151         Emergent ecological factors also render  $\delta$  values difficult to interpret in some  
152 circumstances. First, isotopic similarity does not necessarily mean ecological similarity, as two  
153 individuals may have the same isotopic niche, but distinct ecological niches. That is, even though  
154 trophic pathways that may support the two individuals are distinct, the different source pools are  
155 characterized by similar stable isotope values. Second, if different potential resource pools have  
156 overlapping  $\delta$  values, stable isotopes alone may not be able to pinpoint the particular source pool  
157 being utilized. Different isotope values of source pools typically is essential for isotopes to be a  
158 useful analytical tool. Third, when using stable isotopes to reconstruct dietary relationships, both  
159 source and consumer pools must be sampled on spatial and temporal scales that reflect the  
160 relative incorporation rates of the elements and the turnover rates of tissues (Martinez del Rio *et*  
161 *al.*, 2009b; Post, 2002b). Yet this final point also underpins the strength of isotopes relative to  
162 direct dietary information: when sampled at appropriate scales, stable isotopes provide for time-  
163 and space-integrated representations of trophic relationships in food webs. Such data provide  
164 important insights into food-web structure not possible through snapshot characterizations of  
165 diet.

166 All of the analytical approaches discussed herein deal with either raw  $\delta$  values, or values  
167 that are transformed to represent a specific ecological variable (e.g. trophic position or dietary  
168 proportions from different source pools) (Newsome *et al.*, 2007). Analysis of raw  $\delta$  values  
169 allows inferences regarding feeding relationships, but can be especially sensitive to the relative  $\delta$   
170 values of source pools. For example, broad dispersion among consumers in a  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$   
171 biplot would seem to imply diverse resource use among individuals, but this pattern may also be  
172 a result of high variance in isotope values of source pools. To this end,  $\delta$  space can be  
173 transformed to “proportional” space using isotope values of known source pools (Newsome *et*  
174 *al.*, 2007). Similarly, raw  $\delta^{15}\text{N}$  values can be converted to direct trophic position estimates using  
175 assumed values of  $\delta^{15}\text{N}$  discrimination with trophic transfers, as well as adequate  
176 characterization of isotopic baselines (Post, 2002*b*). Such transformations are often preferred  
177 because they are more ecologically meaningful than raw  $\delta$  values. For example, transforming  
178  $\delta^{15}\text{N}$  values into trophic positions converts them into an actual characteristic of the organism.  
179 But such transformations require considerable additional *a priori* information, including  
180 temporally and spatially appropriate estimates of isotopic baselines and end members, as well as  
181 trophic discrimination factors. If this information is not available or of poor quality, the  
182 transformations may not accurately describe aspects of trophic structure.

183

### 184 **III. INITIAL APPLICATIONS OF STABLE ISOTOPES IN A FOOD-WEB CONTEXT**

185 In a food-web context, many of the first applications of stable isotope data were largely  
186 qualitative, i.e. making general inferences from the relative isotopic values of consumers and/or  
187 resources (Fry, Joern & Parker, 1978; Haines & Montague, 1979; Peterson *et al.*, 1985; Zieman

188 *et al.*, 1984). For example, Peterson *et al.* (1985) suggested the fundamental importance of  
189 *Spartina alterniflora* grass for marsh consumers by qualitatively comparing  $\delta^{34}\text{S}$  and  $\delta^{13}\text{C}$  values  
190 in producer and consumer tissue. Haines & Montague (1979) took a similar approach, using the  
191 variation in  $\delta^{13}\text{C}$  among estuarine primary producers qualitatively to infer the most important  
192 sources for various estuarine consumer species. Hobson & Welch (1992) provided one of the  
193 first insights into the general structure of Arctic food webs using isotope values. All of these  
194 initial advances were critical to laying the foundation for the myriad stable isotope research  
195 programs that are now a fundamental part of the ecological sciences.

196         A logical progression from these early contributions was to apply basic statistics [e.g. *t*-  
197 tests, analysis of variance (ANOVA), multivariate models, etc.] to compare mean  $\delta^{13}\text{C}$  and/or  
198  $\delta^{15}\text{N}$  values among groups, sites or time periods (Oulhote *et al.*, 2011). Some type of basic  
199 statistical comparison can be found in almost any current stable isotope paper in the field.  
200 Simple statistics provide the basic framework for interpreting isotope data, but can be limited as  
201 to the depth of ecological insight that they can provide. Often, basic statistics are used in  
202 conjunction with various other approaches outlined herein. Basic statistical approaches  
203 obviously were not developed for isotope data *per se*, so we focus the remainder of this review  
204 on analytical approaches that are targeted for isotope data sets specifically.

205

#### 206 **IV. WHAT IS THE TROPHIC POSITION OF AN ORGANISM IN A FOOD WEB?**

##### 207 **(1) Species-specific baselines**

208         One of the most important initial advances beyond basic statistics came from the  
209 realization that  $\delta^{15}\text{N}$ , because of the discrimination that occurs with trophic transfers, could be

210 used as a proxy for trophic position (DeNiro & Epstein, 1981; Minagawa & Wada, 1984). In this  
211 context,  $\delta^{15}\text{N}$  provided for a continuous measure of trophic position, a notable difference from  
212 simply assigning organisms to discrete trophic levels based on natural-history observations.

213 Although early studies used the untransformed  $\delta^{15}\text{N}$  values as a measure of trophic position, later  
214 work recognized that  $\delta^{15}\text{N}$  is influenced by local biogeochemistry (baseline variation), trophic  
215 discrimination and the trophic position of an organism. Researchers have taken two approaches  
216 to address baseline variation: (1) using species-specific baselines to estimate relative trophic  
217 shifts and (2) using long-lived organisms or time-series baselines to estimate trophic position of  
218 higher order consumers.

219         Kling, Fry & Obrien (1992) and Post (2003) both used species-specific baselines to  
220 estimate relative differences in trophic position. Kling *et al.* (1992) used herbivorous copepods  
221 as a baseline to estimate the degree of trophic omnivory in copepods, and Post (2003) used  
222 largemouth bass (*Micropterus salmoides*) that had not transitioned to piscivory to estimate the  
223 degree of cannibalism in young-of-the-year individuals. The use of an ecologically relevant  
224 baseline in both of these examples minimized problems related to spatial and temporal  
225 differences between the baseline (herbivorous copepods and non-piscivorous bass) and the target  
226 organism (omnivorous copepods and cannibalistic bass). This baseline method works well for  
227 questions that do not require absolute estimates of trophic position and when the trophic position  
228 of the baseline organism is well understood (i.e. herbivorous copepods). It does not provide an  
229 absolute estimate of trophic position and is, therefore, limited to questions specific to individuals  
230 or a single species.

