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A Concise Atlas of Thyroid Cancer Next-Generation Sequencing Panel ThyroSeq v.2

Tiroid Kanser Genomisinin Güncel Klinik Atlası: ThyroSeq v.2 Yeni Jenerasyon Dizinleme Paneli

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Abstract

The next-generation sequencing technology allows high out-put genomic analysis. An innovative assay in thyroid cancer, ThyroSeq[®] was developed for targeted mutation detection by next generation sequencing technology in fine needle aspiration and tissue samples. ThyroSeq v.2 next generation sequencing panel offers simultaneous sequencing and detection in >1000 hotspots of 14 thyroid cancer-related genes and for 42 types of gene fusions known to occur in thyroid cancer. ThyroSeq is being increasingly used to further narrow the indeterminate category defined by cytology for thyroid nodules. From a surgical perspective, genomic profiling also provides prognostic and predictive information and closely relates to determination of surgical strategy. Both the genomic analysis technology and the informatics for the cancer genome data base are rapidly developing. In this paper, we have gathered existing information on the thyroid cancer-related genes involved in the initiation and progression of thyroid cancer. Our goal is to assemble a glossary for the current ThyroSeq genomic panel that can help elucidate the role genomics play in thyroid cancer oncogenesis.

Keywords: Thyroid cancer, next generation sequencing, ThyroSeq

Öz

Tiroid nodüllerinde kanser teşhisi ve tiroid kanserlerinin prognostik değerlendirilmesi için geliştirilmiş en güncel yöntem DNA dizinleme teknolojisine dayalı ThyroSeq[®] testidir. Bu test ince iğne aspirasyon materyalinde genetik profileme yaparak tiroid kanserinin tetiklenmesinde ve gelişmesinde rol alan mutasyon ve genetik değişiklikleri tespit etmektedir. Bu makalede bu test ile tespit edilebilen genetik değişikliklere konu olan genlerin yapı ve fonksiyonlarını kısaca tanımlıyoruz. Tiroid kanseri onkogenezi bir ölçüde ışık tutan bu test için sözlük görevi tutan bir makale sunulmaktadır.

Anahtar kelimeler: Tiroid kanseri, yeni jenerasyon dizinleme, ThyroSeq

Introduction

Thyroid nodules are prevalent in the general population. Most thyroid nodules are benign and the clinical challenge is to accurately identify those nodules that are malignant and need to be surgically removed (1). Moreover, the extent of initial surgical treatment requires better understanding

of particular tumor biology beyond conventional definitions. Molecular pathology is the new paradigm in cancer diagnosis and prognostication. Thyroid cancer develops and progresses through accumulation of genetic alterations, which does serve as important diagnostic, prognostic, and predictive biological markers (2). Next-generation sequencing technology allows high out-put

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genomic analysis. An innovative assay in thyroid cancer - ThyroSeq® - was developed for targeted mutation detection by next generation sequencing technology in fine needle aspiration and tissue samples. ThyroSeq v.2 next generation sequencing panel offers simultaneous sequencing and detection in >1000 hotspots of 14 thyroid cancer-related genes and for 42 types of gene fusions known to occur in thyroid cancer (3). ThyroSeq is being increasingly used to further narrow the indeterminate category defined by cytology for thyroid nodules. From a surgical perspective, understandably this provides prognostic and predictive information as it relates to determination of surgical strategy. Both the genomic analysis technology and the data collection for the cancer genome atlas are rapidly developing.

This paper reviews basic genomic information on 23 thyroid cancer-related genes involved in thyroid cancer.

We have detailed information in regards to the location, and function of these genes in normal thyroid cells. We also report gathered information as to the consequences mutations to these 23 genes can have on thyroid cancer initiation and progression. Our goal is to provide a detailed glossary for ThyroSeq mutation panel.

Molecular Markers of ThyroSeq Next-Generation Sequencing Panel

B-RAF

The *B-RAF* gene, located on chromosome 7q34, encodes *B-RAF* serine-threonine kinase, which functions as an intracellular effector of the *RAS*/MAPK signaling cascade (Figure 1). This is one of the three isoforms of the *RAF* serine-threonine kinase and the predominant isoform found

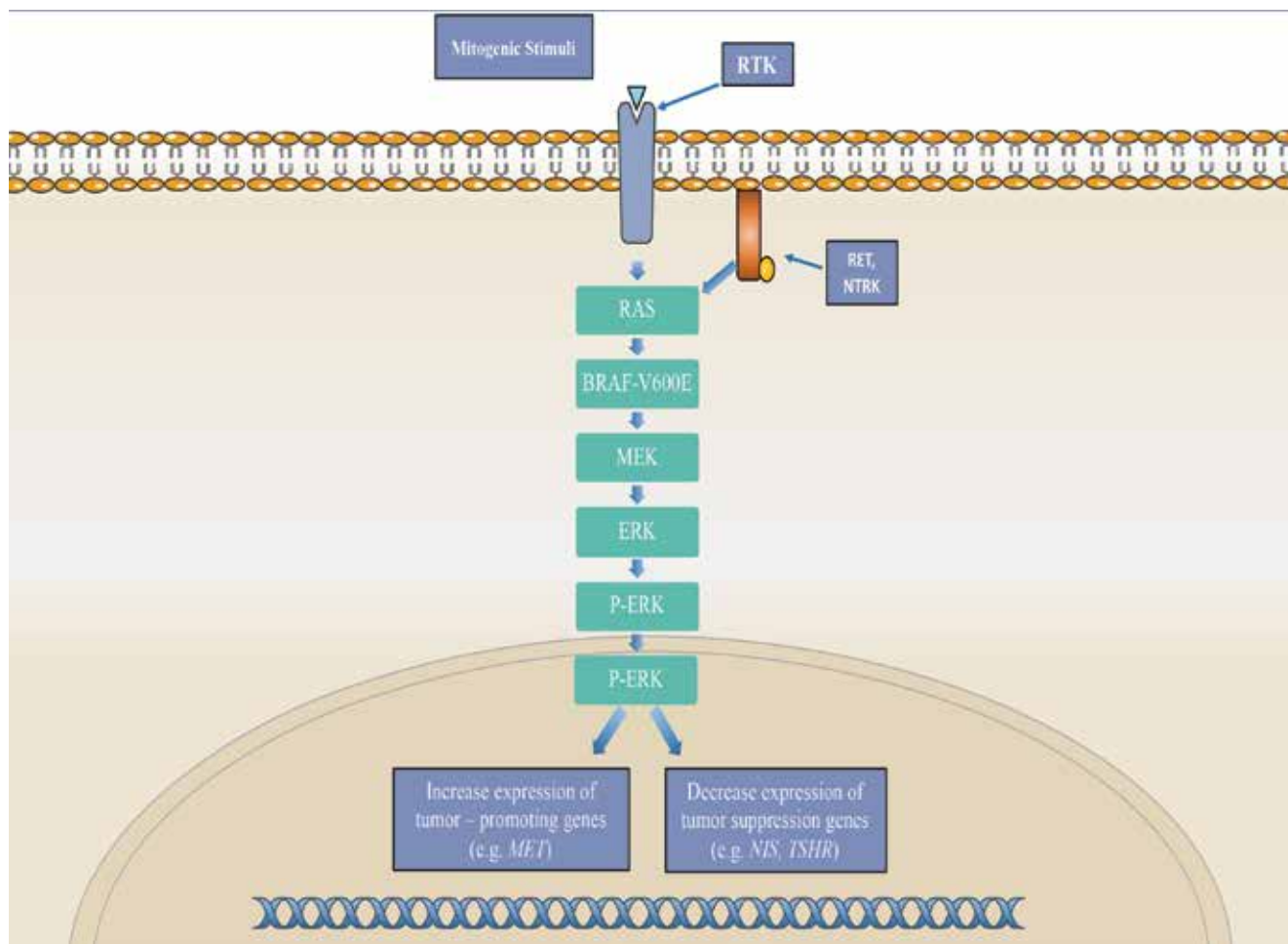


Figure 1. MAPK/ERK pathway

*This pathway begins with a mitogenic stimulus binding to a receptor tyrosine kinase, activating it. This triggers a cascade of protein activation, beginning with RAS and culminating with ERK. Once ERK is phosphorylated, it enters the nucleus and influences transcription, increasing expression of tumor-promoting genes and decreasing expression of tumor-suppressing genes. MAPK signaling can also be stimulated by genetic alterations in proteins RET and NTRK

in thyroid follicular cells. In wild-type forms of this gene, the *B-RAF* protein is activated through binding of a *RAS*-GTP protein complex with the *B-RAF*'s *RAS* binding domain along with simultaneous conformational changes in the protein. Once activated, the *B-RAF* protein phosphorylates the next protein in the signal cascade-MEK and ERK. The protein's function contributes to the *RAS*/MAPK pathway's role in cell proliferation, migration, and differentiation (4,5). The most common *B-RAF* mutation found in thyroid carcinomas is a point mutation at residue 600 involving a substitution from valine to glutamate (V600E). This mutation results in the constitutive activation of the *B-RAF* protein and subsequently the *RAS*/MAPK pathway. The activation of the *B-RAF* protein seems to be caused by a disruption of the hydrophobic interactions between its activation loop and the ATP binding site. In wild-type *B-RAF*, these hydrophobic interactions help maintain the protein in an inactive conformation. When disrupted, *B-RAF* remains in an active, catalytic conformation. This results in the constitutive phosphorylation of its downstream targets (4). The *B-RAF* V600E point mutation is most prevalent in papillary thyroid carcinomas (PTC)-the most common form of well differentiated thyroid carcinoma-found in 45% of PTC cases. Though it is rare in follicular variants of thyroid carcinoma, *B-RAF* is an ideal genetic marker for use in a thyroid cancer sequencing panel. It is found in all forms of thyroid carcinoma and seems to play a very important role early in tumorigenesis as a driver mutation (4,5,6).

RAS

The *RAS* genes consist of a family of highly homologous isoforms: *K-RAS*, *N-RAS*, and *H-RAS*. Located on chromosomes 12p12.1, 11p5.5, and 1p13.1 respectively, all three genes encode G-proteins located on the inner surface of the cell membrane. These proteins help convey signals from receptor tyrosine kinases (RTKs) to the pathways *RAS* regulates: *RAS*/MAPK and PI3K/AKT signaling cascades (Figure 2). Once bound to GTP, the *RAS* proteins proceed to activate the *RAS*/MAPK pathway. The *RAS*-GTP complex quickly becomes inactive as a result of the protein's innate GTPase activity (7).

Point mutations in the *RAS* gene are the most common mutations. Mutations in codons 12 and 13 lead to an increased affinity for GTP. Mutations in codon 61 lead to inactivation of the *RAS* protein's innate GTPase function. Overall, these mutations result in the constitutive activation of the *RAS* protein and thus the activation of the downstream signaling pathways it regulates, a critical step in thyroid tumorigenesis. Thyroid carcinoma has been associated with mutations in all three isoforms of the *RAS* genes, though it seems the more prevalent mutated isoform is that of *N-RAS*. However, the actual pattern of isoform frequencies in thyroid carcinoma remains unclear (4,7).

RAS mutations occur with variable frequency in all types of thyroid follicular-derived tumors. *RAS* point mutations are most common in follicular thyroid carcinoma (FTC), occurring in 40-50% of tumors, as well as in poorly differentiated and anaplastic thyroid carcinoma (PDC and ATC). It is more infrequent in PTCs, occurring in about 10% of tumors. The prevalence of *RAS* mutations in thyroid carcinoma make it a viable genetic marker as well as a useful prognostic tool, given that, studies suggest it may increase the potential for malignant transformation and tumor progression (7).

