Florida International University FIU Digital Commons

FCE LTER Journal Articles

FCE LTER

2007

Tropical seagrass-associated macroalgae distributions and trends relative to water quality

Ligia Collado-Vides Southeast Environmental Research Center, Florida International University, colladol@fiu.edu

Valentina G. Caccia Southeast Environmental Research Center, Florida International University, cacciav@fiu.edu

Joseph N. Boyer Southeast Environmental Research Center, Florida International University, boyerj@fiu.edu

James W. Fourqurean Department of Biological Sciences and Southeast Environmental Research Center, Florida International University, fourqure@fiu.edu

Follow this and additional works at: https://digitalcommons.fiu.edu/fce lter journal articles

Recommended Citation

Collado-Vides, L., V.G. Caccia, J.N. Boyer, J.W. Fourqurean. 2007. Tropical seagrass-associated macroalgae distributions and trends relative to water quality. Estuarine Coastal and Shelf Science 73: 680-694.

This material is based upon work supported by the National Science Foundation through the Florida Coastal Everglades Long-Term Ecological Research program under Cooperative Agreements #DBI-0620409 and #DEB-9910514. Any opinions, findings, conclusions, or recommendations expressed in the material are those of the author(s) and do not necessarily reflect the views of the National Science Foundation. This work is brought to you for free and open access by the FCE LTER at FIU Digital Commons. It has been accepted for inclusion in FCE LTER Journal Articles by an authorized administrator of FIU Digital Commons. For more information, please contact dcc@fiu.edu, jkrefft@fiu.edu.

1	Tropical Seagrass-associated Macroalgae Distributions and Trends relative
2	to Water Quality
3	
4	
5	Ligia Collado-Vides ¹ , Valentina G. Caccia ¹ , Joseph N. Boyer ¹ and James W.
6	Fourqurean ^{1,2}
7	
8	¹ Southeast Environmental Research Center, OE 167, ² Department of Biological
9	Sciences, Florida International University, University Park, Miami, FL 33199, USA.
10	
11	Corresponding author: Ligia Collado-Vides
12 13 14 15	email: <u>colladol@fiu.edu</u> Fax 305 348 1986 Southeast Environmental Research Center University Park OE 167

16 Miami, FL 33199

1 ABSTRACT

2 Tropical coastal marine ecosystems including mangroves, seagrass beds and coral 3 reef communities are undergoing intense degradation in response to natural and 4 human disturbances, therefore, understanding the causes and mechanisms 5 present challenges for scientist and managers. In order to protect our marine 6 resources, determining the effects of nutrient loads on these coastal systems has 7 become a key management goal. Data from monitoring programs were used to 8 detect trends of macroalgae abundances and develop correlations with nutrient 9 availability, as well as forecast potential responses of the communities monitored. 10 Using eight years of data (1996 to 2003) from complementary but independent 11 monitoring programs in seagrass beds and water quality of the Florida Keys 12 National Marine Sanctuary (FKNMS), we 1) described the distribution and 13 abundance of macroalgal groups, 2) analyzed the status and spatiotemporal trends 14 of macroalgal groups, and 3) explored the connection between water quality and 15 the macroalgal distribution in the FKNMS. In the seagrass beds of the FKNMS 16 calcareous green algae were the dominant macroalgae group followed by the red 17 group; brown and calcareous red algae were present but in lower abundance. 18 Spatiotemporal patterns of the macroalgae groups were analyzed with a non-linear 19 regression model of the abundance data. For the period of record, all macroalgal 20 groups increased in abundance (Ab_i) at most sites, with calcareous green algae 21 increasing the most. Calcareous green algae and red algae exhibited seasonal 22 pattern with peak abundances (Φ_i) mainly in summer for calcareous green and 23 mainly in winter for red. Macroalgae Ab_i and long-term trend (m_i) were correlated in 24 a distinctive way with water quality parameters. Both the Ab_i and m_i of calcareous

1	green algae had positive correlations with NO_3^{-} , NO_2^{-} , total nitrogen (TN) and total
2	organic carbon (TOC). Red algae Ab_i had a positive correlation with NO ₂ ⁻ , TN, total
3	phosphorus and TOC, and the m_i in red algae was positively correlated with N:P. In
4	contrast brown and calcareous red algae Ab _i had negative correlations with N:P.
5	These results suggest that calcareous green algae and red algae are responding
6	mainly to increases in N availability, a process that is happening in inshore sites. A
7	combination of spatially variable factors such as local current patterns, nutrient
8	sources, and habitat characteristics result in a complex array of the macroalgae
9	community in the seagrass beds of the FKNMS.
10	
11	Key words: Macroalgae, Florida Keys National Marine Sanctuary, monitoring,
12	nutrients, seaweeds, spatiotemporal distribution, synchrony, water quality.
13	
14	Abbreviations: FKNMS Florida Keys National Marine Sanctuary, CGT Calcareous
15	Green Total, GO Green Other, BA Batophora-Acetabularia, RO Red Other, CR
16	Calcareous Red, BO Brown Other.
17	
18	1. Introduction
19	Tropical coastal marine ecosystems including mangroves, seagrass beds,
20	and coral reef communities are undergoing intense degradation in response to

and coral reef communities are undergoing intense degradation in response to
natural and human disturbances (Jackson et al. 2001, McManus and Polsenberg
2004, Orth et al. 2006). Since 1987, several ecosystem-scale disturbances in
Florida Bay and the Florida Keys have occurred, such as seagrass die-off
(Robblee et al. 1991), cyanobacterial blooms (Phlips and Badylak 1996), sponge

1 mortality (Buttler et al. 1995), and a decline in fisheries (Tabb and Roessler 1989, 2 Tilmant 1989). These alterations, combined with growing human population 3 pressure and an economy based on ocean-related tourism provided the impetus to 4 protect and study this marine ecosystem. In 1990 Congress designated the Florida 5 Keys a National Marine Sanctuary (FKNMS). The FKNMS contains diverse 6 assemblages of terrestrial, estuarine, and marine fauna and flora, encompassing over 9500 km². In order to protect the FKNMS, understanding the effects of 7 8 nutrient loads on this coastal system has become a key management goal.

9 A basic ruling premise in plant communities is that nutrient addition shifts the 10 competitive balance from slow-growing primary producers to faster-growing 11 species. In seagrass beds, a gradual shift is expected to occur as nutrient loads 12 are increased (Duarte 1995; Valiela et al. 1997; Hauxwell et al. 2001; McGlathery 13 2001, Fourgurean and Rutten 2003), where macroalgae proliferations might 14 overgrow and displace seagrasses. Nitrogen (N) is frequently a limiting nutrient in 15 coastal systems, but increasing evidence for phosphorus (P) limitation suggests 16 that both N and P enrichment are of concern in nearshore habitats (Howarth 1988). 17 Under short term experimental conditions it has been shown that in P- (Lapointe 18 1989) and N-limited (Larned 1998) environments, tropical macroalgal response to 19 nutrient enrichment varies among regions and is highly species-specific, 20 suggesting that tropical macroalgae exhibit interspecific variation in responses to 21 nutrient enrichment along gradients corresponding to background nutrient influence 22 (Fong et al. 2003). This suite of short term experiments suggests a close 23 interaction between nutrients and macroalgae; and that results are determined by 24 the initial conditions (Ferdie and Fourgurean 2004), as well as by biotic or abiotic

factors, such as grazing pressure, space, or level of disturbance (Artmitage et al
 2005).