231 **(2) Long-lived consumers as baselines**

232           The second approach involves a more general baseline that allows for absolute estimates  
233 of trophic position, thereby facilitating comparisons among species and across ecosystems  
234 (Hobson, Piatt & Pitocchelli, 1994). Cabana & Rasmussen (1996) first suggested that long-lived  
235 primary consumers in temperate lakes (e.g. mussels) may be used to create an isotope baseline  
236 for fish. This was expanded to include multiple sources by Vander Zanden & Rasmussen (1999)  
237 and Post (2002*b*). Vander Zanden & Rasmussen (1999) proposed creating a baseline by fitting a  
238 logistic curve to the isotope values of primary consumers in the  $\delta^{13}\text{C}$ - $\delta^{15}\text{N}$  bi-plot and using this  
239 baseline to calculate the trophic position of higher order consumers. Post (2002*b*) developed a  
240 more general solution by using a two-end member mixing model to create a baseline from which  
241 trophic position could be calculated (see Section V for detailed discussion of mixing models).  
242 Because the isotope estimates of trophic position calculated using these methods can be  
243 compared directly across diverse, complex food webs, this method has been widely adopted for  
244 calculating food-chain length, the number of transfers of energy from the base to the apex of a  
245 food web (Post, 2002*a*). The isotope method has allowed researchers to make considerable  
246 progress in addressing fundamental questions about variation in and environmental controls of  
247 food-chain length in lakes and ponds (Doi *et al.*, 2009; Post *et al.*, 2000), streams (McHugh,  
248 McIntosh & Jellyman, 2010; Sabo *et al.*, 2010; Walters & Post, 2008) and islands (Takimoto,  
249 Spiller & Post, 2008).

250           Trophic position estimates are perhaps the most widely reported metric in food-web  
251 studies employing stable isotopes. But these measures are characterized by fundamental  
252 limitations that are often not appreciated when trophic positions of individuals are calculated.  
253 First, trophic position calculation is dependent on establishing an adequate baseline. In some  
254 fresh-water ecosystems, basal resources are relatively easily isolated at a coarse level (e.g. seston

255 and benthic microalgae in northern U.S. lakes; Post, 2002b), or can be aggregated into  
256 ecologically meaningful categories (e.g. autochthonous *versus* allochthonous pools in rivers;  
257 Layman *et al.*, 2005b). But as food webs become more complex, and the number of potential  
258 basal resource pools increases, establishing an adequate baseline becomes more problematic. In  
259 systems with resource pools that have numerous and variable  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values, it may be  
260 extremely difficult to establish an accurate baseline using just a few isotopes, rendering any  
261 trophic position estimates problematic (Layman, 2007). Second, and equally important, is  
262 identifying  $\delta^{15}\text{N}$  discrimination values for each trophic transfer (Martinez del Rio *et al.*, 2009b).  
263 Discrimination provides the stepwise correction that allows one to convert baseline isotope  
264 values into a trophic position for a consumer. Typically, this value is chosen based on available  
265 meta-analyses (Caut, Angulo & Courchamp, 2009; McCutchan *et al.*, 2003; Post, 2002b;  
266 Vanderklift & Ponsard, 2003), but numerous physiological and environmental factors can affect  
267 discrimination in  $\delta^{15}\text{N}$  (Martinez del Rio *et al.*, 2009b). Values from the meta-analyses are valid  
268 approximations when averaged over a large number of trophic pathways, as is done for  
269 estimating food-chain length (Post, 2002a). But when used for estimating the trophic position of  
270 individuals or single species, literature values can prove misleading, and should be used with  
271 caution, until the causes of variation in trophic discrimination are better understood (Martinez del  
272 Rio *et al.*, 2009b). Until recently, few studies propagated such error in assumed values in  
273 calculations (but see Vander Zanden & Rasmussen, 2001). Resampling from distributions of  
274 baselines and trophic discrimination factors to produce ranges of estimates for trophic position,  
275 with an associated error term, is now more commonly employed (including the frequently used  
276 Bayesian models, see Section V.3). Such estimates are more accurate depictions of possible  
277 solutions that account for potential variation in discrimination factors (Jackson *et al.*, 2011).

278

## 279 V. WHICH RESOURCE POOLS SUPPORT CONSUMERS?

280 Stable isotope analysis can reveal dietary patterns by suggesting specific resources used  
281 by a consumer. In simple systems, where consumers only use two food resources, basic  
282 qualitative comparisons can be made using a single elemental tracer. For example, many of the  
283 first studies that applied stable isotope analysis in a food-web context capitalized on  
284 differentiation in carbon isotope ratios in various basal carbon resource pools (e.g. C3 *versus* C4  
285 plants) to identify sources of primary productivity (Fry *et al.*, 1978; Zieman *et al.*, 1984). In  
286 some cases, traditional multivariate analyses (e.g. canonical discriminant analysis or non-metric  
287 multidimensional scaling) using  $\delta^{15}\text{N}$ ,  $\delta^{13}\text{C}$ , and/or  $\delta^{34}\text{S}$  may be sufficient to suggest source  
288 contributions (e.g. Litvin & Weinstein, 2004). But as the number of potential resources  
289 increases, the ability accurately to identify dietary contributions becomes more problematic.  
290 Over the last two decades, a number of isotope mixing models have been proposed to identify  
291 the relative contributions of various food resources to a consumer's diet.

292

### 293 (1) Geometric approaches

294 Early mixing models used geometric methods to estimate the proportional contribution of  
295 three or more food resources to a consumer's diet using  $\delta$  values (BenDavid, Flynn & Schell,  
296 1997; Kline *et al.*, 1993; Peterson & Howarth, 1987; Whitley & Rabeni, 1997). Euclidean  
297 distances between consumer and sources were calculated in isotopic niche space, and an inverse  
298 relationship was assumed between these distances and the relative contribution of each source to  
299 the consumer's diet. Although this method provides a visually appealing graphical representation

300 of dietary contribution and is a useful heuristic tool (BenDavid *et al.*, 1997; Kline *et al.*, 1993;  
 301 Peterson & Howarth, 1987; Whitley & Rabeni, 1997), Phillips (2001) demonstrated that the  
 302 equations used in these approaches failed accurately to identify dietary contributions. Euclidean  
 303 methods underestimate commonly used food sources and overestimate rare food sources, and the  
 304 equations provide inaccurate estimates when a consumed resource is excluded from the analysis.  
 305 These Euclidean-based approaches have largely been supplanted by other mixing-model  
 306 approaches (Phillips, 2001), but are still employed in isolated cases (e.g. Wengeler, Kelt &  
 307 Johnson, 2010).