CTNNB1

The *CTNNB1* gene, located on chromosome 3p21, encodes a cytoplasmic protein known as β -catenin. This protein plays several important roles in the cell. It is involved in E-cadherin mediated cell to cell adhesion, found primarily in adherens junctions. It is also an intermediate in the Wnt signaling pathway. Once activated by proteins upstream in the pathway, β -catenin is able to accumulate in the cytoplasm and eventually is translocated into the nucleus. In the nucleus, the protein works with T-cell factor/lymphoid enhancing factor (TCF/LEF) to modulate gene expression (8,9).

When β -catenin is not activated, free cytoplasmic levels are kept low through ubiquitin-proteasome degradation (Figure 3). However, point mutations in exon 3 of *CTNNB1* confer added stability to the protein and thus inhibit the degradation of β -catenin. This results in the accumulation and localization of β -catenin in the cytoplasm and nucleus of the cell and the constitutive activation of its targeted genes (ex: Cyclin D1, C-MYC, C-JUN) (8,10,11).

Mutations in *CTNNB1* are most commonly found in PDC and ATC but not in well differentiated forms. It is typically described as a late event in thyroid cancer progression, but dysregulation of β -catenin has been shown to contribute to tumor proliferation (10,11).

CTNNB1 can serve as a useful marker for PDC and ATC. Its involvement in the Wnt pathway contributes to its viability as a marker, given this pathway is constitutively active in 50% of PDC and ATC and it is involved in the dedifferentiation process of thyroid carcinoma (Figure 4) (10). This gene is thus a useful diagnostic marker as well as prognostic tool to determine the progression of the cancer and its propensity to spread.

NTRK1

The *NTRK1* gene, which is located on chromosome 1q21-22, encodes the neurotrophic RTK-1, also known as TrkA. It is a high affinity receptor for nerve growth factor- β (NGF- β) that provides instructions for the growth and development of nerve cells. It also induces the proliferation of a number of cell types, such as lymphocytes and keratinocytes. Once it is

bound to NFG- β , *NTRK1* dimerizes and autophosphorylates its five tyrosine residues, which then act as binding sites for several target proteins such as *RAS* and *PI3K*. By activating these proteins, *NTRK1* subsequently activates the *RAS*/MAPK and *PI3K*/AKT signaling cascades, thus mediating NGF effects on the proliferation and differentiation of cells (12,13).

Mutations in the *NTRK1* gene are frequently found in PTC. These mutations involve chromosomal rearrangements between *NTRK1* and at least three other identified genes: *TPM3*, *TPR*, and *TFG*. The chimeric oncogenes that are formed (TRK oncogenes) encode constitutively active RTKs. It remains active due to the fact that its kinase domain and the tyrosine binding sites are preserved in the chimeric oncogene. The only portion that is lost is a portion of its extracellular domain. It seems that the extracellular

domain may contain important regulatory elements. This provides constant activation of the protein signals for cell proliferation, contributing to PTC (12,13).

About 60% of PTC carry chimeric oncogenes created through chromosomal rearrangement. Though *RET* rearrangement and not *NTRK1* is the most prevalent chimeric oncogene detected in PTC, thyroid epithelium seems to have a propensity for chromosomal rearrangements. That fact, along with *NTRK1* association with pathways implicated in thyroid carcinoma (*RAS*/MAPK and *PI3K*/AKT) make this gene a viable marker for diagnostic and prognostic purposes. Nevertheless, further investigation is necessary to better understand its role in thyroid carcinogenesis (13,14).

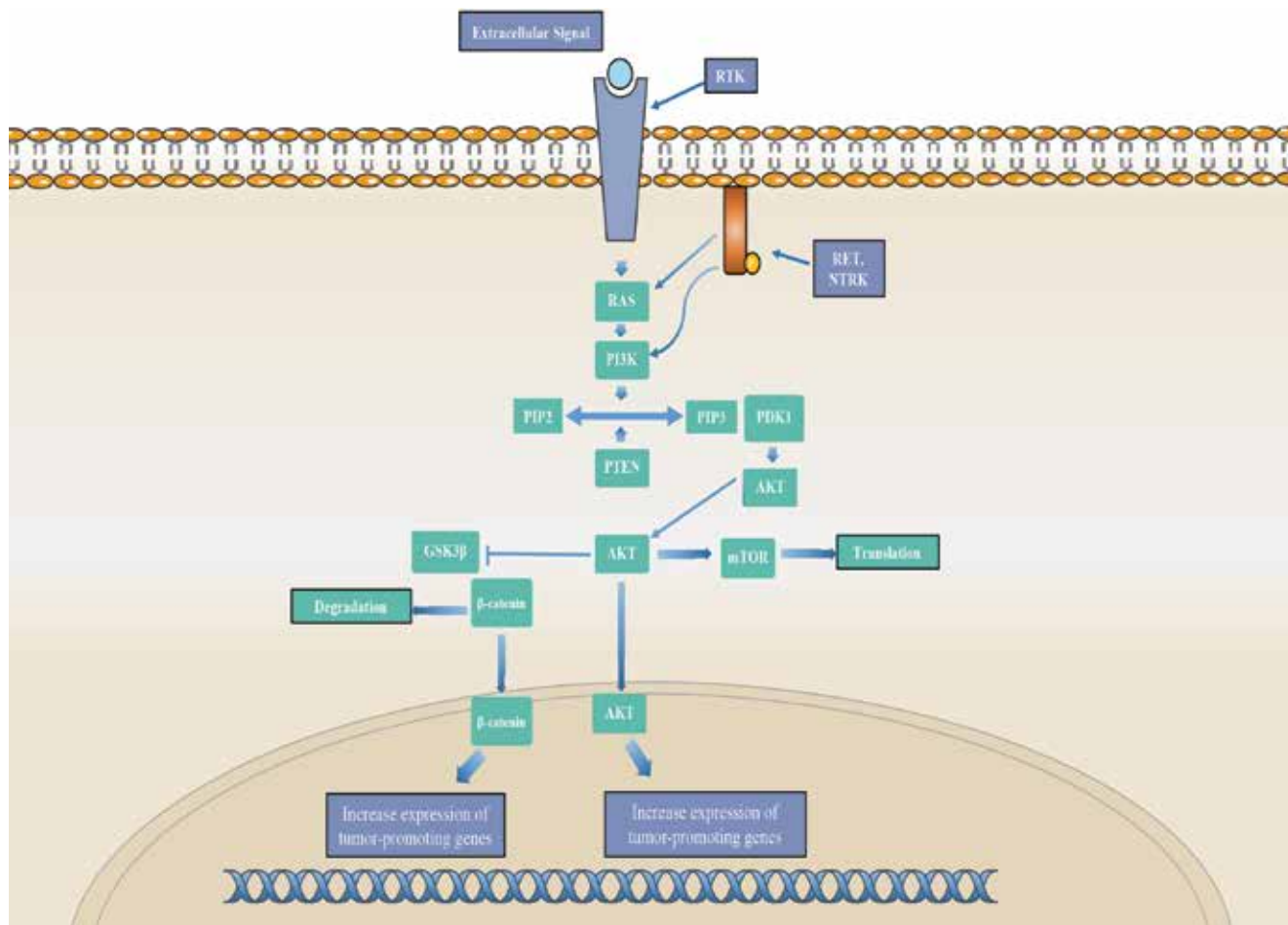


Figure 2. PI3K/AKT pathway

*The pathway begins with an extracellular stimuli activating a receptor tyrosine kinase, triggering the activation of *RAS* followed by *PI3K*. Activated *PI3K* catalyzes the conversion of phosphatidylinositol (4,5)-biphosphate (*PIP2*) to phosphatidylinositol (3,4,5)-trisphosphate (*PIP3*), a step that is regulated by *PTEN*. *PIP3* activates the 3-phosphoinositide-dependent protein kinase 1, which targets and phosphorylates *AKT*. Phosphorylated *AKT* then enters the nucleus, where it increases the expression of tumor-promoting genes. Activated *AKT* also produces changes in the cytoplasm by activating *mTOR* and thus promoting translation. *AKT* can also phosphorylate glycogen synthase kinase 3- β (*GSK3- β*), inactivating the protein. *GSK3- β* inhibition results in a concomitant stimulation of β -catenin, allowing it to enter the nucleus and increase expression of tumor-promoting genes

NTRK3

Another member of the neurotrophic tyrosine receptor kinase family, *NTRK3* encodes a RTK with a high affinity for its ligand neurotrophin-3. The gene is located on chromosome 15q25. Once bound, *NTRK3* autophosphorylates and proceeds to phosphorylate target proteins primarily involved in the *RAS*/MAPK and PI3K/AKT signaling pathways. Primarily expressed in the central nervous system, *NTRK3* is involved in cell proliferation and differentiation as well as neuronal cell processes (14,15).

The mutation of *NTRK3* observed in thyroid carcinomas is a chromosomal rearrangement involving *ETV6*, a gene encoding a transcription factor from the ETS transcription factor family. This particular oncogenic fusion involves the fusion of exons 1-4, the SAM domain, of *ETV6* and exons 12-18, the tyrosine kinase domain, of *NTRK3*. This results in the constitutive activation of the RTK. It is this constant

activation that contributes to thyroid carcinogenesis (15,16).

Chromosomal rearrangements of the NTRK genes are uncommon in sporadic PTC, only occurring in about 2% of cases. However, there is higher prevalence of these chimeric oncogenes in pediatric and radiation-related PTC, with the majority of *ETV6-NTRK3* fusions demonstrating the follicular variant of PTC. Mutations resulting in *RAS*/MAPK activation in radiation-related PTC tend to differ from those found in sporadic PTC as there tends to be more chromosomal rearrangements than point mutations (15,16).

RET

The *RET* gene is a proto-oncogene that is highly expressed in parafollicular C-cells. Located on chromosome 10q11.2, this gene encodes a RTK that is involved in the *RAS*/MAPK