3 Long term trends in seagrasses and macroalgae abundance may be reliable 4 indicators of changes in nutrient availability in coastal ecosystems, but the 5 application of such long-term data requires a consistent monitoring of these 6 organisms at multiple sites with different nutrient conditions. In 1995, such a long-7 term monitoring program was established in the FKNMS. In this study, we used 8 data from the long term seagrass monitoring program (Fourgurean et al. 2001, 9 Fourgurean et al. 2003), and the water quality monitoring program (Boyer and 10 Jones, 2002) to detect the long term abundance trends of macroalgal groups and 11 their correlations with median nutrient concentrations for a period of eight years in 12 30 different sites of the FKNMS. Our objectives were to 1) describe the distribution 13 of the abundance of macroalgal groups and water guality parameters, 2) analyze 14 the status and spatiotemporal trends of macroalgal groups, and 3) explore the 15 connection between water quality and macroalgal distribution and trends in the 16 FKNMS.

17

18 2. Materials and methods

19 2.1 Study area

20 The Florida Keys are an archipelago of sub-tropical islands of Pleistocene 21 origin extending over 354 km in length in a southwesterly direction from the 22 southern tip of Florida (Fig.1). The area includes mangrove-fringed shorelines, 23 mangrove islands, seagrass meadows, hard bottom habitats, thousands of patch 24 reefs. and the third largest coral reef system in the world

1 (http://www.fknms.nos.noaa.gov/). The FKNMS is generally divided into three main 2 geographical regions: Upper Keys, Middle Keys, and Lower Keys (Fig. 1). The 3 Lower Keys are most influenced by cyclonic gyres that spin off the Florida Current, 4 the Middle Keys by exchange with Florida Bay, while the Upper Keys are 5 influenced by Florida Current frontal eddies and to a certain extent by exchange 6 with Biscavne Bay (Klein and Orlando 1994). All three regions are also divided into 7 ocean- or bay-side. Ocean-side regions are influenced by wind and tidally driven 8 lateral Hawk Channel transport (Pitts, 1997). The two bay-side regions of the 9 Lower and Middle Keys are distinguished as the Backcountry and Sluiceway (Fig. 10 1). The Backcountry region is a shallow water area associated with many small 11 islands on the Lower Keys, and is influenced by water moving south along the SW 12 Shelf. The Sluiceway may be considered part of western Florida Bay as it is 13 strongly influenced by water transport from Florida Bay, the SW Florida Shelf, and 14 Shark River Slough (Smith, 1994). Many of the Key channels that exchange water 15 between the Gulf of Mexico and the Atlantic Ocean are located in this region, 16 making for large currents and tides.

17 Regional currents may influence water guality over large areas by the 18 advection of external surface water masses into and through the FKNMS (Lee et 19 al. 1994, Lee et al. 2002) and by the intrusion of deep offshore ocean waters onto 20 the reef tract as internal tidal bores (Leichter et al. 1996, Leichter et al. 2003). 21 Local currents become more important in the mixing and transport of freshwater 22 and nutrients from terrestrial sources (Smith 1994; Pitts 1997). As a result of this 23 complex set of currents, water quality of the FKNMS may be directly affected both 24 by external nutrient transport and internal nutrient loading sources.

1 The subtidal benthic marine habitats of the FKNMS are well-described. 2 Most of the benthos of the FKNMS is carpeted by seagrass communities of varying 3 density and species composition (Fourgurean et al. 2002). A smaller, but vitally 4 important, portion of the FKNMS supports coral communities (Porter, 2002). 5 Macroalgae are important components of both the seagrass and coral 6 communities, but for this study we focused on the data from the seagrass 7 monitoring sites. The most common seagrass in the part of the FKNMS that 8 contains long-term seagrass monitoring sites is Thalassia testudinum, which is 9 found from the shoreline across Hawk's Channel to the back-reef area. 10 Syringodium filiforme is commonly encountered as well especially at the more off-11 shore monitoring sites. Halodule wrightii is occasionally present at the monitoring 12 sites. The density and species composition of the seagrasses in south Florida is 13 strongly controlled by nutrient availability (Fourgurean et al 1995, Ferdie and 14 Fourgurean 2004)

15

16 2.2 *Methods*

17 2.2.1 Water quality

Eight years of data were analyzed from the Water Quality Monitoring and Protection Project of the FKNMS, conducted by Southeast Environmental Research Center at Florida International University Water Quality Monitoring Network (Boyer 2005). This project is based on quarterly sampling events (1995 to present) and includes 154 sites within the FKNMS. For this study we used data from March 1996 to May 2003 including 29 quarterly sampling events at 30 sites (Fig. 1). We selected the years and sites to correspond with the macroalgae data

available. Field sampling and laboratory analyses are extensively described in
Boyer and Jones (2002) and are the same used to analyze the present data. All
analyses were completed within 1 month after collection in accordance to SERC
laboratory QA/QC guidelines. All concentrations are reported as µM. All elemental
ratios discussed were calculated on a molar basis, and salinity was measured
using the Practical Salinity Scale.

7 Data from the 30 selected sites were processed to obtain the medians of the 8 8 year record (1996-2003) for selected water quality parameters, as well as the 9 minimum and maximum value for each nutrient. Contour maps of nutrient 10 distributions were produced (Surfer 8, Golden Software), using a kriging algorithm 11 for the medians of Total Nitrogen (TN), Nitrate (NO₃), Nitrite (NO₂) Ammonium 12 (NH_4^+) , Total Phosphorous (TP), and Total Organic Carbon (TOC). A holistic 13 analysis of all 154 sampling sites and 8 years of the nutrient trends can be found in 14 Boyer (2005).

15 2.2.2 Macroalgae

16 Macroalgae abundance was measured guarterly from 1996 to 2003 at 30 17 permanent sites (Fig. 1). Fine scale taxonomic identification of the macroalgae 18 was not always possible in the field, so macroalgae were grouped into easily 19 identifiable groups: Calcareous Green Total (CGT), Batophora-Acetabularia (BA), 20 Green Other (GO), Calcareous Red (CR), Red Other (RO), and Brown Other (BO). 21 Abundance of these groups was scored using a modified Braun-Blanquet method 22 (Fourgurean and Rutten 2003). At each site, the abundance of taxa was recorded in ten randomly located 0.25 m² guadrats along a 50 m permanent transect. The 23

abundance of each group observed in each quadrat was assigned a score
between 0 and 5. A score of 0 indicated that the genus or functional group was
absent, 0.1 indicated the presence of a solitary individual covering < 5% of the
quadrat area, 0.5 indicated few individuals covering < 5%, 1 indicated numerous
individuals covering <5%, 2 indicated 5-25% cover, 3 indicated 25-50% cover, 4
indicated 50-75% cover, and 5 indicated 75-100% cover. Site-specific abundance
of each taxon (Ab_i) was calculated as:

8

$$Ab_i = (\Sigma^n_{i=1} S_{ij})/N_i$$

9 where N_i is the number of quadrats at a site in which taxon *i* occurred, *n* is the total 10 number of quadrats observed, and, S_{ij} is the Braun Blanquet score for taxon *i* in 11 quadrat *j*. Note that the range of possible taxon-specific abundance scores was 0 12 < Ab_i < 5. The spatial distribution of the eight year (1996-2003) mean Ab_i of each 13 macroalgal group was obtained by interpolating mean values throughout the study 14 area with a kriging interpolation routine (point kriging using linear variogram and no 15 nugget, Surfer 8, Golden Software).