## 308 (2) Linear mixing models

309 Phillips (2001) suggested that partitioning of resources could most accurately be  
 310 identified using a basic set of algebraic mass-balance equations (linear mixing model), and this  
 311 has become a fundamental framework for understanding stable isotope data in a food-web  
 312 context. A linear mixing model can determine the relative contribution of  $p$  unique food  
 313 resources from the isotope ratios of  $q$  elemental tracers when  $p \leq q + 1$  (i.e. the number of  
 314 sources cannot exceed the number of elemental tracers by more than one) by solving a series of  
 315 equations (Phillips, 2001). For example, in a simple system with only three possible food  
 316 resources and two isotope tracers, solving a set of three linear mass-balance equations,  
 317 containing three unknowns, will determine the exact proportional contribution of each resource.  
 318 Assuming  $^{15}\text{N}$  and  $^{13}\text{C}$  are the two isotopes, the equations would be represented:

$$319 \quad \delta^{13}\text{C}_T = f_A \delta^{13}\text{C}_A + f_B \delta^{13}\text{C}_B + f_C \delta^{13}\text{C}_C, \quad (1)$$

$$320 \quad \delta^{15}\text{N}_T = f_A \delta^{15}\text{N}_A + f_B \delta^{15}\text{N}_B + f_C \delta^{15}\text{N}_C, \quad (2)$$

$$321 \quad f_A + f_B + f_C = 1, \quad (3)$$



322 where  $\delta_T$  is the isotopic composition of a consumer's tissue and  $f_A$ ,  $f_B$ , and  $f_C$  are fractional  
323 contributions of sources A, B, and C. Although the linear mixing model and mass-balance  
324 equations had been previously used in palaeo-diet research (Schwarcz, 1991), Phillips (2001)  
325 was the first to promote their use in present-day diet studies. These linear mixing models have  
326 since been extended to account for uncertainty in source partitioning (Isoerror: Phillips &  
327 Gregg, 2001) and concentration dependence (Isoconc: Phillips & Koch, 2002). Nearly all of the  
328 more advanced models outlined below have their foundation in the same basic set of algebraic  
329 equations.

330         Most food webs are too complex to use simple linear mixing models because the number  
331 of source pools exceeds the number of useful isotope tracers by more than one. When this is the  
332 case, we move from a mathematically determined system to a mathematically undetermined  
333 system. The latter implies that there are multiple feasible solutions for combinations of source  
334 contributions. To this end, Phillips & Gregg (2003) developed the model IsoSource, which has  
335 become one of the most common analytical tools in the field. The model does not generate exact  
336 values for proportional contributions of each source, but instead provides a range of possible  
337 contributions or feasible solutions. The model examines every possible combination of source  
338 proportions (summing to 100%) incrementally (typically in increments of 1%), then calculates  
339 the predicted isotope value for each combination using linear mass-balance equations. These  
340 predicted values are then examined to determine which ones fall within some tolerance range  
341 (typically 0.1‰) of the observed consumer isotope value, and all of these feasible solutions are  
342 recorded. One of the main advantages of this model, besides its public availability and ease of  
343 use, is the relatively limited amount of input data required (average isotope values of the  
344 consumer and potential sources). Additionally, the model can be adjusted further to consider

345 source pooling (Phillips, Newsome & Gregg, 2005). As with all mixing models, a series of  
346 critical assumptions must be made, and these will be discussed in detail below. But perhaps the  
347 most common problem with studies employing IsoSource is not related to the intrinsic structure  
348 of the model, but instead, to interpretation of its output. That is, researchers often interpret some  
349 measure of central tendency (e.g. the median or mode) as the definitive solution, a conclusion  
350 which is clearly not justified by the structure of the model (Phillips & Gregg, 2003).

351         Three other notable approaches have been developed to identify proportional source  
352 contributions. First, Lubtekin & Simenstad (2004) proposed two models (SOURCE and STEP)  
353 that are computationally less demanding than IsoSource. These models identify the outer bounds  
354 of possible mixtures in  $n$ -dimensional Euclidean space, instead of examining every single  
355 biologically possible solution. The output of these models is considerably reduced relative to  
356 that of IsoSource, but there is little reason to believe that the output of SOURCE and STEP  
357 would be significantly different from that of IsoSource (Maier & Simenstad, 2009). Second, the  
358 Moore-Penrose pseudoinverse model (Hall-Aspland, Hall & Rogers, 2005a; Hall-Aspland,  
359 Rogers & Canfield, 2005b) attempts to provide a unique solution of source contributions to a  
360 consumer using a single isotopic tracer and matrix algebra. Although output data often match up  
361 well with mean/modal resource values generated by IsoSource (S.A. Hall-Aspland, personal  
362 communication), this approach provides only a single solution and fails to acknowledge other  
363 feasible source combinations as provided by IsoSource. Third, a linear programming (LP) model  
364 employs linear algorithms instead of an iterative approach to determine the minimum and  
365 maximum possible proportions of each source to a consumer (Bugalho *et al.*, 2008). The results  
366 are similar to those produced by IsoSource, with the LP model explicitly identifying which  
367 sources definitively do or do not contribute to an individual consumer. Likely because of the

368 availability and ease of use of the IsoSource software, it is used much more frequently than  
369 SOURCE, STEP, Moore-Penrose, or LP models.

### 370 **(3) Bayesian mixing models**

371 A major limitation of all of the above mixing models (apart from Isoerror) is that they do  
372 not incorporate uncertainty and variation in input parameters (such as variation within source  
373 pools or trophic discrimination factors). In other words, much of the inherent variability in  
374 natural systems is ignored by use of mean resource isotope values or assumed trophic  
375 discrimination values. To this end, models (e.g. MixSIR and SIAR) have emerged, all of which  
376 are based on a series of related linear equations, that utilize Bayesian statistical techniques to  
377 identify proportional contributions of source pools (Jackson *et al.*, 2009; Moore & Semmens,  
378 2008; Parnell *et al.*, 2010; Solomon *et al.*, 2011). Importantly, these approaches allow for  
379 incorporation of available prior information, thereby allowing for more realistic representations  
380 of variability in input terms. Outputs from the Bayesian models are in the form of true  
381 probability distributions, not just summaries of all feasible solutions. As such, unlike in  
382 IsoSource, measures of central tendency from the outputs can be used in subsequent analyses  
383 (Parnell *et al.*, 2010). Further, parameter transformations, as suggested by Semmens *et al.*  
384 (2009b), provide a framework for utilization of general linear model approaches. This allows for  
385 incorporation of fixed and random covariates into models, which can provide the ability to  
386 partition particular drivers of source contribution variation (Francis *et al.*, 2011). Largely  
387 because of the additional input data, the models often substantially narrow the reported ranges of  
388 source pool contributions to consumers (Moore & Semmens, 2008; Moreno *et al.*, 2010).  
389 Bayesian approaches are evolving rapidly, greatly expanding in capability and scope (Jackson *et*  
390 *al.*, 2011; Ward *et al.*, 2011), and are being applied to yield novel insights into aspects of trophic

391 structure (e.g. Francis *et al.*, 2011; Rutz *et al.*, 2010; Solomon *et al.*, 2011). As with IsoSource,  
392 the Bayesian models MixSIR and SIAR can be freely accessed online (Moore & Semmens,  
393 2008; Parnell *et al.*, 2010).