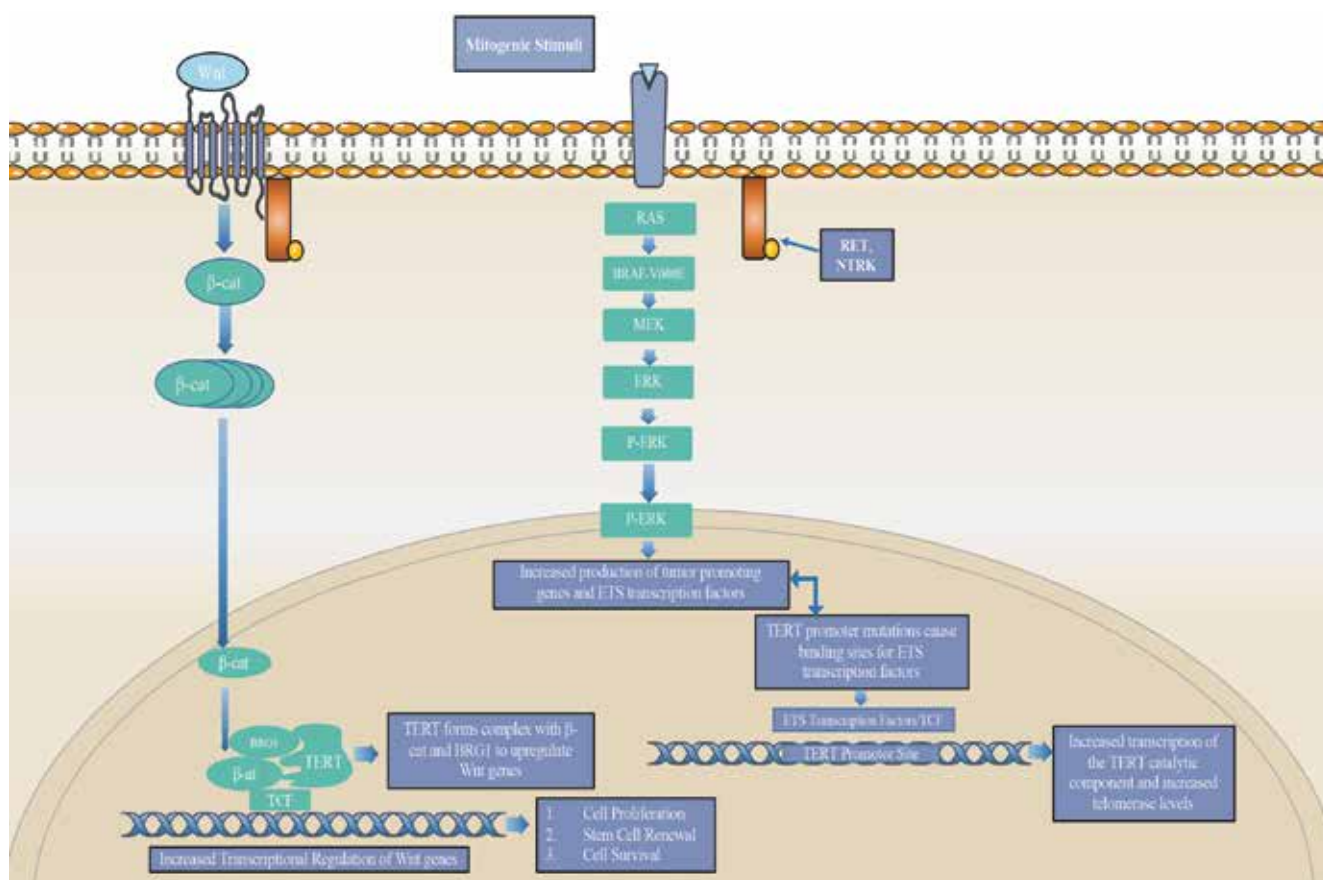


Figure 3. CTNNB1/TERT signaling

*Dysregulation of telomerase is an almost universal feature of cancer, either through genetic amplification of the gene's own locus or through crosstalk activity with other signaling pathways. In this pathway mutations to the promoter site of *TERT* (C228T and C250T) increase the transcription of *TERT* through the creation of consensus binding sites for ETS transcription factors. Constitutive action of the MAPK pathway due to B-RAF or RAS mutations leads to upregulation of *TERT* from ETS transcription binding activity. *TERT* also possess a role as a transcriptional modulator of the Wnt-β-catenin signaling pathway. Wnt stimulation at the plasma membrane leads to increased levels of β-catenin in the cytoplasm, and translocation to the nucleus. In the nucleus β-catenin forms a complex with *TERT* and the Wnt transcription factor *BRG1*. Once formed this complex binds to the promoters of Wnt target genes regulating the expression of these oncogenes

signaling pathway. The *RET* protein responds to signals from the cell's environment and then proceeds to activate itself through autophosphorylation. Once activated, the *RET* protein elicits changes within the cell often involving cell proliferation and development (4,17).

Chromosomal rearrangements involving *RET* represents one of the most common causes of PTC. At least 11 rearrangement variants have been isolated but the most common translocations are *RET/PTC1* and *RET/PTC3*. The aforementioned rearrangements are intrachromosomal paracentric inversions and account for the vast majority of variants found in PTC. These rearrangements result in a constitutively active *RET* protein, leading to activation of the *RAS*/MAPK pathway and driving thyroid carcinogenesis (17,18).

The *RET/PTC1* rearrangement involves interaction between *RET* and the *H4* gene. This particular rearrangement is the dominant type in sporadic PTCs. The *RET/PTC3* rearrangement involves interaction between *RET* and the *ELE1* gene. This particular rearrangement is the dominant type in radiation-related PTCs, which tend to be more aggressive and exhibit a higher frequency of chromosomal rearrangements (4,17).

AKT1

AKT1 codes for the *AKT1* kinase, one member of a family of homologous isoforms. Located on chromosome 14q32.322, *AKT1* kinase helps regulate cell growth and survival as well as the process of apoptosis, mainly through interaction with signaling pathways like PI3K/AKT. The protein contains a pleckstrin homology domain, making it a target for phospholipids generated by PI3K activity. Once activated through phosphorylation, *AKT1* binds to chaperone proteins and proceeds to phosphorylate its downstream effectors. Through this pathway, activated *AKT1* is able to inhibit apoptosis and induce cell growth in thyroid cells (19).

Activation of *AKT1* is known to promote thyroid tumorigenesis in Cowden's syndrome (CS). It is the overexpression and overactivation of *AKT1*, rather than mutation of the *AKT1* gene, that contributes to the pathogenesis of sporadic thyroid cancers, particularly in follicular thyroid cancers (FTC). In FTCs, activation of *AKT1* kinase and the localization of the protein in the nucleus promotes the PI3K/AKT pathway, contributing to tumor invasion and metastasis. It seems that overexpression of *AKT1* is capable of promoting cell

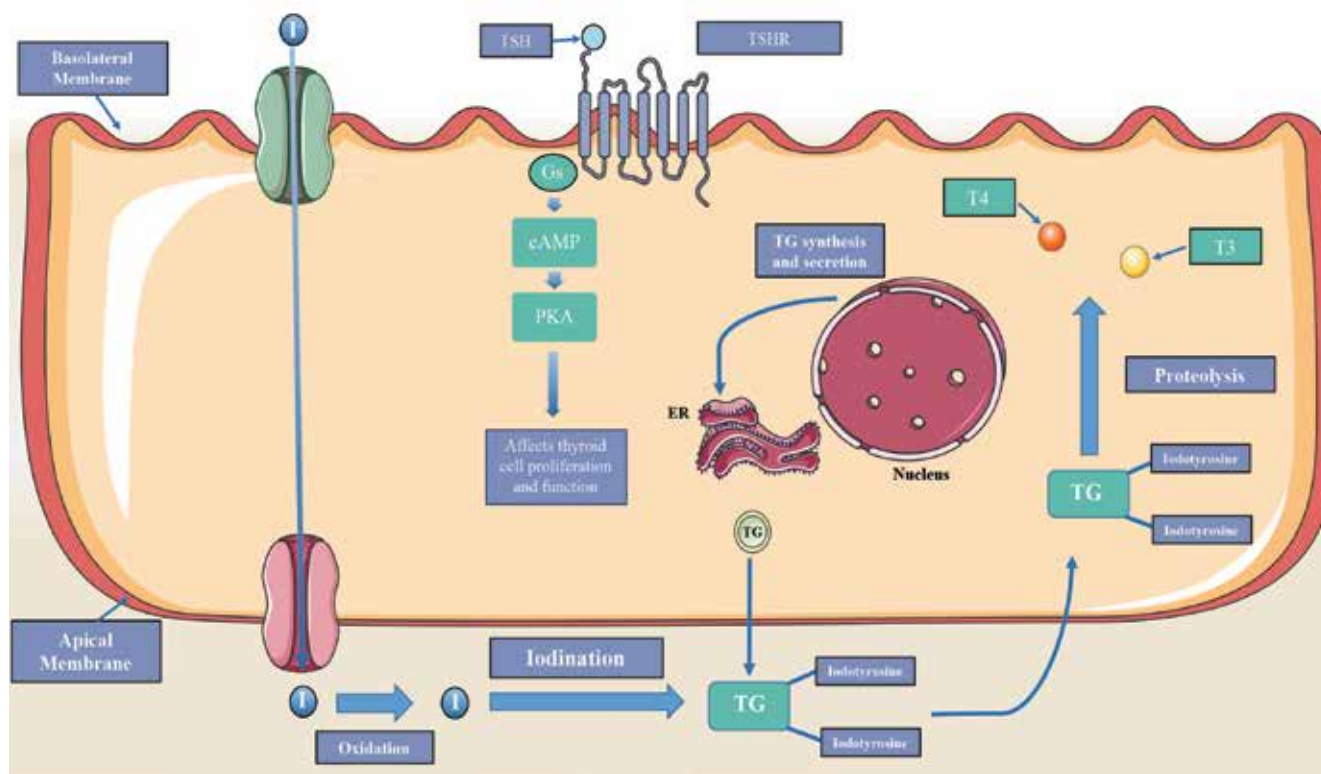


Figure 4. Thyroid hormone signaling

*Follicular cells contain thyroid stimulating hormone receptors (TSHR). Once bound to the hormone TSH, this G-protein coupled receptor generates the second messenger cyclic AMP. This second messenger activates protein kinases that trigger signal cascades responsible for thyroid cell proliferation and function. TSH stimulation of TSHR also leads to iodide transport into the cell via the sodium iodide symporter. This iodide is oxidized and then bound to tyrosine residues on thyroglobulin (TG), a process catalyzed by thyroperoxidase. This primed form of TG can then be cleaved through proteolysis in the cell to generate the thyroid hormones T3 and T4

proliferation, sensitization to TSH, and inhibiting apoptosis, but it is not sufficient to transform thyroid cells by itself (19,20).

This alteration in *AKT1* occurs in about 15% of metastatic thyroid cancers, while also occurring in small fractions of ATCs and FTCs. Studies have shown that there is an association between *AKT1* activation and tumor aggressiveness. This association with metastasis in thyroid carcinoma was found in both PTC and FTC. However, the aggressiveness seen is likely attributed to a combination of genetic alterations rather than solely on dysregulation of *AKT1* activity (19,20,21).

TERT

The *TERT* gene, located on chromosome 5p15.33, encodes the protein telomerase reverse transcriptase. It is the catalytic subunit of the telomerase enzyme and determines telomerase activity. The *TERT* subunit associates with a RNA component TERC as well as additional components to form the telomerase enzyme. Telomerase serves an important role in synthesizing and maintaining telomeres, which helps prolong the life of cells in the body. When telomeres become too short, apoptosis is triggered in the cell. As a result, telomerase activity is highly regulated and expressed at very low levels in most tissues in the body (22).

Dysregulation of telomerase is an almost universal feature of cancer, with over 90% exhibiting overexpression of the enzyme. This imparts cancer cells with an infinite capability to divide. The up-regulation of *TERT* that is seen could be the result of epigenetic deregulation or genetic amplification of the gene's locus (Figure 3). Other factors contributing to the dysregulation of *TERT* expression involve changes in transcription factors that target the *TERT* gene. Such factors include *TP53* and the ETS transcription factors. Both are disrupted in thyroid carcinoma as *TP53* is often mutated and ETS transcription factors are stimulated by several oncogenes including *RAS*. These associations indicate a relationship between an up-regulation of *TERT* expression and the transformation of thyroid cells. One recently discovered mechanism has been mutations in the *TERT* promoter, the most important regulatory element of telomerase activity (22,23).

These promoter mutations, C228T and C250T, seem to increase the transcription of *TERT*, likely through the creation of consensus binding sites for ETS transcription factors. Promoter mutation C228T is the most common one found in thyroid carcinoma with no overlaps reported in the mutations. There seems to be a higher prevalence of *TERT* promoter mutations in more advanced forms of thyroid carcinoma like ATC. Studies have shown a strong association between *TERT* promoter mutations and *B-RAF* or *RAS* mutations as well. The evidence uncovered so far shows that mutations in the *TERT* promoter seems to represent a late stage genetic event in tumorigenesis as

well as an indicator of aggressiveness given its prevalence in ATC (24,25).