In order to analyze the temporal patterns in abundance (e.g. long-term trends, seasonal cycles) for each group at each monitoring site we applied a nonlinear regression model (using the statistical package SPSS) with parameters to incorporate both long-term changes as well as seasonal fluctuations. Time series analyses were conducted using the following model:

21

$$Ab_i = \beta_i + m_i t + \alpha_i \sin(t + \phi_i)$$

where Ab_i was the abundance of group *i*, β_i represented the initial abundance of group *i*, m_i represented the long-term linear trend in abundance of group *i*, *t* was time since the beginning of the time series (time in radians, 1 year = 2 π radians),

1 α_i represented the magnitude of seasonal changes in abundance of group *i*, and ϕ_i 2 (phase angle in radians) represented the timing of seasonal changes in abundance 3 of group *i*. This particular model was chosen for our analyses because a similar 4 approach has been successful in describing the temporal patterns of other aspects 5 of the seagrass and algal communities in the region (Fourgurean et al 2001, 6 Collado-Vides et al. 2005). The model was applied to the time series of 7 abundances for the two most common groups, CGT and RO, for all sites. Because 8 of the patchy distributions of other macroalgal groups (i.e. GO, BA, CR and BO), 9 only the time series from sites with consistent abundance during all studied period 10 were selected.

In order to detect seasonality of macroalgal groups abundance, we evaluated if the α_i parameter estimate was significantly different from zero. Once we detected seasonality, we applied a t-test to compare ϕ mean between CGT and RO, the only groups with a clear seasonal pattern.

15 To evaluate any relationships between temporal patterns in population 16 abundance and geographic location at different spatial scales, a Kruskal-Wallis test 17 was used to test group-specific differences in Ab_i, m_i , α_i and ϕ_i as a function of 18 different geographic divisions of the FKNMS based on three different criteria. We 19 tested for differences among the FKNMS segments proposed by Klein and Orlando 20 (1994): Upper Keys (UK), Middle Keys (MK), Lower Keys (LK) on the ocean side of the Florida Keys; and Sluiceway, Hawk Channel and Backcountry (BC) with two 21 22 sub-segments BC3 and BC4 on the bay side (Fig. 1). We also tested for 23 differences among strata of offshore distances because of the spatial pattern in 24 nutrient limitation along this gradient (Fourgurean and Zieman 2002). The final

classification was based on alongshore distance representing the longitudinal
 distance from the highly urban area of Miami.

To detect any relationship between macroalgal group abundance *Ab_i*, long
term trends *m_i*, and water column nutrient concentrations, a non-parametric
correlation analysis (Kendall's τ-b) was applied to the site specific data.

6

7 3. Results

8 3.1 Water quality

9 For the period studied, the Florida Keys had a median surface water 10 temperature of 27.7 °C, with maximum values during summer (35.4 °C) and 11 minimum during winter (16.0 °C). Salinity median was 36.3 with maximum values 12 during summer (39.7) and minimum during winter (27.9) with low variability 13 spatially.

14 In general, the FKNMS exhibited oligotrophic water quality condition with 15 median NO₃, NO₂ and NH₄⁺ concentrations of 0.09 μ M, 0.05 μ M and 0.29 μ M, 16 respectively. NH_4^+ was the dominant DIN species in almost all of the samples (~70 17 %). However, DIN (NO₃⁻, NO₂⁻ and NH₄⁺) comprised a small fraction (4 %) of the 18 TN pool with TON making up the bulk (median 10.78 μ M) and TP median was 0.20 19 µM. Molar ratios of TN:TP suggested a general P limitation of the water column 20 (median = 58). TOC median was 189.4 μ M; a value higher than open-ocean levels 21 but consistent with coastal areas.

DIN concentrations were highest in the Backcountry and Sluiceway subregions of the Lower and Middle Keys. NO_3^- was highest at site 260 (0.34 μ M) the ocean side of the Lower Keys region; site 285 (0.24 μ M) in the Sluiceway

1 subregion of the Middle Keys, and site 235 (0.24 μ M) the ocean side of the Middle Keys. NO_2^- exhibited the same behavior. NH_4^+ showed several sites of high 2 3 concentration (>0.5 μ M): site 314 in the Backcountry sub-region of the Lower Keys, 4 site 260 in ocean side of the Lower Keys, site 235 and 241 in the ocean side of the 5 Middle Keys. The distribution of TN and TON were very similar, exhibiting their 6 highest concentrations (14-18 μ M) in the Bay side of the Lower and Middle Keys. 7 Backcountry subregion in sites 296, 307 and 314, Sluiceway subregion sites 284, 8 285 and 287, and in the ocean side in sites 260 in the Lower Keys and 235 in the 9 Middle Keys (Fig. 2).

10 The highest concentrations of TP (>0.26 μ M) were found in all five 11 Backcountry sites. The TN:TP ratio showed a similar distribution pattern than the 12 inorganic nutrients. TOC was higher in Sluiceway and the Backcountry (>230 μ M), 13 and was also distributed as a gradient from inshore to offshore (Fig. 2).

In general, the Upper Keys showed very low concentrations of all water quality parameters, except site 214 (the nearest to the coast) that had mediumhigh concentrations of NO_3^- , NH_4^+ , TN:TP (Fig. 2).

17 Depth ranged from 2.7 m in site 296 to 10.6 m in site 216. In general the 18 only region characterized by shallow sites was Sluiceway (2-4 m), the rest of the 19 regions had sites with various depths.

20

21 3.2 Macroalgae

The Florida Keys had mainly tropical macroalgal species as their characteristic aquatic non-vascular flora. In the seagrass beds of the FKNMS, green algae were mainly represented by calcareous algae such as species of the

genera Halimeda, Penicillus, Rhipocephallus, Udotea, or non calcareous green
 algae such as species of the genera Avrainvillea, Caulerpa, Acetabularia,
 Batophora, Anadyomene among others. Red algae were represented by species of
 the genera Laurencia, Chondria, Acanthophora, Gracilaria among others. Brown
 algae were mainly represented by species of the genera Sargasum, and Dictyota.
 Many other species were epiphytic on seagrass blades but were not included in
 this study.

8 Results of the monitoring program show that all algal groups were present 9 and encountered year-round and throughout the eight-year span of our data, but 10 there were large differences in the frequency of encounter and mean abundances 11 of the algal groups. The consistently most abundant group of algae during the 8 12 year period was the CGT, followed by the RO. The rest of groups were present, 13 but in an order of magnitude lower mean abundance (Fig. 3). Each group had a 14 unique distribution. CGT was characterized by the highest abundance and widest 15 distribution, with some high abundance spots in Backcountry (site 307) and 16 Sluiceway (site 285); lower abundance was found at the ocean side of the Keys at 17 sites 243, 255, and 273 (Fig. 3). RO had an intermediate level of abundance and a 18 distribution more or less similar to that of the CGT; high abundance levels for RO 19 were found mainly at sites 285 and 294 both in Sluiceway (Fig. 3). GO, CR and BO 20 were characterized by low abundance and very patchy distribution.

The fits of our non-linear regression model to the abundance time series varied between the algae groups, the model generally described the time series data reasonably well for CGT and RO (Fig. 4), but the efficacy of the model varied among sites for the rarer groups (BO, BA, GO and CR). For this reason, we have

1 only analyzed the spatial patterns in the model parameters m_i , α_i and ϕ_i for CGT 2 and RO.

3 Seasonality in the time series of macroalgal group abundance was 4 assessed using model estimates of the α_i parameter; if the parameter estimate was 5 significantly different from zero at the 0.05 confidence level (i.e., if the asymptotic 6 95% confidence interval for the value of the parameter did not contain zero) we 7 concluded that there was a significant seasonal pattern in the time series. Using 8 this criterion, only the time series of the two most abundant groups, CGT and RO, 9 displayed significant seasonality for most sites. The Φ_i or timing of peaks in 10 abundance between these macroalgal groups was significantly different (T-test, p< 11 0.04). Both groups showed variability with peaks in different seasons for different 12 sites. For CGT 13 sites out of 30 peaked in summer, 8 in fall, 5 in spring, and only 13 4 in winter. In contrast, for RO 11 sites peaked in winter, 9 in fall, 6 in spring, and 14 only 4 in summer.