394         Solomon *et al.* (2011) provide one example of how Bayesian approaches can yield  
395 powerful insights into the contribution of sources to consumers. Their goal was to quantify  
396 resource use for zooplankton, zoobenthos, and fishes in four low-productivity lakes, using  
397 models that incorporated multiple sources of potential variance and error. Informative priors  
398 (and/or associated variance components) utilized in the model included the proportion of  
399 hydrogen in consumer tissues derived from environmental water, trophic position of organisms,  
400 trophic discrimination factors, source isotopic signatures and a term to estimate unexplained  
401 variation. The results provided strong evidence that both terrestrial and benthic basal resource  
402 pools were integral in supporting consumer production in the lake systems. That is, even when  
403 accounting for many of the sources of input error that could have biased model output, terrestrial  
404 and benthic basal resource pools were identified as particularly important contributors. More  
405 generally, even though determined source ranges may still be broad in Bayesian models, there is  
406 greater assurance in their validity because of the incorporated error terms.

407         It is important to recognize that all mixing models, including Bayesian-based approaches,  
408 are not a quick fix or a substitute for poor sampling strategy; moreover, they are not particularly  
409 useful for asking questions about systems where complementary information is largely lacking.  
410 Indeed, all of the mixing models described above are subject to a core set of limitations, with  
411 many of the guidelines in Section II applying here. First, some information on turnover rate,  
412 trophic discrimination and macronutrient composition (e.g. free lipid and carbonate content)  
413 associated with the consumer tissues is needed. Second, prey sources must have different

414 isotope values. The more similar the resource pool isotope values, the less power the models  
415 have to delineate proportional contributions. Third, in many cases, *a priori* grouping may be  
416 necessary to constrain model outputs (Phillips *et al.*, 2005; but Bayesian approaches may also be  
417 useful in this respect, see Ward *et al.*, 2011), a decision that requires extensive knowledge of the  
418 basic natural history of the system. Fourth, prey should ideally be sampled on a time frame that  
419 coincides with the period during which the consumer tissue is synthesized, and all prey items  
420 must be known in order to provide the most meaningful results (although SIAR has an additional  
421 error term whereby the solution is not constrained to be merely a function of the identified  
422 sources, which would allow for some unknowns to be incorporated into the model). Fifth, as  
423 spatial and temporal variability in source pool values increases, so does the sampling effort  
424 necessary to determine adequately the appropriate input mean (and standard deviation in  
425 Bayesian models) values that should be used. As with any model, Bayesian tools such as  
426 MixSIR and SIAR are especially sensitive to the quality of the input data (Moore & Semmens,  
427 2008). Finally, inclusion of prior information into models can lead to more uncertain outputs,  
428 depending on the nature of input data (Moore & Semmens, 2008).

429         Although still beholden to the quality of input data, the ability to incorporate prior  
430 information and propagate error using Bayesian frameworks holds much promise in the  
431 development of the field. One of the primary criticisms of isotope applications in food-web  
432 studies is the lack of specific information on the underlying biochemical processes that affect  
433 isotopic signatures (Martinez del Rio *et al.*, 2009b); since Bayesian approaches allow for  
434 incorporation of uncertainty in input parameters, the models tacitly address this criticism by  
435 providing for more realistic estimates of source contributions to consumers.

#### 436 **(4) Spatially based approaches**

437           Mixing models typically utilize values for source pools that have fixed, distinct isotope  
438 values, but continuous variables can also be incorporated into mixing-model formats (Francis *et*  
439 *al.*, 2011; Gray *et al.*, 2011; Rasmussen, 2010). For example, Rasmussen (2010) describes a  
440 model that can be applied when isotope signatures are not necessarily distinct (e.g. when  $\delta^{13}\text{C}$  of  
441 sources overlap), but patterns of spatial variation occur. This model may be a useful tool in  
442 systems where source variability is predictable across some spatial axis (e.g. altitude, latitude, or  
443 river distance). The model uses the slopes of change along the spatial axis to estimate source  
444 contributions to the consumer by assuming that the consumer's isotope signature is a weighted  
445 mixture of the sources along the linear gradients. For example, aquatic and terrestrial resource  
446 pools may have the same mean isotope value across a distance gradient in river systems, but  
447 aquatic resources vary predictably with river distance (a predictable linear slope of distance  
448 *versus*  $\delta^{13}\text{C}$ ) while terrestrial sources remain consistent (Gray *et al.*, 2011). These relationships  
449 between distance and  $\delta^{13}\text{C}$  for terrestrial and aquatic resource pools allows the calculation of  
450 proportional resource contributions to the stream invertebrates (Rasmussen, 2010). The strengths  
451 of this approach are that it can overcome challenges involving overlap in resource-pool isotope  
452 values, and that it explicitly considers spatial variability. The main drawbacks are twofold. First,  
453 the proportions of the sources in a consumer's diet must be constant along the relevant gradient.  
454 Second, a detailed understanding of the underlying isotope gradients may be difficult to develop,  
455 if they exist at all, and the model will rapidly become mathematically intractable as the number  
456 of resource pools increases. As such, this approach may not be relevant in many systems.

457           Two other approaches also take advantage of spatial correlations to identify possible  
458 resource pools supporting consumers. Melville & Connolly (2003) sampled a consumer and its  
459 possible resource pools at many spatially distinct locations. For the isotopic niche, they

460 calculated the Euclidean distances ( $D$ ) between average consumer values and the resource pool  
461 averages at each location. Since the magnitude and directions of change of  $D$  were consistent  
462 across sampling locations, they suggested the consumer was “tracking” that resource pool and,  
463 therefore, it was an important part of the diet of that consumer. This approach does not provide  
464 estimates of the proportional contributions of sources, just an indication of which sources may or  
465 may not be important. In a similar across-site comparative approach, Vanderklift & Wernberg  
466 (2010) demonstrated, using partial regression analysis (controlling for within- and among-site  
467 source and consumer variation), that large-scale spatial variability in isotope signatures among  
468 sites can be used as a tool to identify diet sources of consumers. The strength of these two  
469 models is that they explicitly account for spatial variability in consumers and resource pools.  
470 There are two primary weaknesses. First, the models rest on the assumption that consumer diet  
471 items have unique isotope signatures and consumers have a relatively consistent, constrained,  
472 diet across sites. These factors must hold to link directly spatial variability in isotope values  
473 among source pools and consumer tissues. Second, many other ecological variables affect large-  
474 scale variability in isotope signatures, so ascertaining specific mechanisms giving rise to the  
475 isotopic niche may be difficult. These spatial-based approaches are likely to be most effective  
476 when used in conjunction with one of the aforementioned mixing models.

477

## 478 **VI. WHAT ADDITIONAL INFORMATION DOES RELATIVE POSITION OF** 479 **CONSUMERS IN ISOTOPIC SPACE REVEAL ABOUT FOOD-WEB STRUCTURE?**

480 In addition to estimating vertical position in a web and quantifying proportional  
481 contributions of source pools to consumers, stable isotope data can provide more general  
482 depictions of food-web structure. That is, important information may be gleaned simply from

483 relative spacing of target groups in isotopic niche space. Two general types of relative position  
484 metrics have been proposed: (1) one based on quantifying the amount of isotopic space occupied  
485 and (2) one quantifying relative change in position of target groups across temporal or spatial  
486 environmental gradients. These metrics often are applied in conjunction with trophic position  
487 and source contribution estimates to provide detailed information regarding trophic structure.  
488 However, they also may be informative even when limitations of particular data sets, e.g. lack of  
489 an adequate baseline or isotope source pools without distinct values, preclude precise  
490 calculations of trophic position and source contributions. In such instances, these two types of  
491 tools still allow for quantification of aspects of food-web structure, especially when these data  
492 are complemented with additional data sources (Layman & Post, 2008).