GNAS

The *GNAS* gene provides instructions for the stimulatory α -subunit of heterotrimeric G protein complexes. The gene is located on chromosome 20q13.3. This subunit stimulates the activity of adenylate cyclase and controls the production of several hormones that help regulate the activity of the endocrine glands such as the thyroid gland. Mutations in *GNAS* are often detected at codons 201 and 227. These mutations result in the constitutive activation by impairing GTP hydrolysis. This in turn results in the constitutive activation of the cyclic AMP (cAMP) cascade. The increase in cAMP levels results in thyroid cell proliferation and increased thyroid hormone production. Despite this disruption in function, these mutations are almost never malignant (20,26).

Activating mutations of *GNAS* has been shown to overactivate thyroid stimulating hormone receptor (*TSHR*) signaling. This often leads to benign, hyperfunctional follicular thyroid adenomas (FTAs). Mutation of *GNAS* is also the main cause for autonomously functioning thyroid nodules. Studies show that mutations in *GNAS* resulting in overactivation of the G protein alpha subunit leads to a hyperfunctioning thyroid. Serving as a potential marker for benign thyroid nodules, *GNAS* does not seem to serve as a driver of thyroid carcinogenesis. However, it seems likely that this disruption in thyroid function in conjunction with other genetic alterations plays a role in thyroid carcinoma (26).

PIK3CA

PIK3CA, phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha, is a gene located on chromosome 3q26.3. It encodes the p110 alpha subunit of the PI3K kinase. This particular subunit is the catalytic subunit and is responsible for the kinase function of the protein. PI3K plays a very important role in the PI3K/AKT signaling pathway, one of the major pathways implicated in thyroid carcinoma. As part of this signaling pathway, PI3K helps regulate cell proliferation, motility, adhesion, and cell cycle progression. The pathway begins with signals transmitted from the environment to the inside of the cell through RTKs. These RTKs then engage PI3K, recruiting the protein to the cell membrane and generating phosphatidylinositol-3,4,5-triphosphate (PIP3). PIP3 acts as a second messenger, recruiting protein kinases phosphoinositide-dependent protein kinase 1 (PDK1) and AKT to the membrane. Once activated via phosphorylation, these kinases target their downstream effectors, producing concomitant changes to cell growth and differentiation. This pathway is regulated by the *PTEN* lipid phosphatase, which dephosphorylates PIP3 and thus shuts off the PI3K/AKT pathway (27).

The PI3K pathway is frequently activated in thyroid carcinoma and studies have reported the overexpression of *PIK3CA* in FTC and ATC. The most common *PIK3CA* mutations observed with oncogenic effects tend to cluster around exons 9 and 20. These mainly somatic mutations result in the constitutive activation of PI3K. Consequently, this leads to overactivation of the PI3K/AKT pathway and thyroid tumorigenesis. Genetic alterations have been found in all differentiated stages of thyroid carcinoma. However, it seems to be the most frequent in ATC, occurring in about 24% of ATC cases. Given the occurrence of *PIK3CA* mutations in all stages of thyroid carcinoma, it supports the implication of the PI3K/AKT signaling pathway in both the initiation and progressive dedifferentiation of the disease (27,28,29).

PTEN

PTEN, phosphatase and tensin homolog, encodes a phosphatase enzyme found in almost all tissues in the body. Located on chromosome 10q23.3, *PTEN* plays an important role in the PI3K/AKT pathway. Once activated via phosphorylation, *PTEN* works to terminate the signaling pathway through its phosphatase activity. It converts PIP3 into PIP2 and shuts off the PI3K/AKT pathway and reduces the recruitment of kinases PDK1 and AKT. As a result, this protein helps regulate cell proliferation and differentiation (28,29).

A tumor suppressor gene, *PTEN* is found mutated frequently in thyroid tumors. Point mutations, deletions, and promoter methylation in *PTEN* have been reported in thyroid carcinoma, particularly undifferentiated ones like ATC. Overall, studies have shown that *PTEN* inactivation is a critical step in thyroid tumor progression. This is evident in the fact that *PTEN* is downregulated or absent in highly malignant thyroid carcinoma like ATC, though mutations have been found in more differentiated forms. *PTEN* downregulation has been reported in about 37% of FTCs. Ultimately, this inactivation results in the overactivation of the PI3K/AKT signaling pathway, resulting in rampant cell proliferation (28,29,30).

PTEN mutations tend to be most frequent in ATC, with an occurrence in about 15% of cases. Germline mutations of *PTEN* is also a leading cause of CS, a disorder that can result in benign and malignant tumors. Individuals with CS have a 10% risk of developing thyroid cancer. This association between CS and *PTEN* further supports its role in thyroid tumorigenesis (27,29,30,31).

TSHR

The *TSHR* gene, located on chromosome 14q31, encodes the *TSHR*. This G protein-coupled receptor spans the membrane of follicular cells in the thyroid. Once activated by thyroid stimulating hormone (TSH), *TSHR* produces

corresponding effects in the cell via second messengers like cAMP. This intracellular signaling plays an important role in thyroid cell proliferation and maintenance of thyroid function (Figure 4). Deregulation of *TSHR* seems to play an important role in thyroid carcinogenesis (20).

Studies have shown an association between high levels of TSH and the development of thyroid nodules. The overactivation of *TSHR*, whether through activating mutations in *TSHR* or *GNAS*, has also been shown to lead to hyperfunctional FTA. However, these tumors are rarely malignant, suggesting that *TSHR* could serve a dichotomous role - protecting against malignancy of thyroid tissue but also promoting carcinogenesis likely when activated by other oncogenic alterations. Mouse models have shown that *TSHR* is required for thyroid carcinogenesis. It is still uncertain whether this supports *TSHR*'s role in the initiation of thyroid carcinoma or it is simply due to the *TSHR*-dependent generation of thyroid cells from which carcinogenesis initiates (20,32,33).

Epigenetics has shown to play an integral part in the role of *TSHR* in thyroid carcinoma. Hypermethylation of the *TSHR* promoter has been frequently found in thyroid carcinoma, while it is unmethylated in normal and benign thyroid tumors. This epigenetic silencing is most prevalent in PTC, with a frequency of 34-59%. Epigenetic silencing via hypermethylation is also prevalent in more undifferentiated forms of thyroid carcinoma. Studies have also shown a strong association with *B-RAF* mutations in PTC. This relationship supports the theory that *TSHR* silencing is a secondary genetic event in thyroid carcinogenesis and not essential for the initiation of thyroid carcinoma (32,33).

The silencing of *TSHR* not only contributes to the development of thyroid carcinoma, but it desensitizes the thyroid tumors to radioiodine therapy through its interaction with sodium iodide symporter (NIS) in iodide uptake. The suppression of *TSHR* hinders the cancer cells' ability to concentrate iodine. Though no silencing mutations have been found thus far, it seems that methylation of *TSHR* serves as a marker for thyroid malignancy (33).

TG

The *TG* gene located on chromosome 8q24, encodes the prohormone *TG*. This protein, found in thyroid follicular cells, acts as a precursor for the thyroid hormones, serving as the matrix from which they are produced. It acts as a protein storehouse and when the thyroid hormones are needed, *TG* is altered and broken down to release the needed hormones. Tyrosine residues on the *TG* protein are iodinated, priming the protein for modification into the thyroid hormones T3 and T4 (Figure 4). The synthesis of *TG* is regulated by TSH, with TSH serving to stimulate production (34).

Mutations in *TG* have been reported in thyroid carcinoma. They tend to be more frequently found in differentiated

thyroid carcinomas, like PTC. The majority of PTCs with a mutation in *TG* tend to also carry a mutation in one of the genes in the MAPK signaling pathway. Though rarely mutated, *TG* does seem to contribute to the malignancy of the cancer. As the cancer dedifferentiates, the expression of *TG* seems to decrease, with it becoming absent in ATC. These genetic changes do not seem to be sufficient to initiate thyroid carcinogenesis, but the hormonal imbalance produced seems to aid in its progression (34,35).

Changes in *TG* are more commonly associated with dys-hormonogenesis, occurring with a frequency of 25%. Individuals with the disorder tend to develop goiters, often due to chronic stimulation by TSH. Studies have shown that individuals with these dys-hormonogenic goiters and *TG* mutations have an increased risk of developing aggressive forms of thyroid carcinoma.

TG also serves as a tool for monitoring thyroid cancer recurrence and metastasis after treatment and thyroidectomy. Serum *TG* levels are carefully monitored and upon stimulation by TSH, if levels rise too high it serves as a strong indicator of metastases. Such an association helps support the role of *TG* dysregulation in the development of thyroid carcinoma (35,36,37,38).

PAX8/PPAR γ

A chromosomal rearrangement found in thyroid carcinoma is that between the genes *PAX8* and *PPAR γ* , *PAX8* encodes a transcription factor that is essential for normal thyroid development. The *PPAR γ* gene encodes a nuclear receptor part of the PPAR subfamily. This receptor forms heterodimers with retinoid X receptors and helps regulate the transcription of several genes, playing a major role in adipogenesis and lipid metabolism (3,39).

This rearrangement is the result of a translocation between chromosome regions 2q13 and 3p25. Highly expressed in thyroid follicular cells, it seems this fusion protein plays a role in thyroid tumorigenesis. The exact mechanism is still unknown. Some studies show that *PAX8/PPAR γ* has a dominant negative effect on wild type *PPAR γ* , which is believed to act as a tumor suppressor. The fusion protein contains the transcriptional regulatory domains of both proteins. As a result, it is capable of modulating their respective downstream pathways, usually targeting *PAX8*-responsive promoters (39,40).

PAX8/PPAR γ is most frequently found in FTCs, with an occurrence rate of about 30%, and occurs rarely in the follicular variant of PTC. The presence of this rearrangement has no major impact on the prognosis of thyroid carcinoma. Though still much remains unknown about this chromosomal rearrangement, evidence points to its potential as an oncoprotein and genetic driver of thyroid carcinoma (4,39,40).

IGF2BP3/IMP3

The *IGF2BP3* gene is located on chromosome 7p11.2, and encodes for the insulin-like growth factor-2 (IGF-2) mRNA-binding protein 3. *IGF2BP3* is a RNA-binding protein consisting of two RNA-recognition motifs in its N-terminal part and four KH homology domains in the C-terminal region. Its primary function is as a translational activator of the IGF-2 gene, while also functioning as a post-transcriptional regulator of cell proliferation, and differentiation during embryogenesis. It is important to note that *IGF2BP3* is a member of the highly conserved IGF2BP family of mRNA-binding proteins along with IGF2BP1 and IGF2BP2. As a member of the IGF2BP family *IGF2BP3* shares a high amount of similarity to the other three members at the amino acid level but differs in terms of its expression. *IGF2BP3* is abundantly found in all tissues during embryonic stage, but its expression in normal mature tissue is confined to the placenta and reproductive tissue (41,42,43).

IGF2BP3 has been the member of the IGF2BP family to be most associated with various cancer types such as lung, ovarian, pancreatic, and colorectal. Its role in tumor progression has been suggested to be by promoting cell growth, proliferation and resistance to ionic radiation via an IGF-2 dependent manner. Recent studies have found *IGF2BP3* to play a role in increasing the invasive potential of tumor cells *in vitro* (42,43).