15 The long-term trends (i.e., m_i) were significantly positive for the majority of 16 sites for all groups, indicating that there were widespread increases in macroalgal 17 abundance across the FKNMS, and that the increases were occurring in all 18 monitored algal groups (Fig. 5). However, each group had a unique spatial 19 behavior with highest slopes at different sites. CGT had the highest slopes in the ocean side at sites 235 ($m_i = 0.42/y^{-1}$, 95% confidence interval $0.38 \le m_i \le 0.47$) 20 and 241 ($m_i = 0.23/y^{-1}$ 95% confidence interval 0.17 $\leq 0.23 \leq 0.29$) in the Middle 21 Keys, and site 260 ($m_i = 0.20/y^{-1}$, $0.15 \le m_i \le 0.25$) Lower Keys. RO had the 22 23 highest values at sites 294 at Sluiceway in the bay side of the Lower Keys ($m_{=}$ $0.20/y^{-1}$, $-0.036 \le m \le 0.43$), and in the ocean side RO had high values in the 24

1 Upper Keys, site 214 ($m_i = 0.13y^{-1}$, 0.048 $\le m_i \le 0.21$) Middle Keys site 237 (0.17/y⁻ 2 ¹, 0.027 $\le m_i \le 0.31$) and Lower Keys 273 ($m_i = 0.14/y^{-1}$, 0.002 $\le m_i \le 0.28$) (Figs. 1 3 and 5).

4 Abundance and trends in abundance of macroalgal groups exhibited 5 complicated relations with geographic patterns. Only CGT average Ab_i and m_i 6 showed significant mean differences among offshore strata, (Kruskall Wallis Ab p< 7 0.01, $m_p < 0.02$, with higher values closer to land indicating that CGT was more 8 abundant and Ab increased faster closer to land (Fig. 6). Long-term trends for RO 9 had significant mean Ab, and m_i differences among segment (Kruskall Wallis Ab,p< 10 0.04, m_i p< 0.05) and significant mean Ab differences among alongshore (Kruskall 11 Wallis Ab p< 0.04) strata, with lower values in Backcountry subregion 3 compared 12 with Sluiceway which had low to medium values (Fig. 7). The intra-annual 13 variability α_i and abundance peak Φ_i did not showed any significant differences 14 among the three different geographic categories tested.

15

16 *3.3 Macroalgae and water quality*

Significant positive correlations were found between CGT Ab_i and m_i with different forms of N (NO₃⁻, NO₂⁻, TN, TON) and TOC in the water column (Table 1, Fig. 8). RO Ab_i had a significant positive correlation with NO₂⁻, TN, TP and TOC; and the long-term trend of RO m_i with N:P (Table 1, Fig. 9). CR Ab_i had a significant negative correlation with TN:TP, and BO Ab_i had significant negative correlation with TN:TP (Table 1). BA Ab_i did not have any significant correlation with any water quality parameters (Table 1).

2 4. Discussion

This study show general trends and patterns and simple relationships between the spatiotemporal patterns of macroalgae abundance and median values of nutrients. The trends in abundance were trends only detectable by such a longterm monitoring program. Our analyses suggest that both the abundance and longterm increases in abundance of major macroalgal groups in the FKNMS were highest in the parts of our study areas with the highest availability of N in the water column.

10 Several physical factors such as light, salinity, and nutrients are known to 11 affect the physiology and abundance of macroalgae (Lobban and Harrison 1997). 12 At the physical level the region studied showed a clear seasonal pattern in its 13 temperature and salinity, as the Florida Keys are located in a subtropical region. 14 However at the spatial level differences in salinity and temperature were probably 15 not the factor causing regional patterns in algal abundance. The sites sampled are 16 all located out of the influence of the freshwater entering Florida Bay (Boyer and 17 Jones 2002), unlike the adjacent Florida Bay, where salinity changes are strong 18 and have influence on the abundance and distribution of macroalgae (Biber and 19 Irlandi 2006). However, nutrient concentrations were found to differ spatially across 20 the FKNMS (Fig. 2); it is likely that the spatial patterns in macroalagal abundance 21 were functions of the pattern in nutrient availability.

The phycological flora found in the Florida Keys is very similar to the rest of the Caribbean (Taylor 1960, Littler and Littler 2000, Dawes and Mathieson 2002). The dominant group in the seagrass beds was the CGT, dominated by species of

1 the genus Halimeda (Collado-Vides et al. 2005) followed by the RO, dominated by 2 Laurencia. These results in general are similar to the reported flora by Biber and 3 Irlandi (2006) for Florida Bay however the distribution might differ in particular 4 cases such as Batophora that was found dominant by Zieman et al. (1989) in 5 Florida Bay. Batophora was found in high abundance in Backcountry, which is 6 similar to the general features of Florida Bay, and was present but inconspicuous in the rest of the FKNMS. The physical characteristics of each region and the 7 8 inherent limitation of macroalgae to find the right substrate results in the patchy 9 distribution found in this study. BA (Batophora and Acetabularia) are species 10 characterized by small forms (up to 10 cm), usually found on hard substrata, i.e. 11 small shells or hard rock, limiting its distribution from general sandy seagrass 12 bottoms.

13 Spatiotemporal covariation, also known as synchrony, has been shown to 14 provide helpful information on population dynamics by facilitating detection of 15 common trends in variation at different time and spatial scales (Bjørnstad et al. 16 1999, Driskell et al. 2001). In this study synchrony is represented by Φ_i value of the 17 CGT displayed highly synchronized seasonal patterns of regression model. 18 abundance; with higher abundances during summer and fall when temperatures 19 are high, and lower during winter when temperatures are low reflecting the fact that 20 the Florida Keys are in a subtropical region with a marked seasonal behavior of its 21 populations and they behave synchronically (Lunning 1993, Makarov et al 1999). 22 The red algae also had a seasonal pattern but with high abundance during late 23 fall/winter. This seasonal trend corroborates the findings of other studies conducted 24 on marine coastal lagoons and coral reef environments which described a clear

pattern of increasing abundance in green algal species during summer-fall and a
subsequent decay in winter-spring, and an increase of red algae during the winterspring and decay in summer-fall (Collado-Vides et al. 1994, Lirman and Biber
2000, Vroom et al. 2003, Artmitage et al 2005, Biber and Irlandi 2006).

5 Shifts from seagrass to macroalgal communities have been associated with 6 nutrient increases in subtropical to temperate zones (Deegan et al 2002,

McClanahan 1999, McClanahan et al. 2002, McClanahan et al. 2003, McClanahan
et al. 2005). Similar mechanisms may influence shifts from corals or seagrass bed
to algal dominated communities in the Caribbean and elsewhere too (Lapointe
1999, Duarte 1995; Valiela et al. 1997; Hughes et al. 1999, Hauxwell et al. 2001;
McGlathery 2001).

12 Our results indicate that the abundance of almost all macroalgal groups was 13 increasing in the FKNMS over the course of our study; particularly at sites with high 14 N concentrations suggesting a limitation of N in general for at least CGT and RO. 15 Eutrophication has been blamed for macroalgal bloom in the Florida Keys 16 (Lapointe et al., 1994); and macroalgal increases, as a response of short-term 17 nutrient enrichment, have been characterized by rapid increase of non corticated 18 filaments (Lapointe et al. 2004, Karez et al. 2004). However, we found a slow and 19 steady increase of slow growing calcareous green algae, that can not be defined 20 as a macroalgal bloom, but steady increase of its abundance over 8 years of 21 monitoring.