493         Layman *et al.* (2007a) proposed a series of metrics to quantify the area of isotopic space  
494 occupied by individuals or species. For example, the total area of a convex polygon  
495 encompassing all species within a community can be used as a measure of trophic diversity.  
496 That is, albeit with caveats associated with baseline resource pools, greater degree of isotopic  
497 niche space occupied relates to greater amount of trophic diversity among species (or  
498 individuals) in a community. In this way, overall trophic complexity is characterized by a single  
499 continuous variable which can be used to compare across systems or time periods. Likewise,  
500 other related metrics (e.g. mean nearest neighbour distance) further characterize spacing among  
501 individual data points in isotopic space, providing additional insight into trophic diversity and  
502 species packing within communities (Layman *et al.*, 2007a).

503         Various modifications of these simple metrics have been proffered. For example,  
504 baseline-corrected trophic position estimates have been used instead of absolute  $\delta^{15}\text{N}$  values in  
505 bivariate plots (Mercado-Silva, Helmus & Vander Zanden, 2009; Swanson, Kidd & Reist, 2010).



506 As mentioned above, raw isotope data also can be converted into proportion-space based on the  
507 contributions of the underlying resource pools (Newsome *et al.*, 2007). With this transformation,  
508 traditional metrics (such as Shannon-Wiener diversity) may be used to compare aspects of niches  
509 across species and systems. When possible, such transformations are desirable, although they  
510 become more ambiguous with increasing numbers of potential resource pools. In fact, when  
511 resource pool diversity is substantial, as is the case in many complex food webs, the  
512 transformation to proportional space is impossible (Layman & Post, 2008).

513 Another set of metrics is used to quantify directional shifts within isotopic niche space.  
514 This set of approaches is based on computed vectors of the directional change between mean  
515  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values. Wantzen *et al.* (2002) analyzed these vectors across species using two-  
516 dimensional ANOVA. Schmidt *et al.* (2007) introduced the use of circular statistics in which  
517 changes in the angle and magnitude of vectors in isotope space can be quantified. These vector-  
518 based approaches should prove especially powerful in analyzing changes through time,  
519 reconstructing historical food-web structure, and/or predicting future food-web patterns  
520 (Mercado-Silva *et al.*, 2009; Schmidt *et al.*, 2007; Schmidt, Zanden & Kitchell, 2009).

521 Turner, Collyer & Krabbenhoft (2010) have taken area-based (Layman *et al.*, 2007a) and  
522 directional (Schmidt *et al.*, 2007) metrics a step further, specifically by using nested linear  
523 models and a residual permutation procedure to provide for a quantitative hypothesis-testing  
524 framework. Specifically, their model allows for testing of shifts in (1) location and dispersion  
525 between isotopic groups indicating potential differences in resource use and niche breath (e.g.  
526 because of ontogeny or movements between isotopically distinct habitats) and (2) magnitude  
527 and direction of changes in centroid position between isotopic samples. We recommend the  
528 quantitative approaches of Turner *et al.* (2010) be used in conjunction with the area-based

529 (Layman *et al.*, 2007a) and directional (Schmidt *et al.*, 2007) metrics to provide increased  
530 quantitative rigour.

531         The area-based (Layman *et al.*, 2007a) and directional (Schmidt *et al.*, 2007) metrics  
532 share a fundamental set of strengths and weaknesses that reflect the underlying nature of the  
533 isotope data. Both types of analytical approaches serve to reduce food-web complexity into  
534 continuous metrics, which can subsequently be compared across systems or time periods. The  
535 measures are relatively simple to compute, and provide for direct measures regarding specific  
536 aspects of trophic structure. An additional distinction is that  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  (or other elemental  $\delta$   
537 values) are simultaneously analyzed, revealing insights that may not be clear when focusing on  
538 variation in  $\delta$  values for a single element. Further, intricacies of every factor affecting a single  
539 individual's isotope values (e.g. trophic discrimination) are not essential to elucidate general  
540 patterns in food-web structure.

541         Clear limitations of these approaches are apparent as well. First, as the number of  
542 consumer and resource species in a food web increases, sources of ambiguity become more  
543 likely. Second, comparisons among food webs become increasingly problematic as food webs  
544 become more dissimilar. For example, comparisons of niche width are not as meaningful when  
545 the focal food webs have very different basal resource pools (e.g. comparing a lake to a grassland  
546 food web). Third, the metrics are also especially sensitive to the sources of ecological ambiguity  
547 we introduced in the Section II. For example, similar food-web structures can give rise to very  
548 different metric values if the two webs have resource pools with underlying differences in  
549 relative  $\delta$  values (although, in some cases, it may be possible to scale baseline variation among  
550 the food webs being compared). Finally, two issues apply explicitly to the convex hull-based  
551 measures (Layman, 2007). Sample sizes of the groups being compared can cause interpretation

552 problems because the hull area will tend to increase with number of individual samples (Jackson  
553 *et al.*, 2011), yet this can be addressed by running bootstrap procedures to ensure sample size is  
554 sufficient to characterize fully the isotopic niche (Vaudo & Heithaus, 2011). Finally, a few  
555 individual outliers may result in a relatively large convex hull in which much of the contained  
556 niche space is unoccupied. In such a case, evaluating the relative merits of different ways to  
557 characterize the isotopic niche is warranted (see discussion of convex hulls *versus* Bayesian  
558 ellipse models in Section VII.1). In summary, the quantitative metrics discussed in this section  
559 have a series of caveats but, if qualified appropriately and augmented with additional sources of  
560 data, provide useful insight into particular aspects of food-web structure.

561

## 562 **VII. WHAT IS THE DEGREE OF TROPHIC VARIABILITY AT THE** 563 **INTRAPOPULATION LEVEL?**

### 564 **(1) General approaches**

565       There has been much renewed interest in the role of intrapopulation niche variation  
566 (Araújo, Bolnick & Layman, 2011; Bolnick *et al.*, 2011), with stable isotopes emerging as one of  
567 the primary tools for analysis. Many of the techniques used to examine trophic structure at the  
568 intrapopulation level are extensions of those used to examine the overall structure of food webs  
569 (see above), with intrapopulation groups defined using categories of sex, stage of maturity, or  
570 habitat use. In fact, intraspecific variation in resource use was among the first applications of  
571 stable isotopes in food-web ecology. For example, Fry *et al.* (1978) showed that variance of  
572 individuals'  $\delta^{13}\text{C}$  values was very low for some grasshopper species indicating no among-  
573 individual diet variation, whereas in other species variance was relatively large, suggesting that  
574 individuals consistently fed on either C3 or C4 plants. More recently, Martinez del Rio *et al.*

575 (2009a) adapted the Schmidt *et al.* (2007) vector-based approach, showing great variation in the  
576 magnitude and direction of changes in the isotopic niche of individual ovenbirds across seasons.