Due to the high transcript levels of *IGF2BP3* in neoplastic cells and near undetectable presence in most adult tissue, it's been suggested as a potential biomarker. Through the use of immunoassays, its expression has been exclusively observed in malignant thyroid cancers of follicular origin, with a small portion of classical PTC, and was observed in 59% of cases for poorly differentiated thyroid carcinoma variants. However, through the use of reverse transcription polymerase chain reaction (RT-PCR) and qRT-PCR assay, it was demonstrated that *IGF2BP3* was overexpressed in thyroid carcinoma tissue when compared to benign thyroid lesions. *IGF2BP3*'s overexpression in thyroid carcinoma, its near undetectable levels in adult tissue, and its potential to distinguish between benign and malignant tissue make it an excellent candidate for the thyroid cancer sequencing panel (41,44,45,46).

KRT7/KRT20

The *KRT7* and *KRT20* genes both belong to the keratin gene family, which consists of 54 functional genes. The *KRT7* gene can be found in type 2 cluster on chromosome 12, while the *KRT20* gene is located in the type 1 cluster on chromosome 17. The protein products of these genes are subdivided into type 1 and type 2 based on their molecular weights. *KRT7* and *KRT 20* both encode for cytokeratin proteins that are intermediate filament forming proteins exclusively found in epithelial tissue. The filaments

are formed from the heterodimer pairing of type 1 and type 2 cytokeratin proteins in equimolar amounts. The intermediate filaments created by cytokeratin proteins aid in forming the structural framework of epithelial cells, regulate cell size, growth, division, as well as protection against mechanical stressors (46,47).

Adenocarcinomas comprise the largest group of human epithelial malignancies and can occur in different organs. *KRT7* and *KRT20* along with the other members of the keratin gene family have been utilized as diagnostic markers to determine the origin tissue, due to the characteristic cell-type, differentiation and functional status-dependent expression pattern of cytokeratin in epithelial cells. In most adenocarcinomas, the expression levels of *KRT7* and *KRT20* are variable and thus are often screened for together (47).

Current literature on *KRT7/KRT20* combined screening of thyroid neoplasms is scarce. Almost all thyroidal tumor's expression levels for *KRT7* and *KRT20* have been found to be positive and negative, respectively. In fact, one study looked into the immunohistochemical profile of 43 different types of primary and metastatic thyroid neoplasms. They found that excluding anaplastic carcinomas, 79% of thyroid tumors and 94% of metastatic thyroid carcinoma cases reacted positively with the antibody. While the study found a positive result for *KRT7*, none of the thyroid tumors or their metastases reacted at all to the *KRT20* antibody. A second retrospective study examined 153 thyroid carcinoma samples for *KRT7/KRT20* immunohistochemically, and they found that all papillary carcinomas, follicular carcinomas, and medullary carcinomas were *KRT7* positive and *KRT20* negative. Due to the fact that *KRT7* has been positively stained in almost all thyroid neoplasms while *KRT20* has consistently been undetectable, *KRT7* and *KRT20* are excellent candidates for the thyroid cancer sequencing panel (48,49,50).

TP53

The gene *TP53* encodes tumor protein p53, located on chromosome 17p13.1. This protein is a nuclear transcription factor that functions as a tumor suppressor. P53 contains three functional domains: a transactivation domain, a DNA binding domain, and an oligomerization domain. In the nucleus, it binds directly to DNA in order to help prevent mutated or damaged DNA from dividing. The protein either activates other genes to fix any damages or triggers cell cycle arrest and apoptosis (Figure 5). As a result, this protein plays a crucial role in regulating cell division and preventing tumor formation (51).

Any impairment in the function of *TP53* can lead to destabilization of the genome and an accumulation of mutations. Most changes involve point mutations in exons 5-8, deletions, or inactivation through indirect mechanisms such as methylation. The now inactivated *TP53* protein

becomes unable to bind to DNA and thus is unable to repair damages that arise in the DNA. The protein, as well as damages to the DNA, just accumulates in the nucleus. These type of changes are usually restricted to poorly differentiated thyroid cancers like ATC, reportedly occurring in 60% of cases (8,51).

TP53 exhibits tremendous oncogenic potential, as it is mutated in about 50% of all human cancers. However, inactivating mutations of *TP53* are found in only about 10% of thyroid carcinomas overall. In well differentiated thyroid carcinomas, *TP53* mutations have a reported relevance ranging from 0 to 25%. Though it seems other mechanisms of *TP53* inactivation play a role in thyroid carcinogenesis, as p53 protein is more commonly overexpressed with a frequency of 11-59% (51,52).

CALCA

CALCA (*CALCA-1*) gene is a member of the calcitonin gene family consisting of four known genes, *CALCA-1*, *CALCA-2*, *CALCA-3*, and *CALCA-4* all containing nucleotide sequence homologies. The *CALCA* gene located on chromosome 11 encodes for two separate peptide hormones. This gene consists of six exons in which exon 4 and 5 contain the sequences for the calcitonin and calcitonin gene related peptide (α -CGRP), respectively. Post-transcriptional processing of the *CALCA* gene in thyroid C cells and neural tissue results in tissue specific production of calcitonin and α -CGRP mRNAs. CT is a 32 amino-acid peptide hormone with tissue specific functional roles in the central nervous, skeletal, renal and gastrointestinal systems. However, CT's major role is in calcium and phosphate metabolism. CT modulates calcium levels by binding with CT receptors to inhibit osteoclast motility. All of CT's activities occur through its binding to the calcitonin receptor, which couples its binding to the activation of cAMP and PKC signaling pathways (53,54,55).

Due to the uncharacteristically high concentrations of calcitonin, serum CT is classically used as a diagnostic biomarker for MTC. Studies have demonstrated that preoperative serum CT levels can be strong predictors for tumor size and remission. Preoperative CT serum levels above 1000 pg/mL correspond to 25.0 mm tumor sizes, while levels below 100 pg/mL were associated with 3.0 mm tumor size. Distant metastases are the main cause of death in patients diagnosed with MTC. Metastasis is initially presented in 50% of MTC cases. Postoperative serum CT levels are an important procedure in screening for biochemical remission. CT doubling times have been found to be associated with progressive or stable disease in 80% of patients. In a study they found doubling times shorter than 25 months to have a 94% predictive value of progressive disease along with a predictive value of 86% for patients with a doubling time longer than 24 months (53,56,57,58,59).

CALCA detection studies and procedures involve detection of this peptide from blood serum. There are limitations to these assays due to needing stimulation tests, and possible intra-assay variation. Some research has found that detection of CALCA gene transcripts presented higher clinical sensitivity, specificity, and positive predictive values of 86.67, 97.06, and 92.86%, respectively. In addition to these promising results their readings mirrored measurements obtainable through a pentagastrin stimulation test. Due to its tissue specific production and role in preoperative and postoperative diagnosing of MTC, CALCA is a viable candidate to include in the thyroid cancer mutation panel (53,60).

MET

MET, also known as c-MET, encodes a proto-oncogene that is located on chromosome 7q31. A member of the RTK family, the MET receptor elicits its effects by binding to its ligand, hepatocyte growth factor (HGF). Once bound to HGF, the MET receptor dimerizes and thus becomes activated and helps promote cell proliferation, survival, and

motility. These effects are largely the result of activation of MAPK and PI3K pathways through MET signaling. This process is tightly regulated and any disruption can result in tumorigenesis (61).

In thyroid carcinoma, what is most commonly seen is a deregulation of MET, most commonly in PTC. This deregulation often involves overactivation or overexpression of MET, which has been shown to promote tumor growth. This activation can be the result of activating point mutations often located in the tyrosine kinase domain, or paracrine signaling due to increased sensitivity to HGF. The altered MET gene acts as a mitogenic factor and helps promote cell motility and invasion (61,62).

Expression levels of MET tend to be low and tightly regulated in normal thyroids. However, it is amplified in 75% of PTCs, as well as in many FTCs. This overexpression seems to be secondary to other driver mutations like RAS and B-RAF. Despite its secondary role, evidence points to increased MET signaling having a significant role in the pathogenesis of thyroid carcinoma (62,63,64).

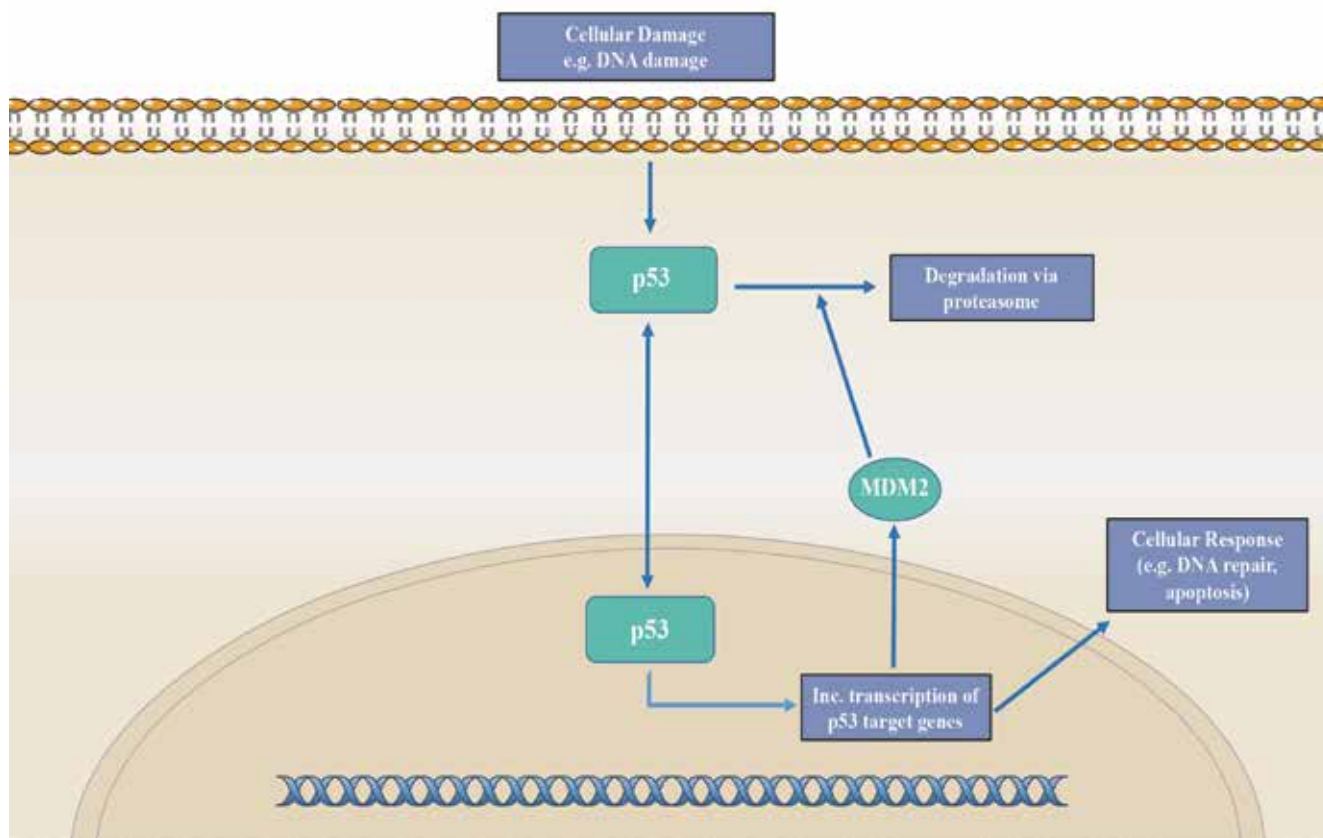


Figure 5. P53 pathway

*Signals of cellular stress, such as DNA damage, triggers activation of p53. Once activated, this nuclear transcription factor travels to the nucleus and binds to DNA, increasing expression of target genes involved in DNA repair and triggering apoptosis. p53 activity also induces expression of MDM2 as well. MDM2 serves to regulate p53 activity, inhibiting transcription of p53 and promoting its degradation. Overall, this pathway functions as an auto-regulatory feedback loop

EIF1AX

This X-linked gene, located on Xp22.12, encodes the eukaryotic translation initiation factor 1A. This particular protein is required for the formation of the preinitiation complex (PIC). It does so by stabilizing the binding of the initiator tRNA (*Met*-tRNA) with the 40S ribosomal subunit and the translation initiation factor eIF2. This is an integral step in the translation of mRNA into proteins, as it is required for the binding of the PIC to the 5' end of capped mRNA and finding the proper start codon (65,66).