As a long term trend, red algae had a positive correlation with N, similar to experimental results in which enrichment with NH₄⁺ resulted in increased photosynthesis and growth during summer of the red algae *Gracilaria tikvahiae* and

1 of Laurencia intricata and Digenia simplex in the Bahamas (Lapointe et al 2004), 2 and Laurencia papillosa and Gracilaria coonopifolia in Taiwan reefs (Tsai et al. 3 2004). It has also been reported, for some temperate red algae, that nutrient 4 uptake is biphasic allowing these algae to exploit transient pulses of high nutrients 5 (Lobban and Harrison 1997). Red algae might be exploiting the transient pulses of 6 high nutrients reported for the FKNMS as upwelling episodic events (Leichter et al. 7 2003), affecting the offshore sites of the Keys, as well as other sources of nutrients 8 coming from land use such as the high nutrient concentrations found close to land 9 (Boyer and Jones 2002).

In contrast, a negative significant correlation between BO abundance and
TN, was found; brown algae growth can be inhibited by high N concentrations
(McClanahan et al. 2005), which is consistent with the negative correlation of BO
found in our data, however no explanation is found still for this response.

14 The N limitation of CGT, has been demonstrated experimentally in this 15 region. Davis and Fourgurean (2001) studied the competitive interaction between 16 the seagrass *Thalassia testudinum* and the calcareous macroalga *Halimeda* 17 *incrassate*; their findings suggest that competition for nutrients was the mechanism 18 of interaction. An increase in nutrients closer to land might relieve the competition 19 between *T. testudinum* and *Halimeda* spp. explaining the increase of the slope of 20 the algae in these areas. These results are consistent with our results in which the 21 higher slopes were found significant correlated to offshore distance, having higher 22 values closer to land. However, Ferdie and Fourgurean (2004) showed that the 23 response to increasing nutrients in seagrass beds might vary as a function of the 24 initial status of nutrient limitation; in their study, enrichment resulted in an increase

1 on the seagrass biomass at offshore sites, and in contrast in the inshore sites the 2 enrichment leaded to an increase in algal biomass including Halimeda. This 3 suggests that a continuous nutrient enrichment could lead to a shift from *Thalassia* 4 testudinum to Syringodium filiforme in offshore sites, and to algal communities at 5 inside shore (closer to land) sites. Also, Armitage et al. (2005) found, in their 6 experimental nutrient enrichment in Florida Bay, that in general nutrient enrichment 7 did not stimulated algal growth to the level to overgrow the seagrass beds, 8 however some increases in calcareous green and ephemeral filamentous red were 9 detected. This suite of results can be interpreted to suggest that in the Florida Keys 10 and Florida Bay seagrass beds, calcareous green algae can be the first group of 11 macroalgae to increase as nutrients loads are increased as well as some 12 ephemeral red filamentous algae as epiphytes on seagrass blades.

13 Short term field studies in tropical regions suggest that it is difficult to find a 14 significant correlation between N or P concentration and abundance of macroalgae 15 (McCook et al. 1997, McCook 1999), and has been explained by the fact that 16 physical and chemical processes controlling the availability of nutrients are very 17 complex (Fong et al. 2001). However, in this long term, large scale region sampling 18 program, we have been able to integrate the seasonal and yearly variability of 19 macroalgal abundance and detect significant correlations between median water 20 quality concentrations and macroalgae patterns in the FKNMS. These findings 21 suggest that in areas with high nutrient concentration CGT and RO had higher 22 slope values. Water quality parameters were higher in the Lower and Middle Keys 23 than in the Upper Keys, and generally decreased from inshore to offshore 24 consistent with a previous transect survey from these areas (Szmant and

Forrester, 1996); high N concentrations were found in the Middle Keys at the sites
nearest to the shore (285, 241 and 235 sites with high CGT slope), these sites
might be influenced by local anthropogenic inputs and the transport of the high N
concentrations found in the western of Florida Bay, Shark River and Florida Shelf.

5 Nutrients are important for the algal communities as shown in this study; 6 however we do not disregard other factors that might be playing a role in the long-7 term trends in the FKNMS macroalgal communities. It is possible that the 8 distribution patterns and trends found may be a response to some unidentified 9 region-wide disturbance in the past. Fourgurean and Rutten (2004) showed that 10 calcareous macroalgae were much more susceptible to disturbance from Hurricane 11 Georges than the sea grasses in the region. However, that same study showed 12 that prestorm abundance of calcareous green macroalgae were reached within 3 13 years of the disturbance. If the increase we found is the result of the 14 reestablishment after a disturbance, that disturbance must have been of 15 significantly greater magnitude than Hurricane Georges.

16 It is well recognized that decrease in herbivore activities is an import factor 17 for observed coastal ecosystems changes including shift of coral dominated 18 communities into macroalgal dominated communities (Jackson et al. 2001, 19 MacManus and Polsenberg 2004, McClanahan et al. 2003, 2005). The Florida 20 Keys is a heavily fished area (Bohnsack et al. 1994), and macroalgal communities 21 in the reef as well as in the seagrass beds might be having a lower pressure, 22 allowing some groups to increase, however not all macroalgal groups respond 23 rapidly to herbivore exclusion in reef environments (McClanahan 1997).

1 The patchy and complex distribution of nutrients, as well as currents and 2 sedimentation pattern play a role in the macroalgal distribution patterns found in 3 this long term study, within a context of disturbance history, particularities of those 4 sites, and history of herbivore activity.

5

6 5. Conclusions

7 The monitoring of the macroalgae at the group level was very useful to give 8 us a general idea of the main trends with a good level of accuracy. A base line or 9 status of the macrolagae and their trends is given with an analysis of their 10 correlations with nutrients availability. The main results show a relationship 11 between the CGT and N having an increasing trend of CGT closer to land sites.

12 The multifactorial processes that determines the nutrient availability, as well 13 as multi-species component of each algal group make difficult to achieve a cause-14 effect interaction between the abundance of macroalgae and water quality results, 15 however, with this type of monitoring programs we have been able to detect trends 16 and set a base line of the status of the macroalgae in the FKNMS that are 17 explained by results of experimental studies. The combination of complex water 18 circulation patterns, diverse sources of nutrients, initial conditions and competitive 19 interactions between benthic vegetation, can determine the increase of macroalgae 20 detected, and these processes can vary at very local scale.

21

22 6. Acknowledgments

Data were provided by the SERC-FIU Water Quality Monitoring Network which is
supported by EPA Agreement #X994621-94-0 and SFWMD/SERC Cooperative