577 Two main categories of inquiry encompass many of the stable isotope applications in this  
578 context. First, many analyses are structured around using simple statistical tools (e.g. *t*-tests,  
579 ANOVA, linear regression) to examine ontogenetic diet shifts within populations. Specifically,  
580  $\delta^{15}\text{N}$  is used as a proxy to assess shifts in trophic position through ontogeny. For example,  
581 Jennings *et al.* (2002) examined the relationship between body size and trophic position for 31  
582 fish taxa in the North Sea, and demonstrated the prevalence of increasing trophic position  
583 through ontogeny for most species. Second, stable isotopes are used to estimate relative niche  
584 width of populations, typically by analyzing individual-level dispersion. Approaches include  
585 range or variance in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  (Bearhop *et al.*, 2004; McClellan *et al.*, 2010; Willson *et al.*,  
586 2010), convex hulls calculated at the individual level (Layman *et al.*, 2007b; Quevedo, Svanback  
587 & Eklov, 2009), relative spacing among individuals (Martinez del Rio *et al.*, 2009a), two-  
588 dimensional confidence intervals based on mean  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  (Layman, Winemiller &  
589 Arrington, 2005a), and standard ellipse areas (SEA<sub>B</sub>), i.e. bivariate equivalents to standard  
590 deviations in univariate analysis (Jackson *et al.*, 2011).

591 Each of these aforementioned approaches has strengths and weaknesses, and we provide  
592 one comparison as an example, i.e. between convex hulls (Layman *et al.*, 2007b) and the recently  
593 developed Bayesian approach (Jackson *et al.*, 2011). The convex hull approach is powerful  
594 because it incorporates each individual sampled, and thus includes information about every part  
595 of isotopic niche space occupied. Conversely, the Bayesian approach is targeted at niche widths  
596 of “typical” members in a population, which could be viewed as the mean or core isotopic niche  
597 of that population (Jackson *et al.*, 2011). The Jackson *et al.* (2011) approach generates standard

598 ellipse areas as this core isotopic niche representation ( $SEA_B$ ). Either the Bayesian approach or  
599 convex-hull-based quantitative analysis (Turner *et al.*, 2010) may be more appropriate with  
600 respect to a particular question of interest and/or the nature of the underlying data set. Convex  
601 hulls may be more appropriate when individual-level niche variation, and thus every niche  
602 position occupied by individuals, is central to the focal research question (Layman *et al.*, 2007b).  
603 When core aspects of a population's isotopic niche are of most interest, other prior information is  
604 available (e.g. on trophic discrimination rates), or error propagation is desirable, then the recently  
605 developed Bayesian-based approaches are preferable to characterize niche widths (Jackson *et al.*,  
606 2011). In some cases, utilization of both of these analytical approaches may be desirable to  
607 reveal different aspects of trophic structure.

## 608 **(2) Numerical simulations**

609 Another research area that has developed rapidly in recent years has been examining  
610 incidence and causes of individual specialization, i.e. variation in resource use among individuals  
611 that is not attributable to age class, size or sex (Araújo *et al.*, 2011; Bolnick *et al.*, 2003, 2011).  
612 Individual specialists utilize a relatively narrow subset of the population's overall resource base  
613 so that there is substantial variability in the specific resources used among individuals. Such  
614 variation among individuals may have several evolutionary and ecological implications,  
615 including driving frequency-dependent disruptive selection (Bolnick *et al.*, 2011; Dieckmann &  
616 Doebeli, 1999; Roughgarden, 1972) or imparting population stability (Agashe, 2009; Bolnick *et al.*,  
617 2011; Lomnicki, 1999). Quantification of individual specialization within populations would  
618 ideally be based on longitudinal samples in which the same individuals' diets are sampled  
619 repeatedly over time (Bryan & Larkin, 1972; Estes *et al.*, 2003; Werner & Sherry, 1987; West &  
620 Williams, 1986). However, in most cases, such longitudinal sampling schemes are too difficult to

621 implement. Because of the time- and space-integrated insight provided by isotopes, they have  
622 become the primary way to investigate instances of individual specialization (Araujo *et al.*, 2007;  
623 Beaudoin *et al.*, 1999; Bolnick *et al.*, 2007; Cherel *et al.*, 2007; Herman *et al.*, 2005), and a new  
624 set of analytical tools have been developed to this end.

625         Matthews & Mazumder (2004) proposed a null-model approach to test directly for  
626 specialization, incorporating information from source pools that could otherwise lead to  
627 erroneous interpretations of consumer isotope values. That is, ostensible sources of dietary  
628 specialization can be inferred directly from measures of variation in individual isotope values  
629 and by carefully selecting among consumer tissues. But for a given degree of individual  
630 specialization, populations using resources that span a wider range of  $\delta^{13}\text{C}$  or  $\delta^{15}\text{N}$  will show  
631 higher variability in consumer isotopes (Bearhop *et al.*, 2004; Newsome *et al.*, 2007). Matthews  
632 & Mazumder (2004) used a null model approach in which individuals sample randomly from a  
633 common resource pool to generate a null distribution of variances in  $\delta^{13}\text{C}$  among individuals.  
634 Empirical  $\delta^{13}\text{C}$  variance is tested against the null distribution, providing a statistical test for the  
635 presence of individual specialization.

636         A subsequent advance was to quantify the degree of individual specialization in a  
637 population, not just identify its presence/absence. Araújo *et al.* (2007) proposed a method that  
638 uses the variance in individual  $\delta^{13}\text{C}$  values in a population of consumers, and the  $\delta^{13}\text{C}$  values of  
639 resources, to calculate two indices of individual specialization that can be compared across  
640 different populations (Bolnick *et al.*, 2002). In this approach, null populations with varying  
641 degrees of individual specialization are generated, for which both isotope variances and indices  
642 of individual specialization are calculated. A curve relating the expected isotopic variances and  
643 indices of individual specialization is built and used to interpolate a measure of individual

644 specialization given an empirical variance in isotopes. This approach is especially useful for  
645 taxa which have relatively few items in their stomachs at any single time (e.g. piscivorous fish),  
646 for which estimations of dietary specialization would not be possible with direct diet analysis  
647 alone. The power of this approach has been illustrated in its first applications on frogs and birds  
648 (Araújo *et al.*, 2007; Woo *et al.*, 2008). However, this model has increased data input  
649 requirements, specifically, direct dietary information that corresponds to the time period that the  
650 isotope values of consumers and sources reflect. Again, collecting appropriately matched diet  
651 and isotope data sets can be difficult, especially as temporal and spatial heterogeneity in  
652 consumers (e.g. prey selection) and resources (e.g. seasonal variation in isotope values)  
653 increases. Such a null-model approach, however, is essential to identify true dietary  
654 specializations and should be employed in all cases in which isotope data are used directly to  
655 infer the degree of dietary specialization within a population.