Mutations in *EIF1AX* have been found in both PTC and ATC. The most common sites of mutation are the N-terminal domain and a C-terminal splice acceptor site between exons 5 and 6. The C-terminal mutations seem to be specific to thyroid carcinoma. In PTC, it is rare and occurs in a mutually exclusive manner, found in only 1-2% of tumors. But, it is more frequent in ATC, found in 10% of tumors. It seems that these mutations result in an overactive *EIF1AX*, as it has been shown to increase cell proliferation (65,66,67,68).

EIF1AX mutations tend to coexist with *RAS* mutations and other driver mutations of thyroid carcinoma like *TP53*. This co-occurrence tends to be found predominantly in poorly differentiated thyroid carcinomas like ATC. This strong association suggests that *EIF1AX* is not sufficient enough to transform thyroid carcinoma into a malignant form. Mutations in *EIF1AX* have also been associated with poor prognosis in PDCs. The mechanisms of *EIF1AX* mutation in thyroid carcinoma are still unclear, especially in terms of its association with *RAS* mutations (65,66,67,68).

TTF1

Also known as NK2 homeobox 1, this gene encodes thyroid transcription factor 1. The *TTF1* protein is expressed in the epithelial cells of the thyroid with a heterogeneous distribution. As a homeoprotein, it plays an important role in thyroid development, cell growth, and differentiation. Essential for thyroid organogenesis, *TTF1* helps modulate thyroid function through its regulation of gene expression in the thyroid, activating the transcription of thyroid-specific genes like thyroglobulin (*TG*) and thyroperoxidase (*TPO*) (69,70).

The mechanisms by which *TTF1* contributes to thyroid carcinoma still remains unclear. Though the gene is expressed in thyroid carcinoma, its level of expression becomes reduced as cell dedifferentiation progresses. This change in expression could be the result of epigenetic silencing, likely achieved through hypermethylation and histone H3 modification. As a result, ATC exhibits the lowest levels of *TTF1*. In thyroid carcinoma as a whole it seems likely that *TTF1*'s molecular functions, DNA binding, and transcription activation are probably disrupted. That could explain why most undifferentiated thyroid carcinomas

have little to no *TG* or *TPO* proteins, both targets of *TTF1* transcription factor activity (41,42,43).

Germline mutations in *TTF1* have also been shown to play a role in thyroid carcinoma, specifically in PTC. These mutations seem to promote cell proliferation as well as activate signaling pathways like PI3K/AKT. The germline mutations observed in *TTF1* have also been associated with multinodular goiter. This provides further evidence of a progression from a benign multinodular goiter to malignant thyroid carcinoma, such as PTC. Given the important role *TTF1* plays in maintaining thyroid architecture and function, it is not surprising that any abnormalities in the gene and its product contribute to the development of thyroid carcinoma (70,71,72).

PTH

The *PTH* gene encodes a member of the parathyroid family of proteins and is located on chromosome 11p15.3. This particular protein binds to the parathyroid hormone receptor and helps regulate blood calcium and phosphate levels in the body. Abnormal levels of *PTH* tend to be associated with parathyroid diseases, like hyperparathyroidism. However, there is evidence to suggest a possible link between *PTH* and the development of thyroid carcinoma (73).

Primary hyperparathyroidism (PHPT) is a disease that has been associated with thyroid carcinoma. PHPT results from an excessive secretion of *PTH*, which causes an elevation of calcium in the blood. The most common form of thyroid carcinoma associated with PHPT is a hereditary form of MTC known as multiple endocrine neoplasia type 2A (MEN2A). Carriers of the MEN2A gene with a mutation at codon 634 concomitantly suffer from PHPT with a frequency of 10-25%. Such an association provides evidence to support the role of *PTH* dysregulation in thyroid carcinoma (73,74).

Associations between non-medullary thyroid carcinoma and PHPT have also been reported. With rates ranging from 2-10%, thyroid malignancy seems to coexist with PHPT, albeit not frequently. A strong relationship has also been found between ionizing radiation and the simultaneous development of PHPT and thyroid carcinoma. *PTH* may also contribute to the development of thyroid carcinoma through chromosomal rearrangement with *PAD1*. This rearrangement results in an overexpression of cyclin D1 and thus an increase in cell proliferation. Though some changes to *PTH* may be the result of metastatic invasion of the parathyroid by thyroid carcinoma, the evidence available shows that the role of *PTH* in thyroid malignancy overall warrants further investigation (75,76,77).

SCL5A5

The *SCL5A5* gene encodes the NIS. NIS is a glycosylated protein with 13 trans-membrane domains and functions

to actively transport one I⁻ and two Na⁺ ions from the sodium ion gradient established by the Na⁺/K⁺ ATPase. NIS expression at both the transcriptional level and posttranscriptional level is modulated by the effects of TSH. TSH causes elevated levels of endogenous cAMP in the thyroid cells. These cAMP levels play a role in stimulating several cis-regulatory elements found in the promoter and upstream enhancer sequences on the *SCL5A5* gene on chromosome 19 (78,79).

The *SCL5A5* gene contains an upstream enhancer sequence (NUE) reported to have both a Pax-8, and cAMP-response element (CRE) binding sites. Pax-8 and cAMP-response element modulator (CREM) transcription factors bind to these cis-elements on the upstream enhancer via cAMP-signaling pathways. These elements are stimulated through cAMP activation of protein kinase-A (PKA) dependent pathways in thyroid cells. PKA dependent activation phosphorylates basic-leucine zipper (B-ZIP) proteins,

CREM, activating transcription factor-1 (ATF-1), and cAMP-response element binding protein (CREB). Studies have found that the binding of these transcription factors are necessary for the NUE's cAMP induced activity on thyroidal responsive genes such as TPO, *TG*, *TSHR*, and NIS (Figure 6). While CREB binding is important for *SCL5A5* transcriptional regulation, full *SCL5A5* regulation is not established without PAX-8 binding. TSH binding increases the expression and translocation of the anti-oxidative stress factor Ref-1. Ref-1 is a nuclear enzyme that reduces Pax-8 and mediates its binding to the NUE sequence. Impaired translocation of this nuclear enzyme has been reported in anaplastic, papillary cell lines as well as in thyroid cancer tissue. TSH also plays a role in the posttranscriptional regulatory role in the transport of NIS to the basolateral membrane (78,79).

While no somatic mutations to NIS have yet been identified, alterations in other genes have been found to be associated with NIS impairment in differentiated thyroid cancers.

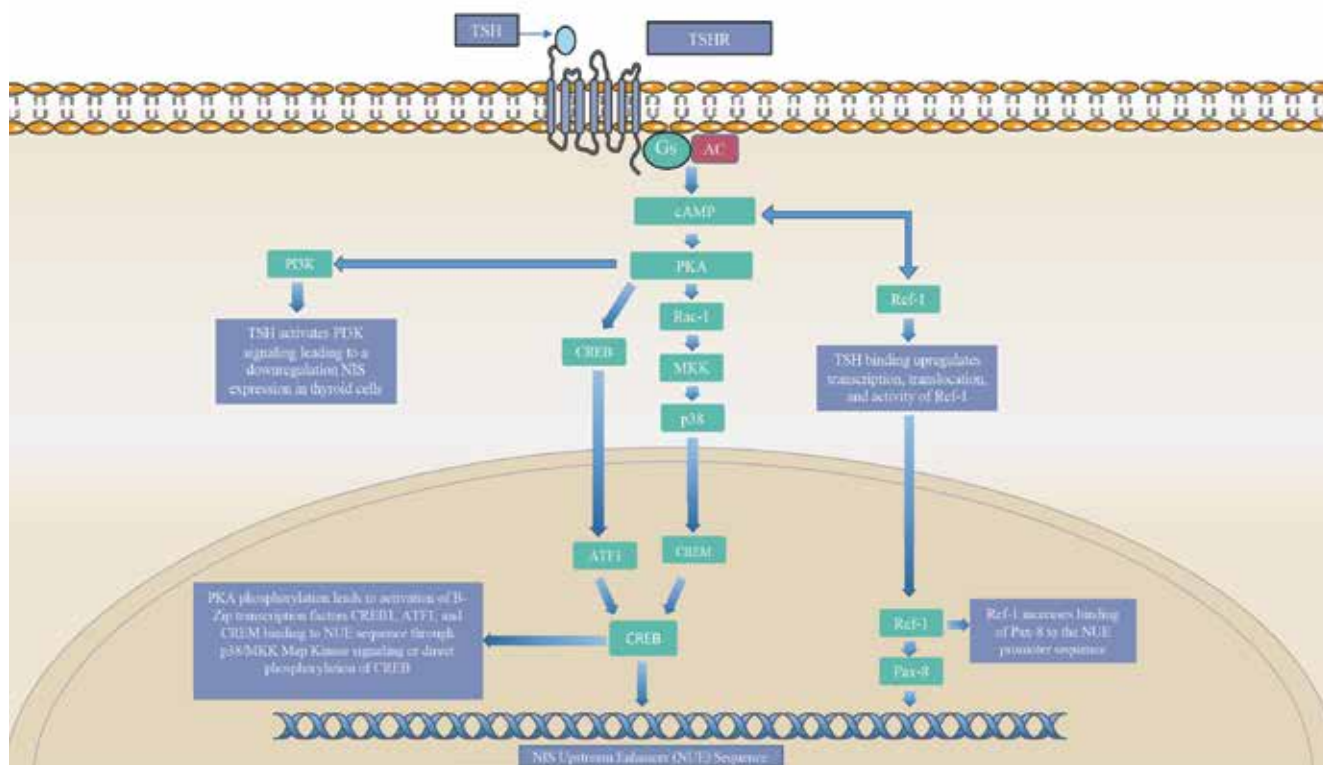


Figure 6. Sodium iodide symporter gene signaling

*The sodium iodide symporter is a transmembrane protein that plays a crucial role in the uptake of iodine by thyroid cells. The *SCL5A5* gene is regulated by the activity of TSH on the sodium iodide symporter upstream enhancer sequence (NUE). In this pathway TSH binding to TSHR stimulates an increase in cyclic AMP (cAMP) levels in the cell. Elevated levels of cAMP lead to the activation of protein kinase-A (PKA) dependent p38/MAPK and PKA-cAMP-response element binding protein (CREB) pathways. The p38/MAPK pathway leads to the recruitment of basic-leucine zipper transcription factors ATF1, and cAMP-response modulator to the nucleus. In the nucleus these transcription factors bind to the cAMP-response element binding sites in the NUE and activating the *SCL5A5* gene. PKA can also directly phosphorylate the cAMP-CREB to directly regulate cAMP responsive genes such as *TSHR*, and *SCL5A5*. Full activation of the *SCL5A5* gene is not reached until the PAX-8 transcription factor binds to the NUE promoter site. TSH signaling upregulates the transcription and nuclear translocation of the oxidative stress factor Ref-1. Ref-1 activates and recruits PAX-8 to bind to the NUE promoter site, leading to full *SCL5A5* gene activity. Inhibition of *SCL5A5* activity can also occur due to the elevated levels of cAMP. These elevated levels can trigger activation of the PI3K signaling pathway leading to the downregulation of the *SCL5A5* gene

B-RAF^{V600E} mutations reduce NIS mRNA expression by inducing the secretion of transforming growth factor- β (TGF- β). TGF- β acts via the SMAD signaling pathway in an MEK/ERK independent pathway. *RET*/*PTC* rearrangement impairs the activity of PAX-8 and PKA. Reducing the activity of these two proteins has been demonstrated to reduce recruitment of B-ZIP proteins and reduced binding to the PAX-8 and CRE elements in NUE (79,80,81).