1	Agreements #C-13178. The macroalgal sampling was conducted with the support
2	of the Water Quality Protection Program of the Florida Keys National Marine
3	Sanctuary, funded by the USEPA (contract X97468102-0) and the National
4	Oceanographic and Atmospheric Administration (contract NA16OP2553). This
5	eight year monitoring program was possible to conduct thanks to the field work of
6	many students and technicians, including L. Rutten, C. D. Rose, A. Willsie, C.
7	Furst, M. Ferdie, D. Byron, V. C. Cornett and K. Cunniff. We also thank Captain
8	Dave Ward of the R/V Magic and Captain Mick O'Connor of the R/V Expedition II
9	for vessel and logistical support. Anna Armitage and two anonymous reviewers
10	made useful comments that helped improve the manuscript. This is contribution #x
11	of the Southeast Environmental Research Center at FIU.
12	
13	7. References
13 14	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental
13 14 15	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and
13 14 15 16	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28 , 422–434.
13 14 15 16 17	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28 , 422–434.
13 14 15 16 17 18	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgae
13 14 15 16 17 18 19	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgal communities along anthropogenic salinity gradient in Biscayne Bay (Florida, USA).
13 14 15 16 17 18 19 20	7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgal communities along anthropogenic salinity gradient in Biscayne Bay (Florida, USA). <i>Aquatic Botany</i> 85, 65-77.
 13 14 15 16 17 18 19 20 21 	 7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgal communities along anthropogenic salinity gradient in Biscayne Bay (Florida, USA). <i>Aquatic Botany</i> 85, 65-77.
 13 14 15 16 17 18 19 20 21 22 	 7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgal communities along anthropogenic salinity gradient in Biscayne Bay (Florida, USA). <i>Aquatic Botany</i> 85, 65-77. Bjørnstad, O. Ims, R. A., & Lambin, X. 1999. Spatial population dynamics:
 13 14 15 16 17 18 19 20 21 22 23 	 7. References Armitage, A., Frankovich T, Heck, K. Jr. & Fourqurean J.W. 2005. Experimental nutrient enrichment causes complex changes in seagrass, microalgae, and macroalgae community structure in Florida Bay. <i>Estuaries</i> 28, 422–434. Biber, P. D. and Irlandi, E. A. 2006. Temporal and spatial dynamics of macroalgal communities along anthropogenic salinity gradient in Biscayne Bay (Florida, USA). <i>Aquatic Botany</i> 85, 65-77. Bjørnstad, O. Ims, R. A., & Lambin, X. 1999. Spatial population dynamics: analyzing patterns and processes of population synchrony. <i>Trend in Ecology and</i>

1	
2	Bohnsack, J. A., Harper, D. E., & McClellan, D. B. 1994. Fisheries trends from
3	Monroe County, Florida. Bulletin of Marine Science 54, 982-1018.
4	
5	Boyer, J. N. & Jones, R. D. 2002 A view from the bridge: External and internal
6	foreces affecting the ambient water quality of the Florida Keys National Marine
7	Sanctuary. In Poter J.W. & Porter, K.G. (eds) The Everglades, Flroida Bary and
8	Coral Reefs of the Florida Keys. CRC PressBoca Raton. 609-40 pp.
9	
10	Boyer, J.N., 2005. Annual Report of the water quality monitoring project for the
11	water quality protection program in the Florida Keys National Marine Sanctuary.
12	Southeast Environmental Research Center, Florida International University.
13	http://serc.fiu.edu/wqmnetwork/FKNMS-CD/index.htm
14	
15	Butler, I., M. J., Hunt, J. H., Herrnkind, W. F., Childress, M. J., Bertelson, R.,
16	Sharp, W., Mathews, T., Field, J. M., & Marshall, H. G 1995. Cascading
17	disturbances in Florida Bay, USA: Cyanobacteria blooms, sponge mortality, and
18	implications for juvenile spiny lobsters Panulirms argus. Marine Ecology Progress
19	<i>Series</i> 129 , 119-125.
20	
21	Collado-Vides, L., González-González, J. & Gold-Morgan M. 1994. A descriptive
22	approach to the floating masses of algae of a Mexican Caribbean coastal lagoon.
23	<i>Botanica Marina</i> 37 , 391–396.

1	Collado-Vides, L., Rutten, L.M. & Fourqurean, J. W. (2005) Spatiotemporal
2	variation of the abundance of calcareous green macroalgae in the Florida Keys: A
3	study of synchrony within a macroalgal functional-form group. Journal of Phycology
4	41 , 742-752.
5	
6	Davis, B. C. & Fourqurean, J. W. 2001. Competition between the tropical alga,
7	Halimeda incrassata, and the seagrass, Thalassia testudinum. Aquatic Botany 71,
8	217-32.
9	
10	Dawes, C. J. & Mathieson A.C 2002. A guide to seaweeds of Florida. Journal of
11	<i>Phycology</i> 38 (s1), 5-6.
12	
13	Deegan, L. A., Wright, A., Ayvazian, S. G., Finn, J. T., Golden, H. & Merson, R. R.
14	2002. Nitrogen loading alters Seagrass ecosystems structure and support of higher
15	trophic levels. Aquatic Conservation: Marine and Freshwater Ecosystems. 12, 193-
16	212.
17	
18	Driskell, W. B., Ruesink, J. L., Lees, D. C., Houghton, J. P. & Lindstrom, S. D.
19	2001. Long-term signal of disturbance: Fucus gardneri after the EXXON VALDEZ
20	oil spill. Ecological Applications 11, 815-27.
21	
22	Duarte, C. M. 1995. Submerged aquatic vegetation in relation to different nutrient
23	regimes. <i>Ophelia</i> 41 , 87–112.

1	Ferdie, M. & Fourqurean, J. W. 2004. Responses of seagrass communities to
2	fertilization along a gradient of relative availability of nitrogen and phosphorus in a
3	carbonate environment. Limnology and Oceanography 49, 2082–2094.
4	
5	Fong, P., Kamer, K. Boyer, K. E. & Boyle, K. A. 2001. Nutrient content of
6	macroalgae with differing morphologies may indicate sources of nutrients for
7	tropical marine systems. Marine Ecology Progress Series 220, 137-152
8	
9	Fong, P., Boyer, K. E., Kamer, K. & Boyle, K. A. 2003. Influence of initial tissue
10	nutrient status of tropical marine algae on response to nitrogen and phosphorus
11	additions. Marine Ecology Progress Series 262, 111–123.
12	
13	Fourqurean, J. W., Boyer, J.N., Durako, M. J., Hefty, L. N. & Bradley, J. P. 2003.
14	Forecasting responses of seagrass distribution to changing water quality using
15	monitoring data. Ecological Applications 13, 474-89.
16	
17	Fourqurean, J. W., M. J. Durako, M. O. Hall, and L. N. Hefty. 2002. Seagrass
18	distribution in south Florida: a multi-agency coordinated monitoring program.
19	Pages 497-522 in J. W. Porter and K. G. Porter, editors. The Everglades, Florida
20	Bay, and the coral reefs of the Florida Keys. CRC Press, Boca Raton.
21	
22	Fourqurean, J. W. & Rutten, L. M. 2003. Competing goals of spatial and temporal
23	resolution: Monitoring seagrass communities on a regional scale. In: Busch, D. E.

1	& Trexler, J. C. (Eds.) Monitoring Ecosystems: Interdisciplinary approaches for
2	evaluating ecoregional initiatives. Island Press, Washington, pp. 257-88.
3	
4	Fourqurean, J. W., Willsie, A. W., Rose, C. D. and Rutten, L. M. 2001. Spatial and
5	temporal patterns in seagrass community composition and productivity in South
6	Florida. Marine Biology 138, 341-354.
7	
8	Fourqurean, J. W. & Zieman, J. C. 2002. Nutrient content of the seagrass
9	Thalassia testudinum reveals regional patterns of relative availability of nitrogen
10	and phosphorus in the Florida Keys USA. Biogeochemistry 61, 229-245.
11	
12	Hauxwell, J., Cebrian, J., Furlong, C. & Valiela, I. 2001. Macroalgal canopies
13	contribute to eelgrass (Zostera marina) decline in temperate estuarine ecosystems.
14	<i>Ecology</i> 82 , 1007–1022.
15	
16	Howarth, R. W. 1988. Nutrient limitation of net primary production in marine
17	ecosystems, In: Johnston, R. F. (ed.), Annual Review of Ecology and
18	Systematics, Annual Reviews, Inc., Palo Alto, California 9, 89–110.
19	
20	Hughes, T. P., Szmant, A. M., Steneck, R., Carpenter, R., & Miller, S. 1999. Algal
21	blooms on coral reefs: what are the causes? Limnology and Oceanography
22	42 ,1583-86.