656         Bayesian mixing models also can be used to examine niche variation and individual  
657 specialization within populations. The model of Semmens *et al.* (2009b) allows the partitioning  
658 of diet variation at different levels (e.g. individuals, sexes, morphs, age classes), providing  
659 insight that is not possible with other methods. The Bayesian framework also allows for  
660 incorporation of sources of variability that are not caused by diet variation, such as isotope  
661 variation within resources or variation in discrimination values among individual consumers.  
662 For populations of gray wolves (*Canis lupus*) in British Columbia, the model was used to show  
663 that not only do populations' diets differ because of geographic location, but diets also differed  
664 among packs and among individuals within packs (Semmens *et al.*, 2009b). It is possible to use  
665 the outputs of Bayesian mixing models to identify more broad patterns of specialization. For  
666 example, SIAR was recently used to define a set of foraging specialists from a population of

667 Northern Gannets *Morus bassanus*, and the output was linked to differences in fitness and  
668 foraging tactics among the specialist group (Votier *et al.*, 2010). Such ability to partition  
669 variance in isotope values across different hierarchical levels holds much potential.

### 670 (3) Different tissue types

671 Stable isotopes can also be used to track changes in individual-level resource use over  
672 time (Bearhop *et al.*, 2004; Hobson, 1993; Tieszen *et al.*, 1983). First, some tissues, such as  
673 hairs, feathers, and the dentine of teeth, are metabolically inert once they are deposited and  
674 therefore represent the isotope signature of a consumer's diet at the time of deposition. If the  
675 rate of tissue deposition is known, these tissues represent a timeline of the consumer's dietary  
676 history. For example, Hobson & Sease (1998) documented ontogenetic isotope shifts in Steller  
677 sea lions (*Eumetopias jubatus*) from tooth annuli. A more quantitative approach was proposed  
678 by Newsome *et al.* (2009) using small sections of Californian sea otter *Enhydra lutris nereis*  
679 whisker as a temporal series of resource use. They applied an ANOVA model to partition the  
680 variance in isotopes into a within-individual component (WIC, variation within an individual sea  
681 otter whisker) and a between-individual component (BIC, measured by differences between  
682 individual sea otter whiskers). Similarly, Jaeger *et al.* (2010) collected multiple feathers from  
683 individual seabirds to estimate Roughgarden's (1974) index of individual specialization  
684 ( $WIC/TNW$  where  $TNW$  is the total niche width of the seabird population), using the variation  
685 within an individual's feathers as an estimate of WIC and the total variation among individuals'  
686 feathers as an estimate of TNW. For such studies, some information on the nature of the inert  
687 tissue's deposition, e.g. whether it is continuous (e.g. the whiskers of some mammal species) or  
688 discontinuous (e.g. feathers), is necessary for appropriate analysis.



689           An alternative approach is based on the fact that different tissues have different turnover  
690 rates and therefore integrate resource use over different time scales (Hesslein, Hallard & Ramlal,  
691 1993). For example, vertebrate blood plasma integrates diets over a time scale of days to weeks,  
692 whereas turnover in muscle tissue is on the scale of months (Dalerum & Angerbjorn, 2005;  
693 MacNeil, Drouillard & Fisk, 2006; Phillips & Eldridge, 2006). As a consequence, individuals  
694 that feed consistently on the same resource(s) over long time scales should have similar isotope  
695 values in tissues with different turnover rates, whereas individuals that switch their diets over  
696 time should show a mismatch between fast and slow turnover tissues. Martinez del Rio *et al.*  
697 (2009a) called the former “isotopic specialists” and the latter “isotopic generalists”. They applied  
698 this framework to three species of ovenbirds and found that one species was made up of isotopic  
699 generalists that switch diets seasonally, another species had a mix of isotopic generalists and  
700 specialists, and the third species was composed of isotopic specialists. Likewise, Matich,  
701 Heithaus & Layman (2010) used different tissues to quantify differences in the degree of dietary  
702 specialization between bull sharks (*Carcharhinus leucas*) and tiger sharks (*Galeocerdo cuvier*).  
703 Importantly, information regarding turnover rates in the different tissues is needed to make  
704 inferences about the degree of dietary specialization. These approaches are especially sensitive  
705 to assumptions regarding isotopic routing and different discrimination factors among tissue  
706 types.

707           A general concern for all the methods outlined herein relates to the temporal and spatial  
708 scales at which individuals are sampled. Sampling individuals at different times or different  
709 locations might artificially inflate variation in isotope values if sources vary temporally and/or  
710 spatially. For example, individuals specialized on the same resource but feeding consistently in  
711 different areas may differ greatly in isotope values if there is spatial heterogeneity in resource

712 isotope values, so that habitat-derived variation in consumers' isotopes will be mistaken as diet  
713 variation (Flaherty & Ben-David, 2010). As is often the case, knowledge of the temporal and  
714 spatial variation in sources' isotopes, as well as organism natural history, will greatly aid in the  
715 interpretation of isotope data.

716

## 717 **VIII. CONCLUSIONS**

718 (1) Quantitative analytical approaches for applying stable isotope data have proliferated rapidly  
719 over the past decades. The numerous choices for analyzing data bode well for the continued  
720 development of stable isotope analysis of food-web structure. We hope this review provides one  
721 framework from which researchers can select the most appropriate tools for particular questions  
722 of interest. Moving forward, we suggest the guidelines listed below for practitioners in the field  
723 using the analytical approaches discussed herein.

724 (2) Stable isotope analysis is not a substitute for a basic understanding of the natural history of  
725 the organism or ecosystem of interest. Stable isotopes are an important tool that can be used to  
726 provide insight into food-web structure, but these data alone cannot elucidate the complexities  
727 that are manifest in food webs.

728 (3) There are still huge gaps in the empirical data needed to support analytical approaches,  
729 including data on isotope incorporation rates and routing into tissues, tissue turnover rates, and  
730 trophic discrimination factors (Martinez del Rio *et al.*, 2009b). Additional field and laboratory  
731 experiments are needed to this end.

732 (4) All models are beholden to the quality of input data available. Further, even basic stable  
733 isotope calculations that are well accepted in the literature, e.g. estimates of trophic position and  
734 food-chain length, should be qualified appropriately because of the lack of the underlying  
735 empirical data (e.g. on trophic discrimination and adequate baselines) necessary to produce  
736 adequate estimates.

737 (5) The validity of assumptions underlying analytical models will vary depending on the  
738 organism or system of interest. For example, the assumption that isotopic routing does not occur  
739 is especially problematic for omnivores (Kelly & Martinez del Rio, 2010; Martinez del Rio *et al.*,  
740 2009b; Voigt *et al.*, 2008). As such, the appropriateness of a specific analytical approach must  
741 be evaluated on a case-by-case basis. By applying corrections, e.g. for concentration dependence  
742 or increasing uncertainty in the trophic discrimination factors, it is becoming increasingly  
743 possible to deal with these issues.

744 (6) Many of the ecological questions discussed herein are necessarily reliant on the fact that  
745 source pools must have distinct isotope values. When sources are not distinct (or overlap to  
746 some degree), stable isotopes may have little utility in answering questions about trophic  
747 relationships.