NIS expression has been reported to be reduced in differentiated thyroid carcinomas. One study found reduced NIS mRNA expression in PTC tissue samples when compared to adjacent normal thyroid tissue. NIS expression was also found to be generally reduced in benign thyroid nodules compared with adjacent normal tissue, but to a lesser extent than in PTC tissue samples. It has also been reported that NIS gene expression is impaired in hypo-functioning thyroid tumors when compared to normal thyroid tissue. Quantification of *SCL5A5* mRNA levels should be added to the classical prognostic factors currently used to predict the outcomes of patients with differentiated thyroid cancer. *SCL5A5* is an excellent candidate for the thyroid cancer mutation panel (82,83,84).

Conclusion

Thyroid carcinoma is the result of an accumulation of genetic changes that impact its initiation and progression. These genetic alterations, thus, serve as both diagnostic and prognostic markers of thyroid carcinoma. ThyroSeq NGS allows for the use of small amounts of DNA to test for a very broad range of mutations and other genetic alterations, helping identify molecular profiles for the different types of thyroid carcinoma. The genotyping of thyroid samples provides invaluable insight in determining the proper course of treatment when faced with a cytologically indeterminate thyroid nodule.

The genetics of thyroid cancer provides us with a greater understanding of the pathophysiology of the disease than is possible from its cellular morphology. The presence of mutations in the molecular pathology profile can not only help more accurately determine malignancy in the thyroid nodules but help better predict the biology of thyroid carcinoma which will have paramount importance to tailor treatment strategies. The presence of certain genetic alterations may prove indicators of aggressiveness of the cancer and predictors for metastasis. As more studies are conducted, and the genomic profile of thyroid carcinoma is further elucidated, the utility of genomic profiling in the diagnosis and prognostication of thyroid cancer will be solidified.

Authorship Contributions

Surgical and Medical Practices: Seza Gulec, *Concept:* Seza Gulec, Jorge Alsina, *Design:* Seza Gulec, Jorge Alsina, Raul Alsina, *Data Collection or Processing:* Jorge Alsina, Raul

Alsina, Seza Gulec, *Analysis or Interpretation:* Jorge Alsina, Raul Alsina, Seza Gulec, *Literature Search:* Jorge Alsina, Raul Alsina, Seza Gulec, *Writing:* Jorge Alsina, Raul Alsina, Seza Gulec.

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References

- Vigneri R, Malandrino P, Vigneri P. The changing epidemiology of thyroid cancer: why is incidence increasing? *Curr Opin Oncol* 2015;27:1-7.
- Nikiforov YE, Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. *Nat Rev Endocrinol* 2011;7:569-580.
- Nikiforova MN, Wald AI, Roy S, Durso MB, Nikiforov YE. Targeted next-generation sequencing panel (ThyroSeq) for detection of mutations in thyroid cancer. *J Clin Endocrinol Metab* 2013;98:E1852-1860.
- Nikiforov YE. Thyroid carcinoma: molecular pathways and therapeutic targets. *Mod Pathol* 2008;21(Suppl 2):S37-43.
- LiVolsi VA. Papillary thyroid carcinoma: an update. *Mod Pathol* 2011;24(Suppl 2):S1-9.
- Wan PT, Garnett MJ, Roe SM, Lee S, Niculescu-Duvaz D, Good VM, Jones CM, Marshall CJ, Springer CJ, Barford D, Marais R, Cancer Genome P. Mechanism of activation of the RAF-ERK signaling pathway by oncogenic mutations of B-RAF. *Cell* 2004;116:855-867.
- Howell GM, Hodak SP, Yip L. RAS mutations in thyroid cancer. *Oncologist* 2013;18:926-932.
- Patel KN, Singh B. Genetic considerations in thyroid cancer. *Cancer Control* 2006;13:111-118.
- Isaeva AV, Zima AP, Saprina TV, Kasoyan KT, Popov OS, Brynova OV, Berezkina IS, Vasil'eva OA, Ryazantseva NV, Shabalova IP, Litvinova LS, Pak YD, Novitskii VV. Comparative Evaluation of beta-Catenin and E-Cadherin Expression in Liquid Aspiration Biopsy Specimens of Thyroid Nodules. *Bull Exp Biol Med* 2016;161:288-291.
- Garcia-Rostan G, Camp RL, Herrero A, Carcangiu ML, Rimm DL, Tallini G. Beta-catenin dysregulation in thyroid neoplasms: down-regulation, aberrant nuclear expression, and CTNNB1 exon 3 mutations are markers for aggressive tumor phenotypes and poor prognosis. *Am J Pathol* 2001;158:987-996.
- Sastre-Perona A, Riesco-Eizaguirre G, Zaballos MA, Santisteban P. ss-catenin signaling is required for RAS-driven thyroid cancer through PI3K activation. *Oncotarget* 2016 doi: 10.18632/oncotarget.10356.
- Pierotti MA, Greco A. Oncogenic rearrangements of the NTRK1/NGF receptor. *Cancer Lett* 2006;232:90-98.
- Vaishnavi A, Le AT, Doebele RC. TRKING down an old oncogene in a new era of targeted therapy. *Cancer Discov* 2015;5:25-34.
- Greco A, Miranda C, Pierotti MA. Rearrangements of NTRK1 gene in papillary thyroid carcinoma. *Mol Cell Endocrinol* 2010;321:44-49.
- Lannon CL, Sorensen PH. ETV6-NTRK3: a chimeric protein tyrosine kinase with transformation activity in multiple cell lineages. *Semin Cancer Biol* 2005;15:215-223.
- Leeman-Neill RJ, Kelly LM, Liu P, Brenner AV, Little MP, Bogdanova TI, Evdokimova VN, Hatch M, Zurnadzy LY, Nikiforova MN, Yue NJ, Zhang M, Mabuchi K, Tronko MD, Nikiforov YE. ETV6-NTRK3 is a common chromosomal rearrangement in radiation-associated thyroid cancer. *Cancer* 2014;120:799-807.
- Giordano TJ, Kuick R, Thomas DG, Misek DE, Vinco M, Sanders D, Zhu Z, Ciampi R, Roh M, Shedden K, Gauger P, Doherty G, Thompson NW, Hanash S, Koenig RJ, Nikiforov YE. Molecular classification of papillary thyroid carcinoma: distinct BRAF, RAS, and RET/PTC mutation-specific gene expression profiles discovered by DNA microarray analysis. *Oncogene* 2005;24:6646-6656.