1	Jackson, J.B. C., Kirkby, M. X., Berger, W. H., Bjorndal, K. A.Botsford, L. W.,
2	Bouque. B. J., Bradbury, R., Cooke, R., Estes, J.A., Hughes, T. P., Kidwell, S.
3	Lange, C. B., Lenihan, H. S., Pandolfi, J. M., Peterson, C. H., Steneck, R. S.,
4	Tegner M. J., Warner, R. 2001. Historical overfishing and the recent collapse of
5	coastal ecosystems. Sciences, 293, 629-638.
6	
7	Klein, C. J. III & Orlando, S. P. Jr. 1994. A spatial framework for water-quality
8	management in the Florida Keys National Marine Sanctuary. Bulletin of Marine
9	<i>Sciences</i> 54 , 1036-1044.
10	
11	Lapointe, B. E. 1989. Macroalgal production and nutrient relations in oligotrophic
12	areas of Florida Bay. Bulletin of Marine Science 44, 312–323.
13	
14	Lapointe, B.E., 1999. Simultaneous top-down and bottom-up forces control
15	macroalgal blooms on coral reefs. Limnology and Oceanography 44, 1586–1592.
16	
17	Lapointe, B. E., Barile, P.J., Yentsch, C. S., Littler, M.M., Littler, D.S. & Kakuk, B.
18	2004. The relative importance of nutrient enrichment and herbivory on macroalgal
19	communities near Norman's Pond Cay, Exumas Cays, Bahamas: a "natural"
20	enrichment experiment. J. Exp. Mar. Biol. Ecol. 298: 275-301.
21	
22	Larned, S. T. 1998. Nitrogen- versus phosphorus-limited growth and sources of
23	nutrients for coral reef macroalgae. Marine Biology 132,409-421.
24	

1	Lee, T. N., Clarke, M. E., Williams, E., Szmant, A. F., & Berger. T. 1994. Evolution
2	of the Tortugas gyre and its influence on recruitment in the Florida Keys. Bulletin of
3	<i>Marine Science</i> 54 , 621-646.
4	
5	Lee, T. N., Williams, E., Johns, E., Wilson, D. & Smith. N. P. 2002. Transport
6	processes linking South Florida ecosystems. In: Porter, J. W., Porter K. G. (eds.),
7	The Everglades, Florida Bay, and Coral Reefs of the Florida Keys: An Ecosystem
8	Sourcebook. CRC Press pp. 309-342.
9	
10	Leichter, J. J., Wing, S. R., Miller, S. L. & Denny. M. W. 1996. Pulsed delivery of
11	subthermocline water to Conch Reef (Florida Keys) by internal tidal bores.
12	Limnology and Limnology and Oceanography 41, 1490-1501.
13	
14	Leichter, J. J., Stewart H. L. & Miller S. L. 2003. Episodic nutrient transport of
15	Florida coral reefs. Limnology and Oceanography 48, 1394-1407.
16	
17	Lirman, D. & Biber, P. 2000. Seasonal dynamics of macroalgal communities of the
18	Northern Florida reef tract. Botanica Marina 43, 305–14.
19	
20	Littler, M.M. & Littler, D. S. 2000. Caribbean reef plants. Offshore Graphics,
21	Washington D.C. 542 pp.
22	
23	Lobban, C. S. & Harrison, P. J. (1997). Seaweed ecology and physiology. NY,

24 Cambridge University Press. 366 pp.

1	
2	Lüning, K. 1993. Environmental and internal control of seasonal growth in
3	seaweeds. Hydrobiologia 260/261, 1-14.
4	
5	Makarov, V. N., Makarov M. V. & Schoschina E. V. 1999. Seasonal dynamics of
6	growth in the Barents Sea seaweeds: endogenous and exogenous regulation.
7	Botanica Marina 42 , 43-49.
8	
9	McClanahan, T. R. 1997. Primary succession of coral-reef algae: Differing patterns
10	on fished versus unfished reefs. Journal of Experimental Marine Biology and
11	<i>Ecology</i> , 218, 77-102.
12	
13	McClanahan, T.R., 1999. Predation and the control of the sea urchin Echinometra
14	viridis and fleshy algae in the path reefs of Glovers Reef, Belize. Ecosystems 2,
15	511–523.
16	
17	McClanahan, T.R., Cokos, B.A., Sala, E. 2002. Algal growth and species
18	composition under experimental control of herbivory, phosphorus and coral
19	abundance in Glovers Reef, Belize. Marine Pollution Bulletin 44, 441–451.
20	
21	McClanahan, T.R., Sala, E., Stickels, P., Cokos, B.A., Baker, A., Starger, C.J. &
22	Jones, S. 2003. Interaction between nutrients and herbivory in controlling algal
23	communities and coral condition on Glover_s Reef, Belize. Marine Ecology
24	Progress Series 261, 135–

1	147.
2 3	
4	McClanahan T.R, Steneck, R. S., Pietri, D., Cokos B. & Jones, S. 2005.
	Interaction between inorganic nutrients and organic matter in controlling coral reef
5	communities in Glovers Reef Belize. Marine Pollution Bulletin 50: 566-575.
6	
7	McCook, L. J., Price, I. R. & Klumpp, D. W. 1997. Macroalgae on the GBR: Causes
8	or consequences, indicators or models of reef degradation? Proceedings of the 8th
9 10	<i>Coral Reef Symposium</i> 2 , 1851-1856.
11	McCook, L. J. 1999. Macroalgae, nutrients and phase shifts on coral reefs:
12	scientific issues and management consequences for the Great Barrier Reef. Coral
13	<i>Reefs</i> 18 , 357-367.
14	
15	McGlathery, K. J. 2001. Macroalgal blooms contribute to the decline of seagrass in
16	nutrient-enriched coastal waters. Journal of Phycology 37, 453-456.
17	
18	McManus, J. W. & Polsenberg, J. F. 2004. Coral-algal phase shifts on coral reefs:
19	ecological and environmental aspects. Progress in Oceanography 60, 263–79.
20	
21	Orth, R. J., T. J. B. Carruthers, W. C. Dennison, C. M. Duarte, J. W. Fourqurean,
22	K. L. Heck, A. R. Hughes, G. A. Kendrick, W. J. Kenworthy, S. Olyarnik, F. T.
23	Short, M. Waycott, and S. L. Williams. 2006. A global crisis for seagrass
24	ecosystems. <i>BioScience</i> 56, 987-996.

2	Phlips, E. J. & Badylak, S. 1996. Spatial variability in phytoplankton
3	standing crop and composition in a shallow innershelf lagoon, Florida Bay, Florida.
4	Bulletin of Marine Science 58, 203-216.
5	
6	Pitts, P. A. 1997. An investigation of tidal and nontidal current patterns in Western
7	Hawk Channel, Florida Keys. Continental Shelf Research 17, 1679-1687.
8	
9	Porter, J. W. 2002. Coral reef monitoring in south Florida. in J. W. Porter and K. G.
10	Porter, editors. The Everglades, Florida Bay, and the coral reefs of the Florida
11	Keys. CRC Press, Boca Raton.
12	
13	Robblee, M. B., Barber, T. B., Carlson, P. R., Durako, Jr. M. J., Fourqurean, J. W.,
14	Muehlstein, L. M., Porter, D., Yabro, L. A., Zieman, R. T., & Zieman J. C. 1991.
15	Mass mortality of the tropical seagrass Thalassia testudinum in Florida Bay (USA).
16	Marine Ecology Progress Series 71, 297-299.
17	
18	Short, F. T., & Wyllie-Echeverria, S. 1996. Natural and human-induced disturbance
19	of seagrasses. Environmental Conservation 23, 17-27.
20	
21	Smith, N. 1994. Long-term Gulf-Atlantic transport through tidal channels in the
22	Florida Keys. Bulletin of Marine Science 54, 602-609
23	