748 (7) Both source and consumer pools must be sampled on proper spatial and temporal scales that  
749 reflect the relative incorporation and turnover rates of tissues. Establishing appropriate isotope  
750 end members and baselines remains of core importance for many of the analytical techniques.

751 (8) Because of the different underlying structure of analytical models, multiple approaches on the  
752 same data set are often warranted. Different analytical approaches may give rise to conflicting

753 output data, the magnitude and importance of which will vary based on the nature of the  
754 underlying data sets (Ikeda *et al.*, 2010; Maier & Simenstad, 2009; Moore & Semmens, 2008).

755 (9) While it is always tempting to favour approaches that provide analytical solutions, it is  
756 important to think about what the associated assumptions and simplifications might mean  
757 biologically.

758 (10) When possible, stable isotope analysis should always be augmented with additional data  
759 sets, particularly diet analysis or other data on feeding behaviour (Layman & Post, 2008). In  
760 fact, many of the approaches discussed herein require specific information on consumer's diets  
761 to parameterize models, *a priori* reduce the number of potential source pools, and define priors  
762 in Bayesian models. Stable isotope data in isolation cannot provide answers for all questions  
763 regarding food-web structure, and traditional dietary analysis will continue to be a core tool.

764 (11) In addition to diet data, stable isotopes used in conjunction with other dietary tracers (fatty  
765 acids, Boecklen *et al.*, in press; Budge *et al.*, 2008; Cheung & Sanyal, 2010) will likely provide  
766 new insights into food-web structure. Technological advances are expanding the potential suite  
767 of tools that can be employed. Compound-specific isotopic analysis may be one of the most  
768 important areas of future development (Chikaraishi, Ogaw & Ohkouchi, 2009b; Chikaraishi *et*  
769 *al.*, 2009a; McMahon *et al.*, 2010; Boecklen *et al.*, in press).

770 (12) Our views of food-web structure have, until recently, been largely constrained to examining  
771 population "means". Stable isotopes have been a core tool in elucidating the importance of  
772 intrapopulation niche variation (Araujo *et al.*, 2007; Layman *et al.*, 2007b; Quevedo *et al.*, 2009).  
773 Results of increasingly powerful analyses of stable isotope data sets will help reveal when  
774 intrapopulation niche variation is necessary to characterize adequately food-web structure.

775 (13) Stable isotopes, irrespective of the way they are analyzed, provide information regarding the  
776 flow of energy or nutrients through food webs. They do not provide definitive information as to  
777 the functional relationships among organisms (e.g. whether a predator controls the abundance of  
778 a given prey), information that typically necessitates controlled experimental manipulations.  
779 This distinction between energy flow and interaction food-web models must always be  
780 considered (Paine, 1980; Polis & Winemiller, 1996).

781 (14) The main caveat to using stable isotopes in a food-web context is that the data are only  
782 indirect indicators of feeding pathways. Nevertheless, the diversity and scope of papers included  
783 in this review reflects the important advances that stable isotopes have provided in food-web  
784 ecology. As analytical approaches become more advanced, stable isotopes should provide for  
785 many more important developments in the field.

786

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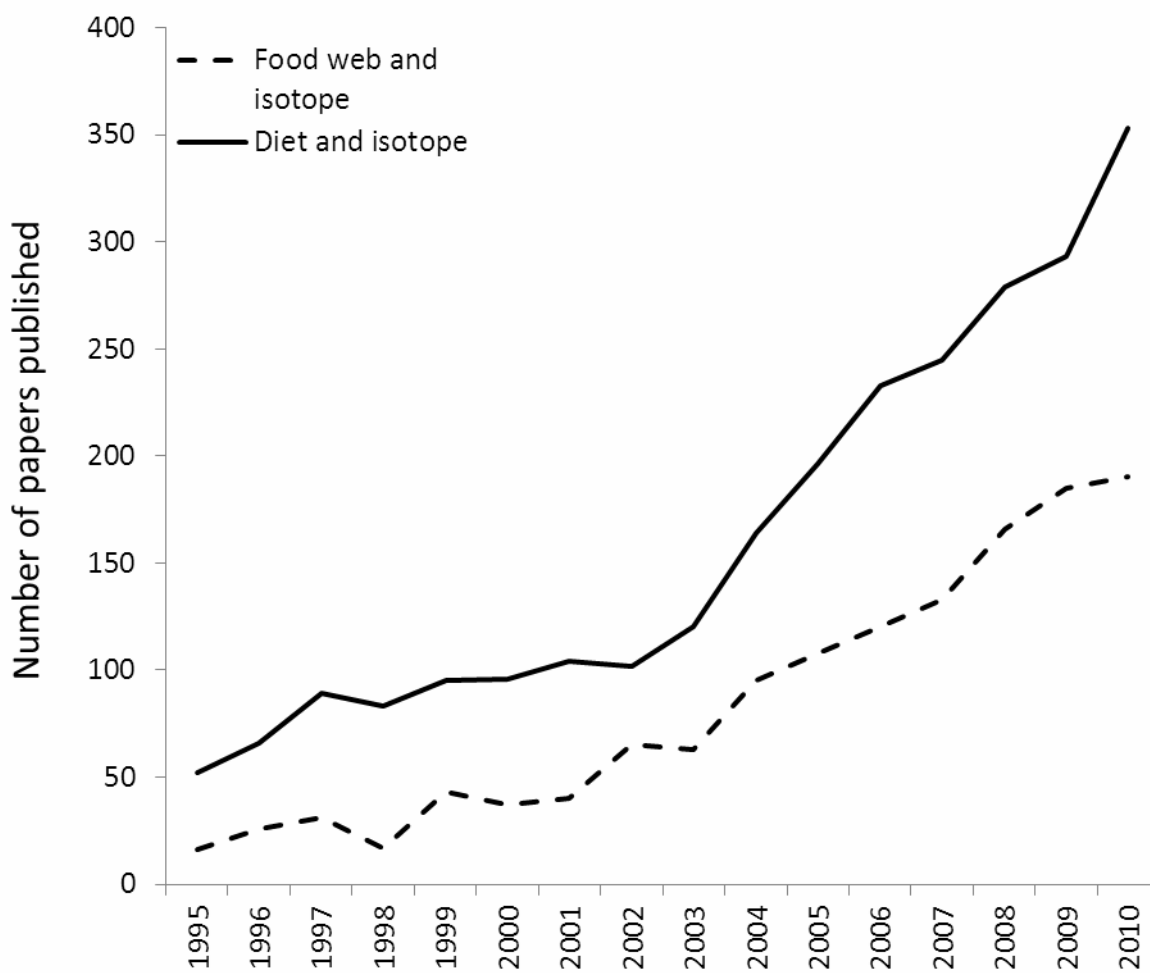
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1150 **Fig. 1.** Number of food web papers, as cataloged by *Web of Science*, employing stable isotopes  
1151 published each year. Papers were identified using the key words “food web” and “isotope” or  
1152 “diet” and “isotope”.

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