18. Marotta V, Guerra A, Sapio MR, Vitale M. RET/PTC rearrangement in benign and malignant thyroid diseases: a clinical standpoint. *Eur J Endocrinol* 2011;165:499-507.
19. Vasko V, Saji M, Hardy E, Kruhlak M, Larin A, Savchenko V, Miyakawa M, Isozaki O, Murakami H, Tsushima T, Burman KD, De Micco C, Ringel MD. Akt activation and localisation correlate with tumour invasion and oncogene expression in thyroid cancer. *J Med Genet* 2004;41:161-170.
20. Xing M. Molecular pathogenesis and mechanisms of thyroid cancer. *Nat Rev Cancer* 2013;13:184-199.
21. Ringel MD, Hayre N, Saito J, Saunier B, Schuppert F, Burch H, Bernet V, Burman KD, Kohn LD, Saji M. Overexpression and overactivation of Akt in thyroid carcinoma. *Cancer Res* 2001;61:6105-6111.
22. Martinez P, Blasco MA. Telomeric and extra-telomeric roles for telomerase and the telomere-binding proteins. *Nat Rev Cancer* 2011;11:161-176.
23. Heidenreich B, Rachakonda PS, Hemminki K, Kumar R. TERT promoter mutations in cancer development. *Curr Opin Genet Dev* 2014;24:30-37.
24. Liu X, Bishop J, Shan Y, Pai S, Liu D, Murugan AK, Sun H, El-Naggar AK, Xing M. Highly prevalent TERT promoter mutations in aggressive thyroid cancers. *Endocr Relat Cancer* 2013;20:603-610.
25. Landa I, Ganly I, Chan TA, Mitsutake N, Matsuse M, Ibrahimspasic T, Ghossein RA, Fagin JA. Frequent somatic TERT promoter mutations in thyroid cancer: higher prevalence in advanced forms of the disease. *J Clin Endocrinol Metab* 2013;98:E1562-1566.
26. Hsiao SJ, Nikiforov YE. Molecular approaches to thyroid cancer diagnosis. *Endocr Relat Cancer* 2014;21:T301-313.
27. Garcia-Rostan G, Costa AM, Pereira-Castro I, Salvatore G, Hernandez R, Hermsem MJ, Herrero A, Fusco A, Cameselle-Teijeiro J, Santoro M. Mutation of the PIK3CA gene in anaplastic thyroid cancer. *Cancer Res* 2005;65:10199-10207.
28. Guerra A, Di Crescenzo V, Garzi A, Cinelli M, Carlomagno C, Tonacchera M, Zeppa P, Vitale M. Genetic mutations in the treatment of anaplastic thyroid cancer: a systematic review. *BMC Surg* 2013;13 (Suppl 2):S44.
29. Nozhat Z, Hedayati M. PI3K/AKT Pathway and Its Mediators in Thyroid Carcinomas. *Mol Diagn Ther* 2016;20:13-26.
30. Smith JR, Marqusee E, Webb S, Nose V, Fishman SJ, Shamberger RC, Frates MC, Huang SA. Thyroid nodules and cancer in children with PTEN hamartoma tumor syndrome. *J Clin Endocrinol Metab* 2011;96:34-37.
31. Xu B, Ghossein R. Genomic Landscape of poorly Differentiated and Anaplastic Thyroid Carcinoma. *Endocr Pathol* 2016;27:205-212.
32. Kartal K, Onder S, Kosemehmetoglu K, Kilickap S, Tezel YG, Kaynaroglu V. Methylation status of TSHr in well-differentiated thyroid cancer by using cytologic material. *BMC Cancer* 2015;15:824.
33. Khan MS, Pandith AA, Masoodi SR, Wani KA, Ul Hussain M, Mudassar S. Epigenetic silencing of TSHR gene in thyroid cancer patients in relation to their BRAF V600E mutation status. *Endocrine* 2014;47:449-455.
34. Whitley RJ, Ain KB. Thyroglobulin: a specific serum marker for the management of thyroid carcinoma. *Clin Lab Med* 2004;24:29-47.
35. Alzahrani AS, Baitei EY, Zou M, Shi Y. Clinical case seminar: metastatic follicular thyroid carcinoma arising from congenital goiter as a result of a novel splice donor site mutation in the thyroglobulin gene. *J Clin Endocrinol Metab* 2006;91:740-746.
36. Francis Z, Schlumberger M. Serum thyroglobulin determination in thyroid cancer patients. *Best Pract Res Clin Endocrinol Metab* 2008;22:1039-1046.
37. Rubio IG, Medeiros-Neto G. Mutations of the thyroglobulin gene and its relevance to thyroid disorders. *Curr Opin Endocrinol Diabetes Obes* 2009;16:373-378.
38. Mazzaferri EL, Robbins RJ, Spencer CA, Braverman LE, Pacini F, Wartofsky L, Haugen BR, Sherman SI, Cooper DS, Braunstein GD, Lee S, Davies TF, Arafah BM, Ladenson PW, Pinchera A. A consensus report of the role of serum thyroglobulin as a monitoring method for low-risk patients with papillary thyroid carcinoma. *J Clin Endocrinol Metab* 2003;88:1433-1441.
39. Raman P, Koenig RJ. Pax-8-PPAR-gamma fusion protein in thyroid carcinoma. *Nat Rev Endocrinol* 2014;10:616-623.
40. Placzkowski KA, Reddi HV, Grebe SK, Eberhardt NL, McIver B. The Role of the PAX8/PPARgamma Fusion Oncogene in Thyroid Cancer. *PPAR Res* 2008;2008:672829.
41. Slosar M, Vohra P, Prasad M, Fischer A, Quinlan R, Khan A. Insulin-like growth factor mRNA binding protein 3 (IMP3) is differentially expressed in benign and malignant follicular patterned thyroid tumors. *Endocr Pathol* 2009;20:149-157.
42. Lederer M, Bley N, Schleifer C, Huttelmaier S. The role of the oncofetal IGF2 mRNA-binding protein 3 (IGF2BP3) in cancer. *Semin Cancer Biol* 2014;29:3-12.
43. Bell JL, Wachter K, Muhleck B, Pazaitis N, Kohn M, Lederer M, Huttelmaier S. Insulin-like growth factor 2 mRNA-binding proteins (IGF2BPs): post-transcriptional drivers of cancer progression? *Cell Mol Life Sci* 2013;70:2657-2675.
44. Findeis-Hosey JJ, Xu H. The use of insulin like-growth factor II messenger RNA binding protein-3 in diagnostic pathology. *Hum Pathol* 2011;42:303-314.
45. Jin L, Seys AR, Zhang S, Erickson-Johnson MR, Roth CW, Evers BR, Oliveira AM, Lloyd RV. Diagnostic utility of IMP3 expression in thyroid neoplasms: a quantitative RT-PCR study. *Diagn Mol Pathol* 2010;19:63-69.
46. Asioli S, Erickson LA, Righi A, Jin L, Volante M, Jenkins S, Papotti M, Bussolati G, Lloyd RV. Poorly differentiated carcinoma of the thyroid: validation of the Turin proposal and analysis of IMP3 expression. *Mod Pathol* 2010;23:1269-1278.
47. Karantza V. Keratins in health and cancer: more than mere epithelial cell markers. *Oncogene* 2011;30:127-138.
48. Lam KY, Lui MC, Lo CY. Cytokeratin expression profiles in thyroid carcinomas. *Eur J Surg Oncol* 2001;27:631-635.
49. Bejarano PA, Nikiforov YE, Swenson ES, Biddinger PW. Thyroid transcription factor-1, thyroglobulin, cytokeratin 7, and cytokeratin 20 in thyroid neoplasms. *Appl Immunohistochem Mol Morphol* 2000;8:189-194.
50. Weber T, Lacroix J, Weitz J, Anman K, Magener A, Holting T, Klar E, Herfarth C, von Knebel Doeberitz M. Expression of cytokeratin 20 in thyroid carcinomas and peripheral blood detected by reverse transcription polymerase chain reaction. *Br J Cancer* 2000;82:157-160.
51. Malaguarnera R, Vella V, Vigneri R, Frasca F. p53 family proteins in thyroid cancer. *Endocr Relat Cancer* 2007;14:43-60.
52. Morita N, Ikeda Y, Takami H. Clinical significance of p53 protein expression in papillary thyroid carcinoma. *World J Surg* 2008;32:2617-2622.
53. Schifter S. Expression of the calcitonin gene family in medullary thyroid carcinoma. *Peptides* 1997;18:307-317.
54. Masi L, Brandi ML. Calcitonin and calcitonin receptors. *Clin Cases Miner Bone Metab* 2007;4:117-122.
55. Pacini F, Castagna MG, Cipri C, Schlumberger M. Medullary thyroid carcinoma. *Clin Oncol (R Coll Radiol)* 2010;22:475-485.
56. Ball DW. Medullary thyroid cancer: therapeutic targets and molecular markers. *Curr Opin Oncol* 2007;19:18-23.
57. Laure Giraudet A, Al Ghulzan A, Auperin A, Leboulleux S, Chehboun A, Troalen F, Dromain C, Lumbroso J, Baudin E, Schlumberger M. Progression of medullary thyroid carcinoma: assessment with calcitonin and carcinoembryonic antigen doubling times. *Eur J Endocrinol* 2008;158:239-246.
58. Cohen R, Campos JM, Salaun C, Heshmati HM, Kraimps JL, Proye C, Sarfati E, Henry JF, Niccoli-Sire P, Modigliani E. Preoperative calcitonin levels are predictive of tumor size and postoperative calcitonin normalization in medullary thyroid carcinoma. *Groupe d'Etudes des Tumeurs a Calcitonine (GETC). J Clin Endocrinol Metab* 2000;85:919-922.
59. Machens A, Schneyer U, Holzhausen HJ, Dralle H. Prospects of remission in medullary thyroid carcinoma according to basal calcitonin level. *J Clin Endocrinol Metab* 2005;90:2029-2034.

60. Camacho CP, Lindsey SC, Melo MC, Yang JH, Germano-Neto F, Valente Fde O, Lima TR, Biscolla RP, Vieira JG, Cerutti JM, Dias-da-Silva MR, Maciel RM. Measurement of calcitonin and calcitonin gene-related peptide mRNA refines the management of patients with medullary thyroid cancer and may replace calcitonin-stimulation tests. *Thyroid* 2013;23:308-316.
61. Organ SL, Tsao MS. An overview of the c-MET signaling pathway. *Ther Adv Med Oncol* 2011;3:57-519.
62. Wasenius VM, Hemmer S, Karjalainen-Lindsberg ML, Nupponen NN, Franssila K, Joensuu H. MET receptor tyrosine kinase sequence alterations in differentiated thyroid carcinoma. *Am J Surg Pathol* 2005;29:544-549.
63. Zenali M, deKay J, Liu Z, Hamilton S, Zuo Z, Lu X, Bakkar R, Mills G, Broaddus R. Retrospective Review of MET Gene Mutations. *Oncoscience* 2015;2:533-541.
64. Parameswaran R, Brooks S, Sadler GP. Molecular pathogenesis of follicular cell derived thyroid cancers. *Int J Surg* 2010;8:186-193.
65. Landa I, Ibrahimipasic T, Boucai L, Sinha R, Knauf JA, Shah RH, Dogan S, Ricarte-Filho JC, Krishnamoorthy GP, Xu B, Schultz N, Berger MF, Sander C, Taylor BS, Ghossein R, Ganly I, Fagin JA. Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. *J Clin Invest* 2016;126:1052-1066.
66. Cancer Genome Atlas Research N. Integrated genomic characterization of papillary thyroid carcinoma. *Cell* 2014;159:676-690.
67. Kunstman JW, Juhlin CC, Goh G, Brown TC, Stenman A, Healy JM, Rubinstein JC, Choi M, Kiss N, Nelson-Williams C, Mane S, Rimm DL, Prasad ML, Hoog A, Zedenius J, Larsson C, Korah R, Lifton RP, Carling T. Characterization of the mutational landscape of anaplastic thyroid cancer via whole-exome sequencing. *Hum Mol Genet* 2015;24:2318-2329.
68. Karunamurthy A, Panebianco F, S JH, Vorhauer J, Nikiforova MN, Chiosea S, Nikiforov YE. Prevalence and phenotypic correlations of EIF1AX mutations in thyroid nodules. *Endocr Relat Cancer* 2016;23:295-301.
69. Katoh R, Kawaoi A, Miyagi E, Li X, Suzuki K, Nakamura Y, Kakudo K. Thyroid transcription factor-1 in normal, hyperplastic, and neoplastic follicular thyroid cells examined by immunohistochemistry and nonradioactive in situ hybridization. *Mod Pathol* 2000;13:570-576.
70. Kimura S. Thyroid-specific transcription factors and their roles in thyroid cancer. *J Thyroid Res* 2011;2011:710213.
71. Ngan ES, Lang BH, Liu T, Shum CK, So MT, Lau DK, Leon TY, Cherny SS, Tsai SY, Lo CY, Khoo US, Tam PK, Garcia-Barcelo MM. A germline mutation (A339V) in thyroid transcription factor-1 (TTF-1/NKX2.1) in patients with multinodular goiter and papillary thyroid carcinoma. *J Natl Cancer Inst* 2009;101:162-175.
72. Fabbro D, Di Loreto C, Beltrami CA, Belfiore A, Di Lauro R, Damante G. Expression of thyroid-specific transcription factors TTF-1 and PAX-8 in human thyroid neoplasms. *Cancer Res* 1994;54:4744-4749.
73. Sharretts JM, Simonds WF. Clinical and molecular genetics of parathyroid neoplasms. *Best Pract Res Clin Endocrinol Metab* 2010;24:491-502.
74. Pacini F, Castagna MG, Cipri C, Schlumberger M. Medullary thyroid carcinoma. *Clin Oncol (R Coll Radiol)* 2010;22:475-485.
75. Sidhu S, Campbell P. Thyroid pathology associated with primary hyperparathyroidism. *Aust N Z J Surg* 2000;70:285-287.
76. Morita SY, Somervell H, Umbricht CB, Dackiw AP, Zeiger MA. Evaluation for concomitant thyroid nodules and primary hyperparathyroidism in patients undergoing parathyroidectomy or thyroidectomy. *Surgery* 2008;144:862-866; discussion 866-868.
77. Carling T. Molecular pathology of parathyroid tumors. *Trends Endocrinol Metab* 2001;12:53-58.
78. Kogai T, Taki K, Brent GA. Enhancement of sodium/iodide symporter expression in thyroid and breast cancer. *Endocr Relat Cancer* 2006;13:797-826.
79. Kogai T, Brent GA. The sodium iodide symporter (NIS): regulation and approaches to targeting for cancer therapeutics. *Pharmacol Ther* 2012;135:355-370.
80. Bizhanova A, Kopp P. Minireview: The sodium-iodide symporter NIS and pendrin in iodide homeostasis of the thyroid. *Endocrinology* 2009;150:1084-1090.
81. Riesco-Eizaguirre G, Rodriguez I, De la Vieja A, Costamagna E, Carrasco N, Nistal M, Santisteban P. The BRAFV600E oncogene induces transforming growth factor beta secretion leading to sodium iodide symporter repression and increased malignancy in thyroid cancer. *Cancer Res* 2009;69:8317-8325.
82. Ward LS, Santarosa PL, Granja F, da Assumpcao LV, Savoldi M, Goldman GH. Low expression of sodium iodide symporter identifies aggressive thyroid tumors. *Cancer Lett* 2003;200:85-91.
83. Trouttet-Masson S, Selmi-Ruby S, Bernier-Valentin F, Porra V, Berger-Dutrieux N, Decaussin M, Peix JL, Perrin A, Bournaud C, Orgiazzi J, Borson-Chazot F, Franc B, Rousset B. Evidence for transcriptional and posttranscriptional alterations of the sodium/iodide symporter expression in hypofunctioning benign and malignant thyroid tumors. *Am J Pathol* 2004;165:25-34.
84. Ringel MD, Anderson J, Souza SL, Burch HB, Tambascia M, Shriver CD, Tuttle RM. Expression of the sodium iodide symporter and thyroglobulin genes are reduced in papillary thyroid cancer. *Mod Pathol* 2001;14:289-296.