1	Szmant, A. M., & Forrester. A. 1996. Water column and sediment nitrogen and
2	phosphorus distribution patterns in the Florida Keys, USA. Coral Reefs 15, 21-41.
3	
4	Tabb, D. C. & Roessler, M. A. 1989. History of studies on juvenile fishes of coastal
5	waters of Everglades National Park. Bulletin of Marine Science 44, 23-34.
6	
7	Taylor, W.R. 1960. Marine algae of the eastern tropical and subtropical coasts of
8	the Americas. University of Michigan Press. Ann Arbor. 870 pp.
9	
10	Tilmant, J. T. 1989. A history and overview of recent trends in the fisheries of
11	Florida Bay. Bulletin of Marine Science 44, 3-22.
12	
13	Tsai, C. C., Chang, J. S., Sheu F. Shyu, Y.T. Yu, A. Y., Wong, S.L., Dai, C.F. &
14	Lee, T.M. 2005. Seasonal growth dynamics of Laurencia papillosa and Gracilaria
15	coronopifolia from highly eutrophic reef in southern Taiwan: temperature limitation
16	and nutrient availability. J. Expe. Mar. Bio. Ecol 315: 49-69.
17	
18	United States Environmental Protection Agency. 1992. Water Quality Protection
19	Program for the Florida Keys National Marine Sanctuary, Phase II. Continental
20	Shelf Associates Inc. and Battelle Ocean Science, Contract No. 68-C8-0105.
21	Washington, D.C.
ഹ	

1	Valiela, I., Mcclelland, J., Hauxwell, J., Behr, P. J., Hersh, D. & Foreman K. 1997.
2	Macroalgal blooms in shallow estuaries: Controls and ecophysiological and
3	ecosystem consequences. Limnology and Oceanography 42, 1105–1118.
4	
5	Vroom, P. S., Smith, C. M., Coyer, J. A., Walters, L. J., Hunter, C. L., Beach, K S.,
6	and Smith, J. E. 2003. Field biology of Halimeda tuna (Bryopsidales, chlorophyta)
7	across a depth gradient: comparative growth, survivorship, recruitment, and
8	reproduction. <i>Hydrobiologia</i> 501, 149-166.
9	
10	Zieman, J. C., Fourqurean, J. W. and Iverson, R. L. 1989. Distribution, abundance
11	and productivity of seagrasses and macroalgae in Florida Bay. Bulletin of Marine

Sciences **44**, 292-311.

1 Table 1. Kendall T-b correlations of median values of nutrients and average abundance

2 and slope values of macroalgal groups. Bold numbers are statistical significant

3 correlations (p <0.05). Ab= Abundance Index, S= Slope.

4 5

	6		BA		BO		CB		BO	
	Ab	S	Ab	S	Ab	S	Ab	S	Ab	S
NO3	0.20	0.22	0.02	0.17	0.15	0.02	0.04	0.01	0.01	0.19
NOS	0.30	0.32	0.02	-0.17	0.15	0.02	0.04	0.01	0.01	-0.10
P	0.01	0.01	0.44	0.09	0.12	0.43	0.40	0.48	0.46	0.08
NO2	0.26	0.25	-0.07	-0.12	0.28	0.12	0.00	0.02	0.00	-0.01
Р	0.02	0.03	0.31	0.18	0.01	0.18	0.49	0.45	0.49	0.46
NH4	0.09	0.15	-0.04	-0.07	0.12	0.03	-0.08	-0.08	-0.04	-0.10
Р	0.24	0.12	0.39	0.29	0.17	0.42	0.27	0.27	0.39	0.23
TN	0.33	0.32	-0.10	0.00	0.23	0.08	-0.06	-0.16	-0.05	-0.07
р	0.00	0.01	0.24	0.49	0.04	0.28	0.32	0.11	0.35	0.28
TON	0.30	0.32	-0.11	-0.01	0.20	0.08	-0.07	-0.20	-0.07	-0.10
р	0.01	0.01	0.22	0.46	0.06	0.27	0.29	0.06	0.31	0.23
TP	0.08	0.14	0.04	-0.10	0.27	0.01	-0.01	-0.19	0.17	0.02
р	0.28	0.14	0.40	0.23	0.02	0.46	0.48	0.08	0.10	0.44
тос	0.30	0.23	-0.06	0.07	0.28	0.05	0.01	-0.13	0.05	-0.04
р	0.01	0.04	0.33	0.30	0.01	0.35	0.48	0.17	0.35	0.39
TN:TP	0.09	0.15	-0.19	-0.14	-0.07	0.15	-0.24	-0.14	-0.36	-0.19
р	0.23	0.12	0.09	0.14	0.30	0.12	0.04	0.15	0.00	0.07
N:P	0.03	0.13	-0.06	-0.09	0.08	0.30	-0.07	0.13	-0.12	0.12
р	0.41	0.15	0.34	0.25	0.27	0.01	0.29	0.16	0.18	0.18

1 Legend of Figures

- 2 Figure 1. Study Area
- 3 Figure 2. Maps displaying interpolated median values for nutrients. Y and X axes show
- 4 latitude and longitude coordinates. Color scale shows the median concentration of each
- 5 nutrient.
- 6 Figure 3. Maps displaying interpolated mean abundance for the macroalgal groups: CG
- 7 Calcareous green, GO Green other, RO Red Other, BA Batophora-Acetabularia, CR
- 8 Crustose Red and BO Brown Other. Y and X axes show latitude and longitude
- 9 coordinates. Color scales show the Braun-Blanquet abundance index.
- 10 Figure 4. Time series showing some examples of sites and algal groups model results:
- 11 Dots = observed data, solid line= non-linear regression curve.
- 12 Figure 5. Histogram showing slope/year values for each group in each site. CG
- 13 Calcareous green, GO Green other, RO Red Other, BA Batophora-Acetabularia, CR
- 14 Crustose Red and BO Brown Other.
- 15 Figure 6. Box-plots showing significant differences of CG *Ab_i*, and *m_i* as a function of
- 16 distance from shore category.
- 17 Figure 7. Box-plots showing significant differences of RO *Ab_i*, *m_i* as a function of segment
- 18 and alongshore categories.
- 19 Figure 8. Scatter-plots showing correlation between CG and nutrients.
- 20 Figure 9. Scatter-plots showing correlation between RO and nutrients.



Figure 1. Study area showing regions and study sites





Figure 2. Maps displaying interpolated median values for nutrients. Y and X axes show latitude and longitude coordinates. Color scale shows the median concentration of each nutrient.



0.15

-80.20

0.24 -0.06

-80.20



Figure 3. Maps displaying interpolated mean abundance for the macroalgal groups: CG Calcareous green, GO Green other, RO Red Other, BA Batophora-Acetabularia, CR Crustose Red and BO Brown Other. Y and X axes show latitude and longitude coordinates. Gray scale shows the Braun-Blanquet abundance index.



Figure 4. Time series showing some examples of sites and algal groups model results: Dots = observed data, solid line= non-linear regression curve.





5 6 7 8 Figure 5. Histogram showing slope/year values for each group in each site. CG Calcareous green, GO Green other, RO Red Other, BA Batophora-Acetabularia, CR Crustose Red and BO Brown Other





Figure 6. Box-plots showing CG *Ab_i*, and *m_i* as a function of distance to shore category.





Figure 7. Boxplot showing differences of RO Ab_i and m_i as a function of Segment and Alongshore categories.





Figure 8. Scatter-plots showing correlation between CG Ab_i and m_i and nutrients







Figure 9. Scatter-plots showing correlation between RO Ab_i and m_i and nutrients